USACE Contract No.: W912DQ-11-D-3004

US Army Corps of Engineers Kansas City District

Draft Quality Assurance Project Plan

Rolling Knolls Landfill Superfund Site
Data Gap Investigation Oversight
Chatham, New Jersey
Task Order No. 019

December 5, 2014





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December 5, 2014

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Attn: CENWK- PM-E/Amy Darpinian
Project: Contract No. W912DQ-11-D-3004

Task Order No. 019

Rolling Knolls Landfill Superfund Site

Chatham, New Jersey

Subject: Draft Quality Assurance Project Plan

Dear Amy:

CDM Federal Programs Corporation (CDM Smith) is pleased to submit an electronic copy of the Draft Quality Assurance Project Plan for Oversight of the Data Gap Investigation at the Rolling Knolls Landfill Superfund Site, located in Chatham, New Jersey.

If there are questions concerning this submittal, please contact me at (732) 590-4663.

Very truly yours,

CDM FEDERAL PROGRAMS CORPORATION

Hazerman

Paul Hagerman, P.E Project Manager

Enclosure

cc: Tanya Mitchell, EPA

Jeniffer Oxford, CDM Smith Field Oversight Staff, CDM Smith Kershu Tan, CDM Smith

file: 6424-019

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(Worksheet#15)

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Appendix C DESA Generic QAPP Worksheets

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Appendix E CDM Smith TSOP and Sampler Guides

This QAPP is prepared in accordance with the UFP-QAPP manual (EPA 2005) and is compliant with EPA's QAPP guidance document EPA QA/R-5 (EPA 2002). The project will be implemented in accordance with the quality procedures in CDM Smith's Quality Assurance (QA) Manual (CDM Smith 2012) and this governing QAPP.

All worksheets are included herein.



References

ARCADIS. 2014. Data Gaps Sampling and Analysis Plan, Rolling Knolls Landfill Superfund Site, Chatham, New Jersey. Prepared for the Rolling Knolls Landfill Settling Parties. November.

ARCADIS. 2014. Quality Assurance Project Plan for the Data Gaps Sampling and Analysis Plan, Rolling Knolls Landfill Superfund Site, Chatham, New Jersey. Prepared for the Rolling Knolls Landfill Settling Parties. November.

ARCADIS. 2012. Site Characterization Summary Report, Rolling Knolls Landfill Superfund Site, Chatham, New Jersey. Prepared for the Rolling Knolls Landfill Settling Parties.

Foster Wheeler Environmental Corporation. 2000. Expanded Site Inspection Report: Rolling Knolls Landfill, Green Village, Chatham Township, Morris County, New Jersey. Volume I of IV. Prepared for the USEPA.

NUS Corporation. 1986. Report of Soil Sampling and Drilling Program at the Green Village Disposal Site. Prepared for the USEPA, Region II Edison, New Jersey.

USFWS Fish and Wildlife Enhancement. 1991. Technical Assistance Report, Contaminants in Fish and Sediments of Great Swamp National Wildlife Refuge, Morris County, New Jersey. Results of 1988 Sampling Efforts. Prepared for the USFWS Refuges and Wildlife (region 5), Massachusetts.

Weston. 2003. CLP Analytical Data of Soil and Sediment Samples Rolling Knolls Landfill Green Village Chatham Township, Morris County, New Jersey. Prepared for the USEPA. (Weston, August 2003); PCB Field Screening Analytical Data of Soil and Sediment Samples Rolling Knolls Landfill Green Village Chatham, Morris County, New Jersey. Prepared for the USEPA. (Weston, April 2003); Sampling Trip Report – Rolling Knolls Landfill. Prepared for the USEPA. April.



Acronyms

ABS absolute difference

AES atomic emission spectrophotometry
ANSETS Analytical Services Tracking System
ASC analytical services coordinator
CCV continuing calibration verification
CDM Smith CDM Federal Programs Corporation
CHMM Certified Hazardous Materials Manager

CIH certified industrial hygienist
CLP contract laboratory program
COC contaminant of concern

CoC chain of custody

CQCP contractor quality control plan
CRQL contract required quantitation limit

CSM conceptual site model

CVAFS cold vapor atomic fluorescence spectrometry
DESA Division of Environmental Science and Assessment

DO dissolved oxygen
DPT Direct-Push Technology
DQI data quality indicator
DQO data quality objective
DV data validation

DMC deuterated monitoring compound

EDD electronic data deliverable

ELAP Environmental Laboratory Accreditation Program
EPA United States Environmental Protection Agency

eV electron volt

FAR Federal Acquisition Regulations

FASTAC Field and Analytical Services Teaming Advisory Committee

FCR field change request
FID flame ionization detector
FLPE fluorinated polyethylene

FOS field team leader

GC/MS gas chromatograph/mass spectroscopy

GWQS Groundwater Quality Standards

HASP Health and Safety Plan H&S health and safety HCl hydrochloric acid

HDPE high density polyethylene ICP inductively coupled plasma

ICP-AES inductively coupled plasma atomic emission spectroscopy ICP-MS inductively coupled plasma mass spectrophotometer

ID identification

IDW Investigation Derived Waste

IR infra-red

LCS laboratory control sample LOQ limit of quantitation

MCAWW Method for Chemical Analysis of Water and Wastes

MDL minimum detection limit MEE methane, ethane, ethane



mg/kg milligram per kilogram mg/L milligram per liter

mL milliliter

MPC measurement performance criteria

MW monitoring well

MS mass spectrophotometer
MSA Master Services Agreement

MS/ MSD matrix spike /matrix spike duplicate

NA not applicable N/A not available

NELAP National Environmental Laboratory Accreditation Program

NIST National Institute of Standards and Technology

NJAC New Jersey Administrative Code

NJDEP New Jersey Department of Environmental Protection
OSHA Occupational Safety and Health Administration

OSRTI Office of Superfund Remediation and Technology Innovation

OU operable unit %D percent difference %R percent recovery PAL project action limit PB preparation blank PM project manager POC point of contact ppb parts per billion ppm parts per million

PQL project quantitation limit
PQLG project quantitation limit goal
PQO project quality objective
PTFE polytetrafluoroethylene
QA quality assurance

QAS quality assurance specialist
QAPP quality assurance project plan

QC quality control
QL quantitation limit
QP Quality Procedure
RA remedial action

RAC Remedial Action Contract
RAO Remedial Action Objective
RAS routine analytical service

RCRA Resource Conservation and Recovery Act RI/FS remedial investigation/feasibility study

ROD record of decision

RPD relative percent difference RPM remedial project manager RRF relative response factor

RSCC Regional Sample Control Coordinator

RSD relative standard deviation SAP sampling and analysis plan SDG sample delivery group

SF square feet

SIM selected ion monitoring



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SOP standard operating procedure

SOW Statement of Work

SSHO site health and safety officer
SSHP site safety and health plan
SVOC semi-volatile organic compound

TAL target analyte list
TAT turnaround time
TBD to be determined
TCL target compound list

TSOP Technical Standard Operating Procedure

UFP Uniform Federal Policy

μg microgram

μg/kg microgram per kilogram μg/L microgram per liter

USGS United States Geologic Survey

USACE United States Army Corps of Engineers

VOC volatile organic compound VTSR verified time of sample receipt

°C degree Celsius



1.0 Introduction

Under the United States Army Corps of Engineers (USACE), Kansas City District, Contract No. W912DQ-11-D-3004, Task Order No. 0019, CDM Federal Programs Corporation (CDM Smith) was directed to perform oversight of the Data Gap Investigation for the Rolling Knolls Landfill Superfund Site located in Chatham, New Jersey.

This Quality Assurance Project Plan (QAPP) addresses oversight of Rolling Knolls Landfill Settling Parties (the Group's) activities performed in support of the Data Gap Investigation which involve sampling and analysis of soil, sediment, surface water, groundwater and porewater to further characterize and delineate the site contamination.

This QAPP has been prepared in accordance with the Uniform Federal Policy (UFP)-QAPP manual (EPA 2005) and optimized worksheets (EPA 2012) and is compliant with EPA's QAPP Requirements document EPA QA/R-5 (EPA 2001). This task order will be implemented in accordance with the quality procedures in CDM Smith's Quality Assurance (QA) Manual (CDM Smith 2012). This QAPP is the governing document for execution of this oversight. CDM Smith will use the various documents prepared by the Group's contractor to verify proper execution of the data gap investigation. The QAPP covers the oversight tasks currently assigned to CDM Smith.

1.1 Site Description

The Rolling Knolls Landfill is an approximately 200-acre, unlined, former municipal landfill located at 35 Britten Road in the Green Village section of Chatham Township. The facility is bounded by the Great Swamp National Wildlife Refuge to the east, south and west; Loantaka Brook and private property to the west; and private residential properties to the north and northwest. The southern and eastern portions of the landfill lie within the boundaries of the Refuge, which is a designated national wildlife refuge and has habitat known to be used by state- and federally designated or proposed endangered or threatened species. The facility is minimally accessible and is not covered by an impenetrable material.

The Rolling Knolls landfill operated as a municipal landfill from the early 1930s through December 1968. During that time, it received municipal solid waste, as well as construction and demolition debris, from surrounding municipalities. Chatham Township Board of Health records indicate that the types of wastes deposited at Rolling Knolls included tree stumps, scrap metal, tires, household refuse, residential septic wastes, and industrial waste. In order to comply with health code regulations adopted in 1959, operational procedures at the facility included the application of herbicides and pesticides to control weeds, insects, and rodents, as well as the application of oil on facility roadways to control dust and daily cover over all exposed surfaces.

Analytical results of surface and subsurface soil samples taken in May 1999 indicated elevated levels of metals, phthalates, and polychlorinated biphenyls (PCBs) at the site. Additional sampling conducted in March 2003 confirmed the presence of elevated levels of PCBs in both the site soil and wetland sediment, on both the privately and federally owned portions of the landfill. Elevated levels of mercury were also detected in the sediment in the southeast portion of the landfill. Additional sampling was needed to further define the nature and extent of contamination at the site.

Notice of liability letters were sent to several potentially responsible parties (the Group). An Administrative Settlement Agreement and Order on Consent (AOC) for Remedial Investigation and Feasibility Study (RI/FS) was reached on 30 September 2005 between U.S. Environmental Protection Agency (EPA) and the "Settling Parties". Approximately twenty percent of the site is owned by the U.S. Department of the Interior, Fish and Wildlife Service. The Settling Parties are listed as follows: Chevron Environmental Management Company, for itself and on behalf of Kewanee industries, Inc.; Lucent Technologies Inc.; and Novartis Pharmaceuticals Corporation as successor to Ciba-Geigy Corporation.



In 2007, on behalf of the Rolling Knolls Landfill Settling Parties (the Group), ARCADIS U.S., Inc., conducted an RI/FS investigation to determine the nature and extent of contamination in groundwater, soil, sediment, surface water and to characterize the chemical constituents of industrial waste, if any, or other waster material identified as potential source material present at the site. The investigation culminated in a Site Characterization Summary Report (SCSR 2012, February).

In March 2013, the USEPA identified several data gaps related to delineation of constituents in environmental media. To address these data gaps, ARCADIS will perform a data gap investigation to:

- Assess the data gaps identified by EPA in August 2014
- Further delineate the extent of the site constituents in soil, groundwater, surface water and sediment
- Characterize surface water and sediment in ponds that were not sampled during previous events
- Characterize the pore water chemistry downgradient of monitoring well MW-10
- Characterize the current groundwater constituent concentrations via monitoring well sampling events
- Investigate the connection between groundwater and surface water on site
- Assess the conditions at the existing Hunt Club well HC-1

1.2 Summary and Purpose of the QAPP

This QAPP serves to detail activities and procedures required to determine the accuracy of the Group's data for the Data Gap Investigation, and to verify that they perform the investigation study activities in accordance with their approved plans. Split samples will be accepted during the following activities:

Phase 1 Field Work

- Soil sampling
- Temporary monitoring well installation and sampling
- Surface water and sediment sampling
- Recovery of pore-water samplers and sample collection
- Redevelopment and sampling of existing wells

Phase 2 Field Work

- Second groundwater sampling event (all new monitoring wells, and selected existing monitoring wells based on the results of the first groundwater sampling event)
- Third groundwater sampling event (all new monitoring wells)

The Data Gap Investigation field activities will be performed by the Group. CDM Smith will perform oversight of the Data Gap Investigation Field Activities. Additionally, CDM Smith will collect split samples at a rate of 10% for analysis of all matrices for Target Compound List (TCL) volatile organic compounds (VOCs), semi- volatile organic compounds (SVOCs) plus selected ion monitoring (SIM), Pesticides, polychlorinated biphenyls (PCBs), target analyte list (TAL) Metals, Mercury and Cyanide and Trace Mercury, Dioxins/Furans and PCB Congeners; and submit a Data Evaluation Report.



QAPP Worksheets #1 and 2: Title and Approval Page (UFP-QAPP Manual Section 2.1) (EPA 2106-G-05 Section 2.2.1)

1.	Pro	Project Identifying Information a. Site name: Rolling Knolls Landfill Superfund Site						
	b.	Site location: Chatham, New Jersey						
	C.	Contract/Work assignment number: W912DQ-11-	D-3004/7	Task Order 0019				
2.	Lea	ad Organization:						
	Un	ited States Army Corps of Engineers (USACE), Kansas	City Dis	trict				
	a.	Project Manager: Amy Darpinian						
		Signature	Date					
3.	Fed	deral Regulatory Agency (name/title/signature/date	e)					
	Un	ited States Environmental Protection Agency (EPA) F	Region 2					
	a.	Remedial Project Manager: Tanya Mitchell						
		Signature	Date	3				
4.	Otl	her Stakeholders: CDM Smith						
	a.	CDM Smith Project QA Specialist: Jeniffer Oxford						
		Signature Inford for	_ Date	12/5/2014				
	b.	CDM Smith Project Manager: Paul Hagerman						
		Signature and Robert Representation	Date	12/5/2014				

5. List plans and reports from previous investigations relevant to this project

- a. Quality Assurance Project Plan for Data Gaps Sampling and Analysis Plan, Rolling Knolls Landfill Superfund Site. Arcadis, October 2014
- Data Gaps Sampling and Analysis Plan, Rolling Knolls Landfill Superfund Site. Arcadis, October 2014
- c. Foster Wheeler Environmental Corporation. 2000. Expanded Site Inspection Report: Rolling Knolls Landfill, Green Village, Chatham Township, Morris County, New Jersey. Volume I of IV. Prepared for the USEPA.
- d. NUS Corporation. 1986. Report of Soil Sampling and Drilling Program at the Green Village Disposal Site. Prepared for the USEPA, Region II Edison, New Jersey.
- e. USFWS Fish and Wildlife Enhancement. 1991. Technical Assistance Report, Contaminants in Fish and Sediments of Great Swamp National Wildlife Refuge, Morris County, New Jersey. Results of 1988 Sampling Efforts. Prepared for the USFWS Refuges and Wildlife (region 5), Massachusetts.
- f. Weston. 2003. CLP Analytical Data of Soil and Sediment Samples Rolling Knolls Landfill Green Village Chatham Township, Morris County, New Jersey. Prepared for the USEPA. (Weston, August 2003); PCB Field Screening Analytical Data of Soil and Sediment Samples Rolling Knolls Landfill Green Village Chatham, Morris County, New Jersey. Prepared for the USEPA. (Weston, April 2003); Sampling Trip Report Rolling Knolls Landfill. Prepared for the USEPA. April.



QAPP CROSSWALK Identifying Information

The following table provides a "cross-walk" between the QAPP elements outlined in the UFP-QAPP Manual, the necessary information, and the location of the information within the text document and corresponding QAPP Worksheet.

Optimized UFP-QAPP Worksheets		2106-G-05 QAPP Guidance Section		
1 & 2	Title and Approval Page	2.2.1	Title, Version, and Approval/Sign-Off	
3 & 5	Project Organization and QAPP	2.2.3	Distribution List	
	Distribution	2.2.4	Project Organization and Schedule	
4,7&8	Personnel Qualifications and Sign-off	2.2.1	Title, Version, and Approval/Sign-Off	
	Sheet	2.2.7	Special Training Requirements and Certification	
6	Communication Pathways	2.2.4	Project Organization and Schedule	
9	Project Planning Session Summary	2.2.5	Project Background, Overview, and Intended Use of Data	
10	Conceptual Site Model	2.2.5	Project Background, Overview, and Intended Use of Data	
11	Project/Data Quality Objectives	2.2.6	Data/Project Quality Objectives and Measurement Performance Criteria	
12	Measurement Performance Criteria	2.2.6	Data/Project Quality Objectives and Measurement Performance Criteria	
13	Secondary Data Uses and Limitations	Chapter 3	QAPP Elements for Evaluating Existing Data	
14 & 16	Project Tasks & Schedule	2.2.4	Project Organization and Schedule	
15	Project Action Limits and	2.2.6	Data/Project Quality Objectives and Measurement	
	Laboratory-Specific Detection / Quantitation Limits		Performance Criteria	
17	Sampling Design and Rationale	2.3.1	Sample Collection Procedure, Experimental Design, and Sampling Tasks	
18	Sampling Locations and Methods	2.3.1	Sample Collection Procedure , Experimental Design, and Sampling Tasks	
		2.3.2	Sampling Procedures and Requirements	
19 & 30	Sample Containers, Preservation, and Hold Times	2.3.2	Sampling Procedures and Requirements	
20	Field QC	2.3.5	Quality Control Requirements	
21	Field SOPs	2.3.2	Sampling Procedures and Requirements	
22	Field Equipment Calibration, Maintenance, Testing, and Inspection	2.3.6	Instrument/Equipment Testing, Calibration and Maintenance Requirements, Supplies and Consumables	
23	Analytical SOPs	2.3.4	Analytical Methods Requirements and Task Description	
24	Analytical Instrument Calibration	2.3.6	Instrument/Equipment Testing, Calibration and Maintenance Requirements, Supplies and Consumables	
25	Analytical Instrument and Equipment Maintenance, Testing, and Inspection	2.3.6	Instrument/Equipment Testing, Calibration and Maintenance Requirements, Supplies and Consumables	
26 & 27	Sample Handling, Custody, and Disposal	2.3.3	Sample Handling, Custody Procedures, and Documentation	



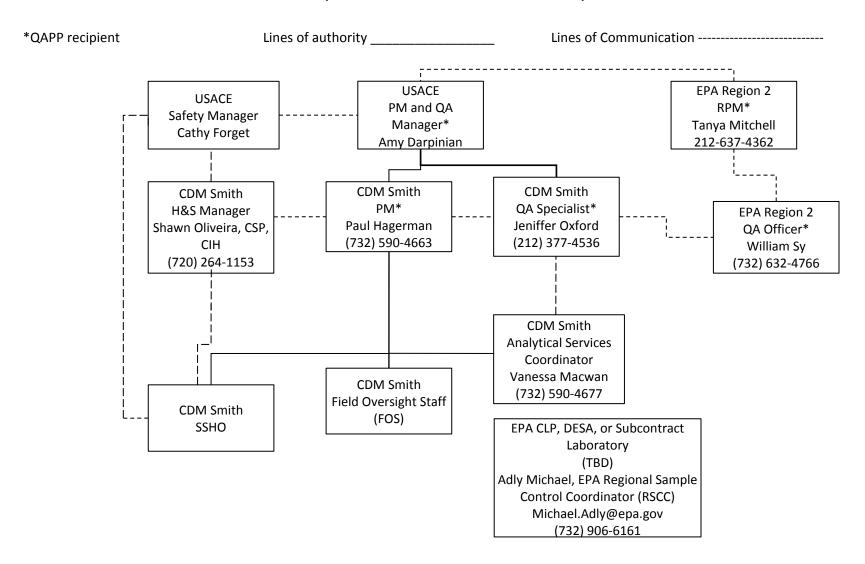
QAPP CROSSWALK Identifying Information

The following table provides a "cross-walk" between the QAPP elements outlined in the UFP-QAPP Manual, the necessary information, and the location of the information within the text document and corresponding QAPP Worksheet.

Optimize	Optimized UFP-QAPP Worksheets		2106-G-05 QAPP Guidance Section		
28	Analytical Quality Control and Corrective Action	2.3.5	Quality Control Requirements		
29	Project Documents and Records	2.2.8	Documentation and Records Requirements		
31, 32 &	Assessments and Corrective Action	2.4	Assessments and Data Review		
33		2.5.5	Reports to Management		
34	Data Verification and Validation Inputs	2.5.1	Data Verification and Validation Targets and Methods		
35	Data Verification Procedures	2.5.1	Data Verification and Validation Targets and Methods		
36	Data Validation Procedures	2.5.1	Data Verification and Validation Targets and Methods		
37	Data Usability Assessment	2.5.2	Quantitative and Qualitative Evaluations of Usability		
		2.5.3	Potential Limitations on Data Interpretation		
		2.5.4	Reconciliation with Project Requirements		



QAPP Worksheet #3 & 5: Project Organization and QAPP Distribution (UFP-QAPP Manual Section 2.3 and 2.4) (EPA 2106-G-05 Section 2.2.3 and 2.2.4)





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QAPP Worksheet #4, 7 & 8: Personnel Qualifications and Sign-off Sheet (UFP-QAPP Manual Sections 2.3.2 – 2.3.4) (EPA 2106-G-05 Section 2.2.1 and 2.2.7)

ORGANIZATION: CDM Smith

Name	Project Title/Role	Education /Experience	Specialized Training/Certifications	Signature/Date
Paul Hagerman	PM - Oversees project and responds to USACE PM and EPA RPM. Manages subcontractors. Responsible for implementing and maintaining QA program. Determine the need for any corrective action.	B.S. – Mechanical Engineering M.S. – Mechanical Engineering; over 20 years of project management and engineering experience	P.E.; Internal PM training modules;	
TBD	FOS - Oversee all field investigation activities		OSHA 40 hour training, annual 8 hour refresher, annual medical monitoring	



QAPP Worksheet #4, 7 & 8: Personnel Qualifications and Sign-off Sheet (UFP-QAPP Manual Sections 2.3.2 – 2.3.4) (EPA 2106-G-05 Section 2.2.1 and 2.2.7)

Name	Project Title/Role	Education /Experience	Specialized Training/Certifications ¹	Signature/Date ²
Shawn Oliveira	Health and Safety Manager - Oversees adherence to Health and Safety requirements	M.S., Environmental Engineering B.S., Chemistry	Certified Industrial Hygienist (CIH)	
Scott Kirchner	Project Chemist - Overall responsibility for laboratory services and data management and evaluating analytical data	B.S., Chemistry B.S, Environmental Science	СНММ	
Vanessa Macwan	ASC - Coordinates with EPA RSCC, Division of Environmental Science and Assessment (DESA) laboratory and subcontract laboratories	B.S., Environmental Science B.S., Engineering Technologies		
Jo Nell Mullins	Quality Assurance Manager - develops and implements the CDM Smith QA program and assesses the implementation of the quality requirements for all projects	M.S., Environmental Health B.S. – Biology/Chemistry 15 years of experience	ASQ Certified Quality Auditor; ISO 14001 Lead Auditor Certified; NQA-1 Lead Auditor Certified; OSHA 40 training and annual 8 hour refresher	
Jeniffer Oxford	QA Specialist - Oversees adherence to QA requirements	B.S., Natural Sciences	СНММ	
Christine Julias	Database Manager - Oversees data management; coordinates with data coordinators and validation staff	B.S., Chemical Engineering M.B.A., Marketing Management	P.E.	

QAPP Worksheet #4, 7 & 8: Personnel Qualifications and Sign-off Sheet (UFP-QAPP Manual Sections 2.3.2 – 2.3.4) (EPA 2106-G-05 Section 2.2.1 and 2.2.7)

ORGANIZATION: Laboratory

Name	Project Title/Role	Education/Experience	Specialized Training/Certifications	Signature/Date ²
EPA CLP Laboratory (TBD)	QA Officer	TBD (Experience vetted by accreditation body)	National Environmental Laboratory Accreditation Program (NELAP)/CLP	
DESA - Sumy Cherukara	QA Officer	TBD (Experience vetted by accreditation body)	NELAP/Trained in EPA and standard analytical methods	
CDM Smith subcontract Laboratory - TBD	QA Officer	TBD (Experience vetted by accreditation body)	NELAP and Environmental Laboratory Accreditation (ELAP)	

Notes:

- 1. CPR/First Aid- Red Cross or CINTAS- periodically as required (1-3 years).
- 2. Signatures indicate personnel have read and agree to implement this QAPP as written.
- 3. EPA Headquarters staff reviews and maintains the resumes of education and experience for key laboratory staff. This information is not available for the QAPP.



QAPP Worksheet #6: Communication Pathways (UFP-QAPP Manual Section 2.4.2) (EPA 2106-G-05 Section 2.2.4)

Communication Driver	Organization	Name	Contact Information	Procedure (Timing, Pathways, Documentation, etc.)
Regulatory agency interface	USACE Project Manager (PM)	Amy Darpinian	816-389-3897	The USACE PM will send all information about the project to the EPA RPM. Major changes will be discussed with the EPA PM prior to
	EPA RPM	Tanya Mitchell	212-637-4362	implementation.
Point of Contact with USACE	CDM Smith PM	Paul Hagerman	732-590-4663	All information about the project will be sent to USACE PM by CDM Smith PM.
Manage field tasks				Act as liaison to USACE PM and EPA RPM concerning investigation activities. Daily communication with project team and PM. Communicate implementation issues to FOS.
QAPP changes: prior to field work , in the field, and during project execution	CDM Smith FOS	TBD	TBD	Notify CDM Smith PM immediately and promptly complete a Field Change Request (FCR) form and/or corrected worksheets. Send FCR forms to Quality Assurance Specialist (QAS).
	CDM Smith PM	Paul Hagerman	732-590-4663	Notify EPA RPM, USACE PM and ASC of delays or changes to field work. Prepare QAPP Addendums or revisions in consultation with the client.
Field corrective actions	CDM Smith FOS	TBD	TBD	CDM Smith FOS will oversee implementation of corrective action and notify auditor, PM by email. CDM Smith PM will complete the corrective action report form.
Daily Quality Control Report (DCQR)				Complete on a daily basis and submit to CDM Smith PM and PE. CDM Smith PM will forward to USACE PM and EPA RPM upon request.
				Submit request to ASC before the timeframe below.
Booking of Analytical Services	CDM Smith ASC	Vanessa Macwan	732-590-4706	Coordinate DESA and Contract Laboratory Program (CLP) analytical services through Regional Sample Control Center (RSCC) 3 weeks prior to sampling.



QAPP Worksheet #6: Communication Pathways (UFP-QAPP Manual Section 2.4.2) (EPA 2106-G-05 Section 2.2.4)

Communication Driver	Organization	Name	Contact Information	Procedure (Timing, Pathways, Documentation, etc.)
Facilitate Database Setup and Data Management Planning	CDM Smith FOS	TBD	TBD	Provide sample and analytical information prior to sample collection to CDM Smith ASC and DC. Provide information on sample and analytical reporting groups, and types of report tables required for project.
Facilitate Data Management				Provide electronic survey data, sample ID, locations and analyses. Transmit completed sample tracking information to data manager by the completion of each sampling case.
Incomplete Electronic Data Deliverables (EDDs) or other EDD issues	CDM Smith Data Manager and Data Coordinator	Christine Julius	732-590-4610	Personnel identifying the issue will request resubmittal of corrected EDD by email.
Data verification issues, e.g., incomplete records	CDM Smith Data Coordinator	Tonya Bennett	212-377-4532	Data Coordinator will send an email to the FOS when an issue is found. FOS will address questions or any discrepancies.
Field Corrective Action	CDM Smith QAS, auditor, FOS	Jeniffer Oxford TBD	212-377-4536 TBD	PM, Task Manager, and FOS, per QA manual requirement corrective actions may also be identified by the field team. FOS initiates corrective action on identified field issues immediately or within QA manager (QAM) recommended timeframe.
Procurement of analytical services	CDM Smith FOS/ASC	TBD	TBD	FOS will prepare laboratory request; ASC will review and send email to RSCC. If needed they will prepare an analytical SOW and submit for project chemist review. FOS initiates laboratory kick-off call with subcontract laboratory (ies) and email agenda.
Analytical Services Support	CDM Smith ASC	Vanessa Macwan	732-590-4706	Act as liaison with RSCC for CLP laboratories, with John Birri for DESA, and with subcontract laboratory (ies).
Laboratory Quality Control Variances and Analytical Corrective Actions	Laboratory Project Manager or QC Officer	TBD	TBD	Communicate with the laboratory staff and regular communication with the CDM Smith ASC, QAS or designee. Provide oversight and direction on technical issues as needed.
Notification of Analytical Issues Sample receipt variances	CDM Smith ASC	Vanessa Macwan	732-590-4706	Notify FOS of any sample collection/shipment issues. Notify RSCC, DESA laboratory or subcontract laboratories to initiate corrective action.



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QAPP Worksheet #6: Communication Pathways (UFP-QAPP Manual Section 2.4.2) (EPA 2106-G-05 Section 2.2.4)

Communication Driver	Organization	Name	Contact Information	Procedure (Timing, Pathways, Documentation, etc.)
Data validation issues, e.g., Non-compliance with procedures; Data review corrective actions	CDM Smith data validator or data assessor	TBD	TBD	Submit a list of questions or issues to USACE and EPA or the subcontract laboratory as appropriate for correction or other appropriate response.
Reporting of Issues Relating to Analytical Data Quality	CDM Smith ASC	Vanessa Macwan	732-590-4706	Communicate to CDM Smith PM as appropriate.
(including ability to meet reporting limits, and usability of data)	CDM Smith Data Assessor	TBD	TBD	Communicate to CDM Smith PM as appropriate. Document situation and effect in a data quality report prepared prior to evaluation of remedial design report.
Release of Analytical Data	CDM Smith ASC	Vanessa Macwan	732-590-4706	Receive and review data packages before data is used. Coordinate validation if a subcontract laboratory is procured.
Site Health and Safety Issues	CDM Smith FOS	TBD	TBD	Make decisions regarding health and safety issues and upgrading PPE. Communicate to CDM smith PM and Health and Safety Manager, as appropriate.



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QAPP Worksheet #9: Project Planning Session Summary (UFP-QAPP Manual Section 2.5.1 and Figures 9-12) (EPA 2106-G-05 Section 2.2.5)

CDM Smith will accept ten percent of samples for analyses of the main contaminants of concern for all matrices collected during the Data Gap Investigation Oversight. Attachment 3 of this QAPP provides additional detail of the project planning session summary.



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QAPP Worksheet #10: Conceptual Site Model (UFP-QAPP Manual Section 2.5.2) (EPA 2106-G-05 Section 2.2.5)

Refer to the Group's QAPP (Appendix A) for information on the Conceptual Site Model (CSM). This Data Gap Investigation will further characterize the CSM; CDM Smith's oversight activities will facilitate verification of compliance with the Group's approved plans and accuracy of the data collected. Ten percent of the Group's samples including all matrices will be accepted and analyzed for definitive level TLC VOCs, SVOC +SIM, Pesticides, PCBs, TAL Metals, Mercury and Cyanide, Trace Mercury, Dioxins/Furans and PCB Congeners to meet the project action limits specified in the Group's document and shown on Appendix A of this QAPP. The split sample results will be compared with the Group's results using the measurement performance criteria described on Worksheet 37, section on Precision.



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QAPP Worksheet #11: Project Data Quality Objectives (UFP-QAPP Manual Section 2.6.1) (EPA 2106-G-05 Section 2.2.6)

The Group's QAPP addresses the Project data quality objective (DQOs). Split samples will be used to support the goals of the oversight program by generating definitive level data for comparison with the Group's data. The problem and framework for oversight are:

1. State the Problem – The field investigation is being led by the Group; USACE and EPA need to determine the accuracy of the Group generated data and to evaluate whether the field work is executed in compliance with approved documents. Oversight will include field observation and acceptance of split samples from the Group's to further delineate the extent of contaminants of concern in soil, groundwater, sediment and surface water.

CDM Smith will assist USACE/ EPA in the oversight of the field activities and will provide field oversight and analysis of split samples accepted from the Group's contractor to verify compliance with their approved project plans and the accuracy of their data. To evaluate the Group's data accuracy, CDM Smith will accept 10 percent split samples of all matrices for analysis at locations determined by coordination with the Group and in consultation with the USACE PM/ EPA RPM.

CDM Smith oversight of the Group's field investigation will include the following activities:

- Technical Review and evaluation of the Group's project plans and reports
- Documentation of field activities observations and deficiencies
- Review of the Group's-selected sampling locations
- Acceptance of split samples
- Sample handling, packaging and shipping to off-site laboratories
- Comparison of data sets to determine any analytical bias
- **2. Identify Study Goals** The goals are to verify, through independent oversight and split sampling analysis, that the Group's activities are in accordance with their Contractor's SAP, QAPP, and HASP and that the Group's data are representative of the site conditions and contaminant concentrations. Oversight and split sample data will be used to answer the environmental questions below:
 - Is the Group contractor complying with the approved plans or approved deviations?
 - Does the Group data adequately characterize the site, and are the data representative and useful for project decisions?
 - Are the Group and CDM Smith data complete and accurate?
 - Are data sets comparable as defined on Worksheet #37?
 - Does the data show any analytical bias?
 - Do the relative percent differences (RPDs) calculated for the Group and CDM Smith data fall within the measurement performance criteria?



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QAPP Worksheet #11: Project Data Quality Objectives (Oversight) (UFP-QAPP Manual Section 2.6.1) (EPA 2106-G-05 Section 2.2.6)

3. Identify Information inputs – The primary required data types will be analytical results from soil, sediment, groundwater, surface water and porewater samples collected from the selected locations.

CDM Smith, in consultation with the USACE PM/ EPA RPM, will determine sample locations to be split. CDM Smith will accept samples during the Group field program and send to a DESA, CLP or sub-contract laboratory to assess data accuracy. The analyses selected to be split are determined to be more critical for oversight evaluation. Chemical analyses will be obtained for the contaminants of concern; physical data will not be obtained unless determined critical to the investigation or as directed by USACE and EPA. The oversight will be used to verify data accuracy and whether the study questions listed in Step 2, Identify Study Goals, are adequately addressed.

4. Boundaries of the Study – CDM Smith will only be collecting split samples at a frequency of 10 percent (%) during Data Gap Investigation field activities. Samples locations are to be determined in consultation with the USACE PM and EPA RPM. Samples selected for split sampling data will cover a range of locations and concentrations, will cover critical items such as areas of potential contamination, and will be collected from each media type. The analyte group to be split is: *TLC VOCs, SVOC +SIM, Pesticides, PCBs, TAL Metals, Mercury and Cyanide, Trace Mercury, Dioxins/Furans and PCB Congeners*.

Sampling oversight will be performed according to the Group's schedule.

5. Analytical Approach – Analytical data and reports will be used to qualitatively assess any potential bias in the Group dataset. Sample results will be evaluated against the Group's project action limits on Worksheet #15 and against the Group's data using the split samples measurement performance criteria on worksheets #12 and 28 (**Appendix A**). Field implementation will be measured against procedures in the Group's field plans. The project decision criteria below will apply.



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QAPP Worksheet #11: Project Data Quality Objectives (Oversight) (UFP-QAPP Manual Section 2.6.1) (EPA 2106-G-05 Section 2.2.6)

Project Decision Conditions ("If..., then..." statements):

- If field work is inconsistent with the Group QAPP and SAP, then the field oversight staff will verify tasks with respect to the Group's QAPP, SAP, and HASP and note deviations with the Group's field project leader and document such discussions in the Weekly Oversight Summary Reports submitted to USACE and EPA. The CDM Smith PM, USACE PM and EPA RPM will be informed verbally of via email within 24 hours if there are deviations.
- If the Group team needs to relocate sampling locations, or there are any changes to the planned field program, CDM Smith will communicate this change to USACE and EPA and document it on the Daily Quality Control Reports.

CDM Smith will present data findings and submit it to USACE and EPA, who will then determine if any additional actions are required.

6. Performance and Acceptance Criteria -

- CDM Smith's QC data will be used to determine split sample data quality and whether sample results are acceptable based on the established project DQOs. Sample results will be compared to the measurement performance criteria (MPC) of the data quality indicators (DQIs).
- EPA's Field and Analytical Services Teaming Advisory Committee (FASTAC) policy for obtaining laboratory resources will be utilized for sampling events. Laboratory analysis will be performed through the DESA and CLP laboratories.
- Definitive level data is required for full validation of the data.
- The project-specific action limits and quantitation limits are specified on Worksheet #15 (**Attachment 1**) for all contaminants of concern. Analytical data generated will be compared to these limits. Data must meet the DQOs that have been specified for the site. Refer to Worksheets #12, 15 and 28.
- Laboratory reporting limits (contract required quantitation limits (CRQLs)) need to be below or equal to the Group's project action limits (PALs).
- In addition, to ensure that measurement performance criteria for usability (criteria for measures of precision, accuracy, representativeness, comparability, completeness, and sensitivity) are met, all data will be subject to validation and the outputs used to perform a data usability assessment.



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QAPP Worksheet #11: Project Data Quality Objectives (Oversight) (UFP-QAPP Manual Section 2.6.1) (EPA 2106-G-05 Section 2.2.6)

7. Detailed Plan for Obtaining Data -

Field sampling and field procedures are described in the Group's QAPP and SAP (Arcadis, November 2014). Sampling oversight will be performed according to the Group's schedule, unless unanticipated delays occur. See the Group's Figures in Attachment 2 and SAP (Appendix B) for potential split sample locations.

The Group Contractor's representative will collect and fill the sample containers and CDM Smith's field personnel will prepare the split samples for shipment. CDM Smith will perform sample management, prepare, package, and ship the split samples to the assigned laboratory. DESA, CLP or subcontract laboratory will generate the data. EPA's RSCC will communicate laboratory assignments to CDM Smith.

CDM Smith field personnel will observe the implementation of field and sampling activities and note any deviations from their Work Plan and QAPP. Deviations will be brought to the attention of the Group's contractor, and reported to the CDM Smith PM who will communicate this information to the USACE PM and EPA RPM. These will be documented in the daily communications and in the CDM Smith data comparability report. The data report will include a discussion of the impact of the deviation(s) on the data quality. CDM Smith will field oversight staff will document the Group contractor's activities will be documented in the field logbook.

Data Reporting

- CDM Smith will prepare a field oversight report for each split sampling event on a monthly basis. Sampling method, number of split samples collected, and documented compliance with the Group's sampling activities will be recorded along with the title of approved plans, USACE and EPA requirements.
- Sampling data results will be emailed to CDM Smith from DESA or the EPA for evaluation and data comparison. Final validated data will be submitted to CDM Smith in electronic format and/or hard copies.
- Following completion of laboratory analyses and receipt of all electronic and hardcopy data, CDM Smith will prepare a report and submit it to EPA and USACE. The report will include tabulated results and a discussion of the data quality and its comparability with the Group's data. This review will be used to evaluate the accuracy of the Group's data.

Data archiving

- Data will be downloaded from the EPA website or emailed to CDM Smith.
- Final CLP validated data will be submitted to CDM Smith in electronic format consistent with CLP deliverables
- Electronic data will be input into the project's EQuIS database.



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QAPP Worksheet #11: Project Data Quality Objectives (Oversight) (UFP-QAPP Manual Section 2.6.1) (EPA 2106-G-05 Section 2.2.6)

- EPA will archive CLP laboratory raw data in its document control system.
- Hard copies of field data including field logs will be archived in the project files.
- Hard copies of analytical data received by CDM Smith will be archived in the project files for 10 years after contract expiration.



QAPP Worksheet #12a: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical GroupTCL VOCs/SOM01.2Concentration LevelTrace or Low (μg/L)

DQIs QC Sample or Measurement Performance Activity		Measurement Performance Criteria
Overall Precision	Split Samples	≤50% RPD when VOCs in both samples ≥ CRQL ABS ≤ 5xQL when one or both results < CRQL
Overall Accuracy/bias (contamination)	Trip Blank*	No analyte > CRQL No target analyte concentrations ≥ CRQL
Precision	Matrix spike(MS)/Matrix spike duplicate (MSD)**	See Worksheet #28 for compound specific values
Accuracy	***Deuterated Monitoring Compounds (DMC); MS/MSD**	See Worksheet #28 for list of compound specific values and range of acceptable %Rs
Sensitivity	Method Blank	Results ≤ CRQL
Completeness	Data Assessment Also See Worksheet #34	≥90% Valid Data versus Total Data Collected and ≥90% Planned Data versus Data Collected Also See Worksheet #34

^{*}Reference EPA Region 2 SOP No. 34 for Trace/ Low VOA - Blank Type Criteria Table



^{**}Optional MS/MSD – Reference CLP SOM01.2, Exhibit D, Table 6 for Criteria – Not typically required for CLP in Region 2

^{***}DMCs - Reference CLP SOM01.2, Exhibit D, Table 5 for Criteria

QAPP Worksheet #12b: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical GroupTCL SVOCs/SOM01.2Concentration LevelLow/Medium (μg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Overall Precision	Split Samples	≤50% RPD when SVOCs in both samples ≥ CRQL otherwise ABS ≤ 5xCRQL
Precision	MS/MSD**	See Worksheet #28 for compound specific values
Accuracy	***DMCs; MS/MSD**	See Worksheet #28 for list of compound specific values and range of acceptable %Rs
Consitivity	LOQ verification or Method Blank	Results ≤ CRQL
Sensitivity	Data Assessment	CRQLs meet project quantitation limit goals (PQLGs)
Completeness	Data Assessment Also See Worksheet #34	≥90% Valid Data versus Total Data Collected and ≥90% Planned Data versus Data Collected Also See Worksheet #34

^{*}Reference EPA Region 2 /Low/Medium Semivolatile SOP shown on Worksheet # 36 or most recent revision http://www.epa.gov/region2/ga/documents.htm



^{**}Optional MS/MSD – Reference CLP SOM01.2, Exhibit D, Table 6 for Criteria – Not typically required for Region 2

^{***(}DMCs) - Reference CLP SOM01.2, Exhibit D, Table 5 for Criteria

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QAPP Worksheet #12c: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group Dioxin/Furans/ EPA 1613B

Concentration Level Low (µg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Split Samples	RPD ≤ 40% if concentration ≥5 QL
Precision	Laboratory duplicate	±20% of mean if concentration >10QL
Accuracy/Bias Precision	Ongoing precision and recovery	RPD ≤ 40%
Accuracy/ Representativeness	Temperature Blank checks/ DV	0 to 6 °C 10 °C (DV)
Precision	Initial precision and recovery standard	Per laboratory SOP
Accuracy/Bias		Various % recovery per laboratory SOP
Accuracy/Bias	Ongoing precision and recovery standard (OPR)	70-130 %recovery, RPD ≤ 40%
Accuracy/Bias	Surrogate standards	17-130% recovery
Comparability	Evaluated during DQA	Comparable units, and methods
Completeness	Evaluated during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)



QAPP Worksheet #12d: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Overall Precision	Split Samples	50% RPD when pesticides in both samples ≥ CRQL ABS ≤ 5xQL when one or both results < CRQL
Precision	MS/MSD**	See Worksheet #28 for compound specific values
Accuracy	***LCS; MS/MSD**	See Worksheet #28 for list of compound specific values
Concibinity	Method Blank	Results ≤ CRQL
Sensitivity	Data Assessment	CRQLs meet PQLGs
Completeness	Data Assessment Also See Worksheet #34	≥90% Valid Data versus Total Data Collected and ≥90% Planned Data versus Data Collected Also See Worksheet #34

^{*}Reference EPA Region 2 Low/Medium Pesticide Data Validation SOP shown on Worksheet # 36 or most recent revision http://www.epa.gov/region2/qa/documents.htm



^{**}MS/MSD – Reference CLP SOM01.2, Exhibit D, Table 3 for Criteria

^{***}LCS - Reference CLP SOM01.2, Exhibit D, Table 2 for Criteria

QAPP Worksheet #12e: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group TCL PCBs/SOM01.2
Concentration Level Low/Medium (µg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Overall Precision	Split Samples	≤50% RPD when PCBs in both samples ≥ CRQL ABS ≤ 5xQL when one or both results < CRQL
Analytical accuracy/bias (contamination)	Method Blank	No analyte > CRQL No target analyte concentrations ≥ ½ CRQL
Precision	MS/MSD**	See Worksheet #28 for compound specific values
Accuracy	***LCS; MS/MSD** Surrogates	See Worksheet #28 for the list of compound specific values
Sensitivity	Method Blank	Results ≤ CRQL
	Data Assessment	CRQLs meet PQLGs
Completeness	Data Assessment Also See Worksheet #34	≥90% Valid Data versus Total Data Collected and ≥90% Planned Data versus Data Collected Also See Worksheet #34

^{*}Reference EPA Region Low/Medium Aroclor Data Validation SOP on Worksheet # 36 or most recent revision http://www.epa.gov/region2/qa/documents.htm



^{**}MS/MSD - Reference CLP SOM01.2, Exhibit D, Table 1 for Criteria - Not typically required for Region 2

^{***}LCS - Reference CLP SOM01.2, Exhibit D, Table 2 for Criteria

QAPP Worksheet #12f: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group PCB Congeners/EPA 1668A

Concentration Level Low (µg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Split Samples	RPD ≤ 40% if concentration ≥5 QL otherwise ABS ≤QL
Precision	Laboratory duplicate	±20% of mean if concentration >10 xQL
Accuracy/Bias	Calibration Verification Sample	Per laboratory or method SOP (70-130% of native analytes and 50-150% for surrogates)
Accuracy/Bias Precision	Initial Precision and Recovery	60-140 %recovery RSD ≤ 40%
Accuracy/Bias	LCS or OPR	Per laboratory SOP Warning 70-130%R; Accept 50-150 %recovery
Accuracy/ Representativeness	Temperature Blank checks DV	0 to 6 °C 10 °C (DV)
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)



QAPP Worksheet #12g: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group TAL Metals /ISM01.3 Concentration Level ICP-AES (µg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Split Samples *	≤50% RPD¹ when both results ≥ CRQL otherwise, ABS ≤ 5xCRQL The validation SOP requires qualification of results ≤20% RPD. For the purpose of data use the 50% RPD criterion is satisfactory.
Precision	Laboratory Duplicate Sample **	<20% RPD**
Accuracy	Matrix Spike ***; LCSW ****	75–125% R; 70–130% R (except Ag and Sb)
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Field rinsate/ Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 ICP-AES Data Validation SOP or most recent revision http://www.epa.gov/region2/qa/documents.htm (includes absolute difference criteria)



^{**}Reference EPA CLP ISM01.3, Exhibit D of inductively coupled plasma (ICP) atomic emission spectrophotometry (AES) for Duplicate Sample Criteria (page D-22) (include absolute difference criteria)

^{***}Reference EPA CLP ISM01.3, Exhibit D of ICP-AES for Spike Sample Criteria (page D-21)

^{****}Reference EPA CLP ISM01.3, Exhibit D of ICP-AES for aqueous LCS (LCSW) Criteria (page D-23) w/exception of silver (Ag) and antimony (Sb)

QAPP Worksheet #12h: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group TAL –Total Mercury/ISM01.3 - Cold Vapor Atomic Absorption (CVAA)

Concentration Level Low (µg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
		≤50% RPD ABS ≤ 5xCRQL when either result ≤ CRQL
Precision	Split Samples	The validation SOP requires qualification of results ≤20% RPD. For the purpose of data use the 50% RPD criterion is satisfactory.
Precision	Laboratory Duplicate Sample **	≤20% RPD*
Accuracy	Matrix Spike***	75–125 %R
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*} Reference EPA Region Hg & CN Data Validation SOP or most recent revision http://www.epa.gov/region2/qa/documents.htm (include absolute difference criteria)



^{**}Reference EPA CLP ISM01.3, Exhibit D of Mercury for Duplicate Sample Analysis, (page D-19) (includes absolute difference criteria)

^{***}Reference EPA CLP ISM01.3, Exhibit D of Mercury for Spike Sample Analysis, (page D-18)

QAPP Worksheet #12i: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group TAL Metals /ISM01.3 Concentration Level ICP-MS (µg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
		≤50% RPD¹ when both results ≥ 5*CRQL ABS ≤ CRQL when either result ≤ 5*CRQL
Precision	Split Samples*	(¹Important Note. The validation SOP requires qualification of results ≤20% RPD. For the purpose of data use the 50% RPD criterion is satisfactory)
Precision	Laboratory Duplicate Sample **	<20% RPD**
Accuracy	Matrix Spike ***; LCSW ****	75–125% R; 80–120% R (except Ag and Sb)
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 ICP-MS Data Validation SOP or most recent revision http://www.epa.gov/region2/qa/documents.htm (include absolute difference criteria)



^{**}Reference EPA CLP ISM01.3, Exhibit D of (ICP) atomic emission spectrophotometry (AES) for Duplicate Sample Analysis (page D-25) (includes absolute difference criteria)

^{***}Reference EPA CLP ISM01.3, Exhibit D of ICP-MS for Spike Sample Analysis (page D-24)

^{****}Reference EPA CLP ISM01.3, Exhibit D of ICP-MS for aqueous LCS (LCSW) Analysis (page D-26)

QAPP Worksheet #12j: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group Mercury (trace)/EPA 1631
Concentration Level Trace (nanogram per liter (ng/L))

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Split Samples *	RPD ≤ 40% if concentration ≥5 CRQL or ABS≤ QL
Accuracy	Laboratory duplicate	RPD ≤ 25% for values ≥10 MDL. No more than 35% of RSDs >25%
Accuracy/Bias	MS/MSD	70-130 %R
Precision	MS/MSD; Initial Precision and Recovery (IPR)	MS/MSD - Laboratory SOP or RPD ≤ 35%; 70-130% R IPR - RSDs <20%; 75-125% R
Accuracy	OPR	Laboratory SOP or 70-130%R
Accuracy/ Representativeness	Temperature Blank checks DV	0 to 6 °C 10 °C (DV)
Comparability	Evaluated during Data Quality Assessment (DQA)	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)



QAPP Worksheet #12k: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Aqueous

Analytical Group TAL –Total Cyanide/ISM01.3 - Colorimeter or Spectrophotometer

Concentration Level Low (µg/L)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
		≤50% RPD ABS ≤ 5xCRQL when either result ≤ CRQL
Precision	Split Samples *	(¹Important Note. The validation SOP requires qualification of results ≤20% RPD. For the purpose of data use the 50% RPD criterion is satisfactory)
Precision	Laboratory Duplicate Sample **	<20% RPD*
Accuracy	Matrix Spike ***	75–125 %R
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*} Reference EPA Region Hg & CN Data Validation SOP or most recent revision http://www.epa.gov/region2/qa/documents.htm (include absolute difference criteria)



^{**}Reference EPA CLP ISM01.3, Exhibit D of Cyanide for Duplicate Sample Analysis, (page D-20) (includes absolute difference criteria)

^{***}Reference EPA CLP ISM01.3, Exhibit D of Cyanide for Spike Sample Analysis, (page D-19)

QAPP Worksheet #12I: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

MatrixSoil/SedimentAnalytical GroupTCL VOCs/SOM01.2Concentration LevelLow/Medium (μg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Split Samples	≤100% RPD ABS ≤ 5xQL when either result < 2X CRQL
Precision	MS/MSD**	%RPD – see worksheet #28
Accuracy	***DMCs; MS/MSD**	Compound specific %Rs are on worksheet #28
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 Low/ Medium VOCs Data Validation SOP shown on Worksheet # 36 or most recent revisionhttp://www.epa.gov/region2/qa/documents.htm



^{**}Optional MS/MSD – Reference CLP SOM01.2, Exhibit D, Table 6 for Criteria – Not typically required for Region 2

^{***(}DMCs) – Reference CLP SOM01.2, Exhibit D, Table 5 for Criteria

QAPP Worksheet #12m: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

MatrixSoil/SedimentAnalytical GroupTCL SVOCs/SOM01.2Concentration LevelLow/Medium (μg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Split Samples	≤100% RPD ABS ≤ 5xCRQL when either results ≤ 2*CRQL
Precision	Laboratory Duplicate; MS/MSD**	Worksheet #28 lists compound specific RPDs
Accuracy	***DMCs; MS/MSD**	Worksheet #28 lists compound specific %Recoveries
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 Low/Medium SVOCs Data Validation SOP shown on Worksheet # 36 or most recent revision http://www.epa.gov/region2/qa/documents.htm



^{**}Optional MS/MSD – Reference CLP SOM01.2, Exhibit D, Table 6 for Criteria – Not typically required for Region 2

^{***(}DMCs) - Reference CLP SOM01.2, Exhibit D, Table 5 for Criteria

QAPP Worksheet #12n: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Soil/Sediment

Analytical Group PCDD/PCDF/EPA 1613B

Concentration Level Low (µg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Split Samples	RPD ≤ 40% if concentration ≥5 CRQL
Precision	Laboratory duplicate	±20% of mean if concentration >10DL
Accuracy/Bias	LCS; MS/MSD	70-130 %recovery or per laboratory SOP
Precision	MS/MSD	RPD ≤ 20% if >10 QL
Accuracy/ Representativeness	Temperature Blank checks DV	0 to 6°C 10 °C (DV)
Precision	Initial precision and recovery	15-50% RSD or per laboratory SOP
Accuracy/Bias	Initial precision and recovery	Various % recovery per laboratory SOP
Accuracy/Bias	Ongoing precision and recovery	15-50% RSD or per laboratory SOP
Accuracy/Bias	Surrogate standards	17-130% R
Comparability	Evaluated during Data Quality Assessment	Comparable units, and methods
Completeness	Evaluated during Data Quality Assessment	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks/DV and DQA	≤ QLs (WS#15)

Laboratory and SOPs TBD. Laboratory is assigned per FASTAC policy.



QAPP Worksheet #12o: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Soil/Sediment

Analytical Group TCL Pesticides/SOM01.2
Concentration Level Low/Medium (µg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Sample Splits	≤100% RPD ABS ≤ 5xCRQL when either results ≤ 2X CRQL
Precision	Laboratory Duplicate; MS/MSD**	See list of compound specific RPDs on Worksheet #28
Accuracy	***LCS; MS/MSD**	See list of compound specific %Rs on Worksheet #28
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 Low/Medium Pesticide Data Validation SOP shown on Worksheet # 36 or most recent revision http://www.epa.gov/region2/qa/documents.htm



^{**}MS/MSD - Reference CLP SOM01.2, Exhibit D, Table 3 for Criteria - Not typically required for Region 2

^{***}LCS - Reference CLP SOM01.2, Exhibit D, Table 2 for Criteria

QAPP Worksheet #12p: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

MatrixSoil/SedimentAnalytical GroupTCL PCBs/SOM01.2Concentration LevelLow/Medium (μg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Sample Splits	≤100% RPD ABS ≤ 5xCRQL when either result is ≤2X CRQL
Precision	MS/MSD**	
	LCS***	See list of compound specific RPDs and %Rs on
Accuracy	MS/MSD**	Worksheet #28
	Surrogates	
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/	Method blanks	≤ QLs (WS#15 and laboratory SOP)
accuracy	assessed during DV and DQA	> CL2 (NA2#12 glin landiatoly 20h)

^{*}Reference EPA Region 2 Low/Medium Pesticide Data Validation SOP shown on Worksheet # 36 or most recent revision http://www.epa.gov/region2/qa/documents.htm



^{**}MS/MSD – Reference CLP SOM01.2, Exhibit D, Table 3 for Criteria – Not typically required for Region 2

^{***}LCS - Reference CLP SOM01.2, Exhibit D, Table 2 for Criteria

QAPP Worksheet #12q: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Soil/Sediment

Analytical Group PCB Congeners/EPA 1668A

Concentration Level Low (µg/kg)

DQIs	QC Sample and/or Activity Used to Assess Measurement Performance	Measurement Performance Criteria
Precision	Sample Splits	RPD ≤ 40% if concentration ≥5 CRQL
Precision	Laboratory duplicate	≤20% RPD; ±QL for samples <10x QL
Accuracy/Bias	Certified Reference Material; Calibration Verification Sample (QC Sample)	70 -130 %R
Accuracy/Bias	Initial Precision and Recovery	60-140 %R
Precision	Initial Precision and Recovery	RSD ≤ 40%
Accuracy/Bias	LCS or Ongoing Precision and Recovery	Per laboratory SOP Warning 70-130%R; Accept 50-150 %R
Accuracy/ Representativeness	Temperature Blank checks Data validation (DV)	0 to 6 °C 10 °C (DV)
Comparability	Data Quality assessment	Comparable units, and methods
Completeness	Data Quality Assessment	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks/ DV and DQA	≤ QLs (WS#15)



QAPP Worksheet #12r: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

MatrixSoil/SedimentAnalytical GroupTAL Metals/ISM01.3Concentration LevelICP-AES (mg/kg)

16. 7.25 (116/16)		
DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
Precision	Sample Splits *	≤100% RPD when both results ≥ 5*CRQL ABS ≤ 5xCRQL when either result ≤ 2xCRQL (¹Important Note. The validation SOP requires qualification of results ≤50% RPD. For the purpose of data use the 100% RPD criterion is satisfactory for most projects)
Precision	Laboratory Duplicate Sample **	\leq 35% RPD* (DV action based on this value)
Accuracy	Matrix Spike***; LCS****	75–125%R 70–130%R
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 ICP-AES Data Validation SOP or most recent revision http://www.epa.gov/region2/qa/documents.htm (include absolute difference criteria)



^{**}Reference EPA CLP ISM01.3, Exhibit D of ICP-AES for Duplicate Sample Analysis, (page D-22) includes absolute difference criteria

^{***}Reference EPA CLP ISM01.3, Exhibit D of ICP-AES for Spike Sample Analysis (page D-21)

^{****}Reference EPA CLP ISM01.3, Exhibit D of ICP-AES for LCS Sample Criteria (page D-23) with the exception of Ag and Sb

QAPP Worksheet #12s: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Soil/Sediment
Analytical Group TAL Metals/ISM01.3
Concentration Level ICP-MS (mg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
		≤100% RPD when both results ≥ 5*CRQL ABS ≤ 5xCRQL when either result ≤ 2x CRQL
Precision	Sample Splits *	(¹Important Note. The validation SOP requires qualification of results ≤50% RPD. For the purpose of data use the 100% RPD criterion is satisfactory)
Precision	Laboratory Duplicate Sample **	≤ 35% RPD* (DV action based on this value)
Accuracy	Matrix Spike***; LCS****	75–125%R 70–130%R
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 ICP-MS Data Validation SOP or most recent revision http://www.epa.gov/region2/qa/documents.htm (include absolute difference criteria)



^{**}Reference EPA CLP ISM01.3, Exhibit D of ICP-MS for Duplicate Sample Analysis, (page D-25) includes absolute difference criteria

^{***}Reference EPA CLP ISM01.3, Exhibit D of ICP-MS for Spike Sample Analysis (page D-24)

^{****}Reference EPA CLP ISM01.3, Exhibit D of ICP-MS for LCS Sample Criteria (page D-26)

QAPP Worksheet #12t: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Soil/Sediment

Analytical Group TAL –Total Mercury/ISM01.3 or current method-Cold Vapor Atomic Absorption (CVAA)

Concentration Level Low (mg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria
		≤100% RPD when both results ≥ 5*CRQL ABS ≤ 5xCRQL when either result ≤2*CRQL
Precision	Sample Splits *	(¹Important Note. The validation SOP requires qualification of results ≤50% RPD. For the purpose of data use the 100% RPD criterion is satisfactory)
Precision	Laboratory Duplicate Sample **	≤ 35% RPD* (DV action based on this value)
Accuracy	Matrix Spike***	75–125% recovery
Comparability	Assessed during DQA	Comparable units, and methods
Completeness	Assessed during DQA	≥ 90% collection and analysis
Sensitivity/ accuracy	Field rinsate/ Method blanks assessed during DV and DQA	≤ QLs (WS#15 and laboratory SOP)

^{*}Reference EPA Region 2 Hg &CN Data Validation SOP or most recent revision http://www.epa.gov/region2/qa/documents.htm (include absolute difference criteria)



^{**}Reference EPA CLP ISM01.3, Exhibit D of Mercury for Duplicate Sample Analysis (page D-19) (include absolute difference criteria)

^{***}Reference EPA CLP ISM01.3, Exhibit D of Mercury for Spike Sample Analysis (page D-18)

QAPP Worksheet #12u: Measurement Performance Criteria (UFP-QAPP Manual Section 2.6.2) (EPA 2106-G-05 Section 2.2.6)

Matrix Soil/Sediment

Analytical Group Total Cyanide /ISM01.3

Concentration Level Low (mg/kg)

DQIs	QC Sample or Measurement Performance Activity	Measurement Performance Criteria		
Precision	Split Samples	RPD ≤ 100% if concentration ≥5 CRQL otherwise ABS ≤2*CRQL		
Accuracy	Laboratory duplicate	≤35% RPD if Results >5xCRQL		
Accuracy/Bias	MS/MSD	75-125%R		
Duratelan	MS/MSD;	Laboratory SOP or RPD ≤ 35%;		
Precision	LCS	Method: RSDs <20%		
	MS/MSD;	Laboratory SOP or 75-125%		
Accuracy	LCS	70-130%R		
Accuracy/ Representativeness	Temperature Blank checks DV	0 to 6 °C and 10 °C (DV)		
Comparability	Evaluated during Data Quality Assessment	Comparable units, and methods		
0 11	5 1 1 1 1 2 2 2 2 2 2 2	≥ 90% Collection and		
Completeness	Evaluated during Data Quality Assessment	≥ 90% Valid data		
Sensitivity/	Method blanks/DV and DQA	≤ QLs (WS#15)		
accuracy	Wethou bidins/DV dilu DQA	≥ QL3 (44.2±1.2)		



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QAPP Worksheet #14 &16: Project Tasks & Schedule (UFP-QAPP Manual Section 2.8.2) (EPA 2106-G-05 Section 2.2.4)

The Group's Proposed Schedule

Phase 1 Field Work

Submittal of permit equivalencies - November 18, 2014

Soil sample collection - November 20 through December 5, 2014

Redevelopment of existing wells - November 24 through November 26, 2014

Temporary monitoring well installation and sampling – December 1 through December 5, 2014

Installation of pore-water samplers – December 2, 2014

Surface water and sediment sampling - December 2 through December 11, 2014

Assessment of Hunt Club well HC-1 - December 5, 2014

Sampling of existing wells - December 8 through December 12, 2014

Recovery of pore-water samplers and sample collection – December 16, 2014

Sample analysis – November 18, 2014 through January 19, 2015

Data Review and Document Preparation

Data validation – January 13, 2015 through February 12, 2015

Preparation of Interim Technical Memorandum - December 12, 2014 through February 12, 2015

Submit Interim Technical Memorandum to USEPA – February 13, 2015

USEPA review of Interim Technical Memorandum – February 16 through March 6, 2015

Receive USEPA approval of proposed permanent well locations – March 6, 2015

Phase 2 Field Work

Obtain well permits – March 9 through March 27, 2015

Install and develop proposed permanent wells – March 30 through April 10, 2015

Second groundwater sampling event (all new monitoring wells, and selected existing monitoring wells based on the results of the first groundwater sampling event) – April 27 through May 7, 2015

Third groundwater sampling event (all new monitoring wells) – July 27 through August 7, 2015

Groundwater sample analysis – April 28 through September 7, 2015

Evaluation of connection between northern ponds and groundwater – April 10 through August 14, 2015

Final Reporting

Data validation – June 7 through September 21, 2015

Begin preparation of final report - September 22, 2015

Submit final report to USEPA - October 30, 2015



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QAPP WORKSHEET # 15a Project Action Limits and Laboratory-Specific Detection/Quantitation Limits (UFP-QAPP Manual Section 2.6.2.3 and Figure 15) (EPA 2106-G-05 Section 2.2.6)

See Attachment 1



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QAPP Worksheet # 17a - Sampling Design and Rationale Oversight Split Sampling (UFP-QAPP Manual Section 3.1.1) (EPA 2106-G-05 Section 2.3.1)

Procurement of Technical Services

CDM Smith will procure an analytical laboratory for PCB Congeners, Dioxins/Furans and Trace Mercury in accordance with the Federal Acquisition Regulation and CDM Smith procedures. A scope of work will be prepared and will include the project's technical and quality requirements to meet the requirements established herein.

Field Planning Meetings

Prior to field activities, each field team member will review all CDM Smith project plans QAPPs, Health and Safety Plans (HASPs), etc., the Group's project plans, and participate in a field planning meeting conducted by the CDM Smith PM or designee to become familiar with the history of the Site, roles and responsibilities, field procedures, field data collection and management procedures, sample naming, split sample acceptance, communication procedures, and related QC requirements. Field oversight staff will also attend an onsite tailgate kick-off meeting immediately prior to the commencement of each stage or step of field activities. All new field oversight staff will receive comparable briefing if they were not at the initial field planning meeting and/or tailgate kick-off meeting. Supplemental meetings may be necessary as required by any changes in site conditions or to review field operation procedures.

The CDM Smith PM will identify any required field electronic data deliverable (EDDs) and assign the team member to be responsible for its preparation. The FOS will review the analytical method codes to be used in Scribe to ensure that they are consistent with EQuIS.

Field Equipment and Supplies

Equipment and field supply mobilization, governed by CDM Smith's Quality Procedures (QP) section 2.1, *Procuring Measurement and Test Equipment* and Section 5.3, *Inspection of Items*, will entail ordering, renting, and purchasing all supplies needed for each part of the Data Gap Investigation. This will also include staging and transferring all supplies to and from the site.

Field Procedures for these Activities are detailed in the Technical Standard Operating Procedures (TSOPs) below:

- TSOP 4-1 Field Logbook Content and Control*
- TSOP 4-2 Photographic Documentation of Field Activities



^{*-} Logbook notes should include field procedures used, descriptions of photos taken, problems encountered and notes of conversations with the Group'sfield staff. Details of samples collected including CLP numbers and visual observations.

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QAPP Worksheet # 17b - Sampling Design and Rationale Oversight Split Sampling (UFP-QAPP Manual Section 3.1.1) (EPA 2106-G-05 Section 2.3.1)

Describe and provide a rationale for choosing the sampling approach:

As part of the Project, the Group is implementing an investigation and field sampling program in support of the Data Gap Investigation. On behalf of the USACE and EPA, CDM Smith will provide oversight and will accept and analyze split samples. The oversight program is designed to provide technical review and evaluation of associated the Group's-implemented QAPPs and SAPs. Worksheet 10 of this QAPP states the oversight activities to occur during the field sampling programs, and Worksheet #11 provides details on the collection of split samples. Oversight forms are provided in Appendix D.

CDM Smith will accept split samples at a rate of approximately 10 percent to ensure that the Group's data is accurate. Locations for the split samples will be selected to cover a range of locations and concentrations, will address critical items such as areas of potential contamination, and will be from each media types samples (i.e. groundwater, surface water, porewater, sediment and soil) and in consultation with EPA/USACE if they provide any directions to split specific locations.

Field activities will be conducted according to the TSOPs below.

Describe the Sampling Action and Rationale in terms of: Matrix to be sampled and Frequency (including seasonal considerations):

Refer to Worksheets #10, 11, Table 1 and the text above for sampling and analysis rationale, matrices to be sampled, and analytical groups to be analyzed. Refer to Worksheets #11 and 18 for number of samples to be taken and sampling frequency. CDM Smith will accept split samples from the Group at a rate of approximately 10 percent to verify accuracy of the Group's generated data and to ensure their results are comparable.

Decontamination Procedures

Equipment decontamination procedures will be implemented by the Group in accordance with their QAPP, SAP and HASP to prevent cross contamination. CDM Smith will follow the Group's HASP prepared by their contractor.

Field Procedures for these Activities are detailed in:

- TSOP 1-2 Sample Custody
- TSOP 2-1 Packaging and Shipping Environmental Samples
- TSOP 4-1 Field Logbook Content and Control
- TSOP 4-2 Photographic Documentation of Field Activities, Sections 5.2.2 General Guidelines for Still Photography and 5.2.4 Photographic Documentation



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QAPP Worksheet #18: Sampling Locations and Methods (UFP-QAPP Manual Section 3.1.1 and 3.1.2) (EPA 2106-G-05 Section 2.3.1 and 2.3.2)

See Table 1



QAPP Worksheet #19 & 30: Sample Containers, Preservation, and Hold Times (UFP-QAPP Manual Section 3.1.2.2) (EPA 2106-G-05 Section 2.3.2)

Laboratory – CLP/DESA

List of required accreditations/certifications: DESA QAPP

Sample Delivery Method: FedEx Overnight

Analyte/ Analyte Group	Matrix	Analytical & Preparation Method/	Accreditation Expiration Date	Container(s) (number, size, and type per sample)	Preservation	Preparation Holding Time	Analytical Holding Time	Data Package Turnaround Time
VOCs				3-40 ml VOC vials with spin bars	0-6° C; store in dark	None	10 days	42 days
Soil Moisture	SOM01.2		Maintained by EPA	1-2oz jar	0-6° C; store in dark	None	10 days	42 days
SVOCs + SIM		SOM01.2		1-8 oz wide-mouth glass jar	0-6° C; store in dark	14 days	40 days	42 days
3VOCS + 31IVI		t			<-10° C; store in dark	1 year	40 days	42 days
Pesticides	Soil/Sediment				1-8 oz wide-mouth glass jar	0-6° C; store in dark	None	14 days
PCBs				1-8 oz wide-mouth glass jar	0-6° C; store in dark	None	14 days	42 days
Metals		ISM01.3	1-8 oz wide-mouth glass jar	0-6° C; store in dark	6 months	None	42 days	
Mercury				a a mana manamananananan	0-6° C	None	28 days	42 days
Cyanide, total				1-4 oz glass jar	0-6° C	None	14 days`	30 days



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QAPP Worksheet #19 & 30: Sample Containers, Preservation, and Hold Times (UFP-QAPP Manual Section 3.1.2.2) (EPA 2106-G-05 Section 2.3.2)

Laboratory – Subcontract Laboratory (TBD)
List of required accreditations/certifications: *Provided upon procurement of laboratory*Sample Delivery Method: *FedEx Overnight*

Analyte/ Analyte Group	Matrix	Analytical & Preparation Method/ SOP	Accreditation Expiration Date	Container(s) (number, size, and type per sample)	Preservation	Preparation Holding Time	Analytical Holding Time	Data Package Turnaround Time
PCB Congeners and homologs	Soil/Sediment	1668A	TBD	1-8 oz wide-mouth glass jar	0-6° C; or <-10° C; store in dark	1 year	None	42 days
Dioxin/Furans		1613B		1-8 oz wide-mouth glass jar	0-6° C; <-10° C; store in dark	1 year	None	42 days



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QAPP Worksheet #19 & 30: Sample Containers, Preservation, and Hold Times (UFP-QAPP Manual Section 3.1.2.2) (EPA 2106-G-05 Section 2.3.2)

Laboratory – CLP/DESA
List of required accreditations/certifications: DESA QAPP

Sample Delivery Method: FedEx Overnight

Analyte/ Analyte Group	Matrix	Analytical & Preparation Method/	Accreditation Expiration Date	Container(s) (number, size, and type per sample)	Preservation	Preparation Holding Time	Analytical Holding Time	Data Package Turnaround Time
VOCs		SOM01.2 Aqueous	SOM01.2	3- 40-ml voa vials	0-6°C, HCL to pH<2	7 days	40 days	42 days
SVOCs + SIM				2- 1-L amber glass with PTFE-lined lid	0-6°C; store in the dark	7 days	40 days	42 Days
Pesticides	Λαμρομε			2- 1-L amber glass with PTFE-lined lid	0-6°C; store in the dark	7 days	40 days	42 Days
PCBs	Aqueous		100	2- 1-L amber glass with PTFE-lined lid	0-6°C; store in the dark	7 days	40 days	42 Days
Metals		ISM01.3		1-1L HDPE	0-6°C; field filter samples or within 24 hours; HNO ₃ to pH <2	None	6 months	42 Days
Cyanide				1-500ml HDPE	0-6°C; NaOH to pH >12	14 days	28 days	42 Days



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QAPP Worksheet #19 & 30: Sample Containers, Preservation, and Hold Times (UFP-QAPP Manual Section 3.1.2.2) (EPA 2106-G-05 Section 2.3.2)

Laboratory – Subcontract Laboratory (TBD)

List of required accreditations/certifications: *Provided upon procurement of laboratory*

Sample Delivery Method: FedEx Overnight

Analyte/ Analyte Group	Matrix	Analytical & Preparation Method/	Accreditation Expiration Date	Container(s) (number, size, and type per sample)	Preservation	Preparation Holding Time	Analytica I Holding Time	Data Package Turnaround Time
PCB congeners and Homologs	Aqueous	1668A		2-1L amber glass PTFE lid	0-6°C; store in the dark	None	1 year	30 days
Dioxin/ Furans	Aqueous	1613	TBD	2-1L amber glass PTFE lid	0-6°C; store in the dark	None	1 year	30 days
Total mercury		1631E		1-250 mL FLPE	0-6°C, HCL to pH <2, no headspace	28 days	90 days	30 days



QAPP Worksheet #20: Field Quality Control Summary (UFP-QAPP Section 3.1.1 and 3.1.2) (EPA 2106-G-05 Section 2.2.6)

Matrix	Analyte/Analytical Group	Method/ SOP	The Group's Environmental Analyses Total	CDM Smith Split Samples	Matrix Spike/Matrix Spike Duplicate (MS/MSD)	Trip Blanks	Total
	VOCs	SOM01.2	40	4	NA*		4
	SVOCs + SIM	SOM01.2	40	4	NA	1	4
	Pesticides	SOM01.2	40	4	1	1	4
Soil	PCBs Aroclors	SOM01.2	40	4	1	NA	4
3011	TAL Metals, Mercury, Cyanide	ISM01.3	40	4	1	INA	4
	PCB Congeners	EPA 1668	14	2	1		2
	Dioxins and Furans	EPA 1613	14	2	1		2
	VOCs	SOM01.2	9	1	NA	1	2
	SVOCs + SIM	SOM01.2	9	1	NA		1
	Pesticides	SOM01.2	9	1	1		1
Groundwater	PCBs Aroclors	SOM01.2	9	1	1		1
(temporary wells)	TAL Metals, Mercury and Cyanide (unfiltered)	ISM01.3	9	1	1	NA	1
	TAL Metals, Mercury and Cyanide (filtered)	ISM01.3	9	1	1		1
	VOCs	SOM01.2	48	5	NA	3	8
	SVOCs+ SIM	SOM01.2	48	5	NA		5
	Pesticides	SOM01.2	48	5	1	1	5
Groundwater	PCB Aroclors	SOM01.2	48	5	1]	5
(permanent wells)	TAL Metals, Mercury and Cyanide (unfiltered)	ISM01.3	48	5	1	NA	5
	TAL Metals, Mercury and Cyanide (filtered)	ISM01.3	48	5	1		5



QAPP Worksheet #20: Field Quality Control Summary (UFP-QAPP Section 3.1.1 and 3.1.2) (EPA 2106-G-05 Section 2.2.6)

Matrix	Analyte/Analytical Group	Method/ SOP	The Group's Total Analyses	CDM Smith Split Samples	Matrix Spike/Matrix Spike Duplicate (MS/MSD)	Trip Blanks	Total
	VOCs	SOM01.2	2	1	NA**	1	2
	SVOCs+ SIM	SOM01.2	2	1			1
	Pesticides	SOM01.2	2	1			1
Porewater	PCBs Aroclors	SOM01.2	2	1			1
Porewater	TAL Metals, Mercury and Cyanide (unfiltered)	ISM01.3	2	1		NA	1
	TAL Metals, Mercury and Cyanide (filtered)	ISM01.3	2	1			1
	VOCs	SOM01.2	11	1	NA	1	2
	SVOCs + SIM	SOM01.2	11	1	NA		1
	Pesticides	SOM01.2	11	1	1		1
	PCBs Aroclors	SOM01.2	11	1	1		1
Surface Water	TAL Metals and Cyanide (unfiltered)	ISM01.3	11	1	1		1
	TAL Metals and Cyanide (Filtered)	ISM01.3	11	1	1		1
	Trace Mercury	EPA 1631E	11	1	1	NA	1
	VOCs	SOM01.2	11	1	NA		
	SVOCs + SIM	SOM01.2	11	1	NA		1
Codinoont	Pesticides	SOM01.2	11	1	1		1
Sediment	PCBs Aroclors	SOM01.2	11	1	1		1
	TAL Metals, Mercury and Cyanide	ISM01.3	11	1	1		1

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QAPP Worksheet #20: Field Quality Control Summary (UFP-QAPP Section 3.1.1 and 3.1.2) (EPA 2106-G-05 Section 2.2.6)

Notes:

*No extra volume required but may need to be designated on chain of custody depending on laboratory assigned. MS/MSDs are not counted as an extra sample they are additional volumes provided for laboratory QC.

** Limited volume sample

Abbreviations NA- not applicable



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QAPP Worksheet #21: Field SOPs (UFP-QAPP Manual Section 3.1.2) (EPA 2106-G-05 Section 2.3.2)

SOP # or reference	Title, Revision, Date, and URL (if available)	Originating Organization	SOP option or Equipment Type (if SOP provides different options)	Modified for Project? Y/N	Comments
1-2	Sample Custody, Rev. 7, January 2012	CDM Smith	NA	N	Sample tags are not required.Scribe generated COCs will be used.Use waterproof ink for any handwritten labels.
2-1	Packaging and Shipping Environmental Samples, Rev. 5, January 2012	CDM Smith	TSOP Section 1.3 lists materials needed	N	Vermiculite shall not be used
4-1	Field Logbook Content and Control, Rev. 7, January 2012	CDM Smith	NA	N	Logbook notes should include decontamination procedures; descriptions of photographs taken; problems encountered and notes of conversations with PM, USACE, EPA, the Group's contractor; and details of samples collected including CLP numbers and visual observations ¹ .
4-2	Photographic Documentation of Field Activities, January 2012	CDM Smith	Camera	N	

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QAPP Worksheet #21: Field SOPs (UFP-QAPP Manual Section 3.1.2) (EPA 2106-G-05 Section 2.3.2)

The following information will be recorded (at a minimum) in the field logbook for each sample collected and shipped:

- Name of field personnel
- CDM Smith assigned sample number/location
- Date sampled
- Date shipped
- Sample location number
- Corresponding CLP routine analytical services (RAS) sample number
- Media type
- Type of analysis to be performed
- Sample volume and containers
- Any unusual discoloration or evidence of contamination
- Field parameter measurements
- Preservatives added to sample
- Courier airbill number and means of delivery to the laboratory
- General observations



QAPP Worksheet #23: Analytical SOPs (UFP-QAPP Manual Section 3.2.1) (EPA 2106-G-05 Section 2.3.4)

SOP#	Title, Date, and URL (if available)	Definitive or	Matrix/ Analytical	SOP Option or	[‡] Modified for Project?
		Screening Data	Group	Equipment Type	Y/N
SOM01.2	Multi-Media, Multi-Concentration, Organic Analytical Service for Superfund. EPA 2005, amended 4.11.2007		VOC, SVOC, Pesticides	GC/MS GC/ECD; FID	
EPA 1613B	Analytical Method for the Determination of Polychlorinated Dibenzodioxins and Dibenzofurans. Revision 20. March 2011.		PCDD/PCDF	High resolution (HR) GC/HRMS	
EPA 1668A	Chlorinated Biphenyl Congeners in Water, Soil, Sediment, Biosolids, and Tissue by HRGC/HRMS. November 2008.	Definitive	PCB Congeners	HRGC/HRMS	
EPA 1631E	Total Mercury Using Atomic Fluorescence Spectroscopy. Revision 2. August 28, 2009.		Mercury (trace)	cold vapor atomic fluorescence spectrometry (CVAFS)	
	CLP SOW for Multi-Media, Multi-Concentration		Target Analyte List (TAL) Metals	ICP-AES/ICP-MS	
ISM01.3	Inorganic Analysis. December 2006.		Mercury	CVAA	
	Inductively Coupled Plasma – Mass Spectrometry Analysis. Revision 2. April 1, 2011.		Cyanide	Colorimeter	

Notes:

- 1. EPA reviews CLP laboratories SOPs. DESA laboratory SOPs will apply and not these generic SOPs when the DESA laboratory is able to perform the analyses. CDM Smith subcontract laboratory specific SOPs are not available (NA) at this stage since the Region 2 FASTAC Policy will be implemented for procuring laboratory services. However, some of the listed analyses will be sent to a MSA subcontract laboratory to match the Respondents specific and unique analytical requirements and facilitate comparison of the data. Subcontract Laboratory SOPs are TBD.
- 2. For non-routine analytical services (RAS) data, the ASC will submit the electronic "Analytical Services Tracking System (ANSETS) Data Requirement" form to the RSCC by the first day of each month for the previous month's sampling.



QAPP Worksheet #24: Analytical Instrument Calibration (UFP-QAPP Manual Section 3.2.2) (EPA 2106-G-05 Section 2.3.6)

Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	Corrective Action	Title/Position Responsible for Corrective Action	SOP Reference
HRGC/ HRMS	Initial Calibration and calibration verification check: Per laboratory SOP	After set up, prior to run and after instrument changes or failures of checks.	% RSD and % recovery per laboratory SOPs.	Check, correct; re-calibrate and rerun all samples analyzed after last valid calibration check	Laboratory analyst / QA officer - TBD	TDD
and HRGC/LRMS	Calibration checks: CCVs per laboratory SOP	Daily: Beginning of run and after every 10 samples and at end of analytical run	% recovery per laboratory SOP	Check, correct; re-calibrate and rerun all samples analyzed after last valid cal check	Laboratory analyst / QA officer - TBD	TBD
GC/MS GC GC/FID	Initial calibration: 5 points standards	Upon award of the contract, whenever the laboratory takes corrective action which may change or affect the initial calibration criteria (e.g., ion source cleaning or repair, column replacement, etc.), or if the continuing calibration acceptance criteria have not been met.	Relative response factor (RRF) ≥ minimum acceptable RRF listed in Table 5 of procedure. All target compounds, initial relative standard deviation (RSD) ≤ 10% or 20% and correlation coefficient (r)> 0.995. %RSD ≤ value in Table 5 of SOM01.2 or other laboratory SOP as applicable.	Inspect system for problems (e.g., clean ion source, change the column, service the purge and trap device), correct problem, re-calibrate.	Laboratory Technician	TBD
GC/MS	Calibration Standards Verification	Each lot of standards	As per laboratory established control limits	Inspect system; correct problem; re-run standard and affected samples	Laboratory GC/MS Technician	
GC/MS	Tuning	Daily: every 12 hours	Response factors and RRF as method specified	Inspect system; correct problem; re-run standard and affected samples	Laboratory GC/MS Technician	TBD
GC/FID	Mass Discrimination Check	Every 12 hours	RF ratio of C32/C20 should be >0.8	Per laboratory SOP	Laboratory GC/FID Technician	

Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	Corrective Action	Title/Position Responsible for Corrective Action	SOP Reference
CVAFS	Per method and laboratory SOP	Calibration	Per method/ laboratory SOP. ICAL ≤15%RSD.	Increase the system correct	Assigned	
		ICV: Check daily when instrument is in use	85-115% R for Total mercury	Inspect the system, correct problem, re-calibrate, and re-analyze samples.	laboratory personnel	TBD
		CCV: Beginning and after every 10 samples	77-123% R for total mercury	re-analyze samples.	personner	
CV-GAS	Calibration; 3 point st andards	After instrument set up	$R^3 \ge 0.995$	Inspect system; correct problem	Laboratory Technician	TBD
	Initial Calibration Verification (ICV)	Before sample analysis	80-120% R; source of standard separate from calibration standards	Do not analyze samples until problem is corrected		
	CCV	10% or every 2 hours, whichever is more frequent	80-120% R	Inspect system, re-calibrate and rerun associated samples		
	See ISM01.3/ per instrument manufacturer's procedures	Initial calibration: daily or once every 24 hours and each time the instrument is set up	ICP-MS: As per instrument manufacturer's procedures, at least 2 standards	Inspect the system, correct problem, re-calibrate, and re-analyze samples		
	Initial calibration	Daily; after tuning and optimizing instrument	Correlation coefficient >0.995 with a minimum of 3 standards and a blank	Repeat analysis; re-prepare calibration standards and reanalyze	Laboratory ICP-AES / ICP-MS	
ICP-MS / AES ISM01.3	ICV	Before sample analysis	90-110% R; source of standard separate from calibration standards	Re-calibrate instrument; prepare fresh ICV standards; do not analyze samples until problem is corrected	Technician or DESA Laboratory analyst / QA officer	TBD
	Reporting Limit Standard	After initial calibration verification standard	80-120% R or concentration ≤ 30% difference (from true value)	Re-analyze failed standard	officer	
	ccv	Every 10 samples and at end of analytical sequence	90-110% R; source of standard separate from calibration standards	Re-check; re-calibrate and rerun all samples analyzed after last valid CCV		

QAPP Worksheet #24: Analytical Instrument Calibration (UFP-QAPP Manual Section 3.2.2) (EPA 2106-G-05 Section 2.3.6)

Instrument	Calibration Procedure	Frequency of Calibration	Acceptance Criteria	Corrective Action	Title/Position Responsible for Corrective Action	SOP Reference
Colorimeter	Initial Calibration; 4 - 9 point standards	Every 3 months or as per Laboratory SOP	90-110 % R	Re-check; re-calibrate		
	Calibration check (Cal Check)	Every 10 samples and at end of analytical run	80-120 % R	Re-check; re-calibrate and rerun all samples analyzed after last valid Calibration Check	Laboratory analyst / QA officer - TBD	TBD
Thermometer	Calibration	Quarterly; serviced annually	±1°C of true value of National Institute of Standards and Technology traceable thermometer	Replace defective thermometer		
pH meter	Daily buffer checks (2 point bracketing sample pH)	Before use/per batch; other checks as per rental company and manufacturer's recommendations	± 0.1 pH units or ± 0.05 pH units	Recheck; replace bufferlutions and recheck. If still fails perform instrument check or place out of service	Laboratory analyst / QA officer - TBD	TBD

Notes:

- 1. The FASTAC decision process will be used for procuring laboratory services. CLP, DESA and CDM Smith subcontract laboratory's calibration and/or method SOPs will be utilized to meet calibration criteria. Specific instrument information (Manufacturer and model) is not available at this time.
- 2. To be determined (TBD) Reference SOP depends on the laboratory assignment. EPA maintains the CLP laboratory SOP information. If a subcontract laboratory is needed, CDM Smith will submit their SOP as a field change request.
- 3. R represents the correlation coefficient.
- 4. The laboratory SOP will include the calibration range information.
- 5. NJDEP=New Jersey Department of Environmental Protection http://www.nj.gov/dep/srp/guidance/fspm/pdf/chapter06e.pdf).



QAPP Worksheet #25: Analytical Instrument and Equipment Maintenance, Testing, and Inspection (UFP-QAPP Manual Section 3.2.3) (EPA 2106-G-05 Section 2.3.6)

Instrument/ Equipment	Maintenance Activity	Testing Activity Inspection Activity	Frequency	Acceptance Criteria	Corrective Action	Title/Position Responsible for Corrective Action	Reference
utilized for analytic	al services. Informatio	n is provided in CDM S	rmation and availability Smith MSA subcontract and inspection frequen	laboratories' QA Ma	nuals. The MSA labor	atory to be utilized (
ICP-MS/AES	As per instrument manufacturer's recommendations	As per instrument manufacturer's recommendations; check connections	As per instrument manufacturer's recommendations	Acceptable re-calibration; see ISM01.3	Inspect the system, correct problem, re-calibrate and/or reanalyze samples	EPA CLP Laboratory ICP-MS/AES Technician	ISM01.3
GC/MS	See SOM01.2; as per instrument manufacturer's recommendations	See SOM01.2; as per instrument manufacturer's recommendations	See SOM01.2; as per instrument manufacturer's recommendations	Acceptable re-calibration; see SOM01.2	Inspect the system, correct problem, re-calibrate and/or reanalyze samples	EPA CLP Laboratory GC/MS Technician	SOM01.2
GC/ECD	See SOM01.2; as per instrument manufacturer's recommendations	See SOM01.2; as per instrument manufacturer's recommendations	See SOM01.2; as per instrument manufacturer's recommendations	Acceptable re-calibration; see SOM01.2	Inspect the system, correct problem, re-calibrate and/or reanalyze samples	EPA CLP Laboratory GC/ECD Technician	SOM01.2
CVAFS	Replace disposables, Flush lines	Sensitivity check Check connections	Daily or as needed	See SOP	See SOP	Analyst or Section Supervisor	EPA 1631



QAPP Worksheet #26 & 27: Sample Handling, Custody, and Disposal (UFP-QAPP Manual Section 3.3) (EPA 2106-G-05 Section 2.3.3)

Sampling Organization: CDM Smith

Laboratory: DESA, CLP

Method of sample delivery (shipper/carrier): FedEx

Number of days from reporting until sample disposal: DESA 30 days or CLP as per contract or subcontract laboratory 60-90 days as specified in SOW

Activity	Organization and title or position of person responsible for the activity	SOP reference		
Sample labeling	CDM Smith - FOS	TSOP 2-1		
Chain-of-custody form completion	CDM Smith – Sample manager	TSOP 1-2		
Packaging	CDM Smith – Sample manager	TSOP 1-2 and 2-1; EPA CLP Guidance for Field Samplers		
Shipping coordination	CDM Smith - FOS, CDM Smith ASC/ CLP coordinator	TSOP 2-1		
Sample receipt, inspection, & log-in	Laboratory custodian (DESA, CLP, or Subcontract)	Analytical Scope of work and Laboratory SOP		
Sample custody and storage	CDM Smith and Laboratories (DESA, CLP, or Subcontract)	TSOP 1-2; Analytical SOW or Laboratory TSOP		
Sample disposal	Laboratory Custodian (DESA, CLP, or Subcontract)	Laboratory TSOP		

Notes:

Trip blanks will be identified using the abbreviation TB followed by the date. For example, TB-070108 indicates that the trip blank was collected on July 1, 2008. Split samples will be identified using the abbreviation CDM- followed by the Group sample name

Filtered samples will be identified by adding the letter F to the end of the location identifier. For example, CDM-xxxx-F would indicate a split filtered sample



QAPP Worksheet #28a: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Aqueous

Analytical Group TCL VOCs Low $(\mu g/L)^{**}$

Analytical Method/SOP Reference SOM01.2

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
Split Samples	1 per 20 samples	None		Notify PM and flag duplicate results	CDM Smith ASC and FOS	≤50% RPD when both samples ≥ CRQL otherwise ABS ≤ 5xQL	
Temperature Blank	1 per cooler	0 to 6 degrees C		Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation	
Trip Blank	1 per cooler	≤ CRQL		Verify results; re-analyze. Flag outliers	Laboratory analyst	≤ CRQL	
Method Blank	1 every 12 hours	No analyte > CRQL*		Suspend analysis unit source recertified	DESA or	No analyte >	CRQL*
Deuterated Monitoring	ring all samples	Vinyl chloride-d3	65-131 %R	Check calculations and instruments,	EPA CLP Laboratory GC/MS	Vinyl chloride-d3	65-131 %R
Compounds		Chloroethane-d5	71-131 %R	reanalyze affected samples	Technician	Chloroethane-d5	71-131 %R

^{*}with the exception of methylene chloride, 2-butanone and acetone which can be up to 2 times the CRQL.



QAPP Worksheet #28a: Analytical Quality Control and Corrective Action (TCL VOCs Aqueous continued)

QC Sample:	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
			TCL Low \	OCs Continued			
		1,1-Dichloroethene-d2	55-104 %R	Check calculations and instruments, reanalyze affected samples; up to 3 DMCs per sample may fail to meet recovery limits	DESA or EPA CLP Laboratory GC/MS Technician	1,1-Dichloroethene-d2	55-104 %R
		2-Butanone-d5	49-155 %R			2-Butanone-d5	49-155 %R
		Chloroform-d	78-121 %R			Chloroform-d	78-121 %R
		1,2-Dichloroethane-d4	78-129 %R			1,2-Dichloroethane-d4	78-129 %R
Davitamatad	all samples	Benzene-d6	77-124 %R			Benzene-d6	77-124 %R
Deuterated Monitoring		1,2-Dichloropropane-d6	79-124 %R			1,2-Dichloropropane-d6	79-124 %R
Compounds [cont'd]		Toluene-d8	77-121 %R			Toluene-d8	77-121 %R
		trans-1,3-Dichloropropene-d4	73-121 %R			trans-1,3-Dichloropropene-d4	73-121 %R
		2-Hexanone-d5	28-135 %R			2-Hexanone-d5	28-135 %R
		1,4-Dioxane-d8	50-150 %R			1,4-Dioxane-d8	50-150 %R
		1,1,2,2-Tetrachloroethane-d2	73-125 %R			1,1,2,2-Tetrachloroethane-d2	73-125 %R
		1,2-Dichlorobenzene-d4	80-131 %R			1,2-Dichlorobenzene-d4	80-131 %R
Internal Standards	all samples	60-140%	Check calculations and instruments, reanalyze affected samples		± 40% of response area, ± 20 sec retention time shift		



QAPP Worksheet #28a: Analytical Quality Control and Corrective Action (TCL VOCs Aqueous continued)

MatrixAqueousAnalytical GroupTCL VOCsAnalytical Method/SOP ReferenceSOM01.2

QC Sample	Frequency/ Number	Method/SOP QC Accepta	ince Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
	1 per 20 samples or less (if requested)	1,1-Dichloroethene	61-145 %R	Flag outliers	- DESA/ CLP Laboratory GC/MS Technician	1,1-Dichloroethene	61-145 %R
Matrix Spike (Not Required)		Trichloroethene	76-127 %R			Trichloroethene	76-127 %R
		Benzene	71-120 %R			Benzene	71-120 %R
		Toluene	76-125 %R			Toluene	76-125 %R
		Chlorobenzene	75-130 %R			Chlorobenzene	75-130 %R
Matrix Spike Duplicate (Not Required)	1 per 20 samples or less (if requested)	1,1-Dichloroethene	0-14 %RPD	Flag outliers		1,1-Dichloroethene	0-14 %RPD
		Trichloroethene	0-11 %RPD			Trichloroethene	0-11 %RPD
		Benzene	0-14 %RPD			Benzene	0-14 %RPD
		Toluene	0-13 %RPD			Toluene	0-13 %RPD
		Chlorobenzene	0-13 %RPD			Chlorobenzene	0-13 %RPD



QAPP Worksheet #28b: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixAqueousAnalytical GroupTCL SVOCsAnalytical Method/SOP ReferenceSOM01.2

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Split Samples	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤50% RPD when both results ≥ CRQL otherwise ABS ≤ 5xQL
Temperature Blank	1 per cooler	0 to 6 degrees C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Method Blank	1 per ≤20 samples or batch	No analyte > CRQL*	Stop analysis, re-extract and reanalyze affected samples	DESA or CLP Laboratory GC/MS Technician	≤ CRQL

^{*}with the exception of bis (2-Ethylhexyl) phthalate which can be up to 5 times the CRQL. (EPA CLP National Functional Guidelines)



QAPP Worksheet #28b: Analytical Quality Control and Corrective Action (continued)

Laboratory QC Sample	Frequency / Number	Method/SOP QC Acceptance Limits TCL SVOC		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
		Phenol-d5	39-106 %R			Phenol-d5	39-106 %R
		Bis(2-chloroethyl)ether-d8	40-105 %R			Bis(2-chloroethyl)ether-d8	40-105 %R
		2-Chlorophenol-d4	41-106 %R			2-Chlorophenol-d4	41-106 %R
		4-Methylphenol-d8	25-111 %R			4-Methylphenol-d8	25-111 %R
		Nitrobenzene-d5	43-108 %R			Nitrobenzene-d5	43-108 %R
		2-Nitrophenol-d4	40-108 %R	Check calculations and instruments, reanalyze affected samples; up to 4		2-Nitrophenol-d4	40-108 %R
		2,4-Dichlorophenol-d3	37-105 %R		DECA CLD	2,4-Dichlorophenol-d3	37-105 %R
Deuterated	l all	4-Chloroaniline-d4	1-145 %R		DESA or CLP Laboratory GC/MS Technician	4-Chloroaniline-d4	1-145 %R
Monitoring		Dimethylphthalate-d6	47-114 %R			Dimethylphthalate-d6	47-114 %R
Compounds	samples	Acenaphthylene-d8	41-107 %R	DMCs may fail to		Acenaphthylene-d8	41-107 %R
		4-Nitrophenol-d4	33-116 %R	meet recovery		4-Nitrophenol-d4	33-116 %R
		Fluorene-d10	42-111 %R	limits		Fluorene-d10	42-111 %R
		4,6-Dinitro-2-methylphenol -d2	22-104 %R			4,6-Dinitro-2-methylphen ol-d2	22-104 %R
		Anthracene-d10	44-110 %R			Anthracene-d10	44-110 %R
		Pyrene-d10	52-119 %R			Pyrene-d10	52-119 %R
		Benzo(a)pyrene-d12	32-121 %R			Benzo(a)pyrene-d12	32-121 %R
		,		Check	DESA or CLP		
Internal	all	50-100% of area, <u>+</u> 20 second	l retention	calculations/instru	Laboratory	50-100% of area, <u>+</u> 20 second retention	
Standards	samples	time shift		ments reanalyze	GC/MS	time shift	
				affected samples	Technician		

QAPP Worksheet #28b: Analytical Quality Control and Corrective Action (TCL SVOCs Aqueous continued)

MatrixAqueousAnalytical GroupTCL SVOCsAnalytical Method/SOP ReferenceSOM01.2

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
	Phenol	12-110 %R			Phenol	12-110 %R	
	1 per 20	2-Chlorophenol	27-123 %R	Flag outliers		2-Chlorophenol	27-123 %R
Matrix Spike (Not san	samples or less (if requested)	N-Nitroso-di-n-propylamine	41-116 %R			N-Nitroso-di-n- propylamine	41-116 %R
		4-Chloro-3-methylphenol	23-97 %R			4-Chloro-3-methylphenol	23-97 %R
		Acenaphthene	46-118 %R		DESA/ CLP Laboratory GC/MS Technician	Acenaphthene	46-118 %R
		4-Nitrophenol	29-94 %R			4-Nitrophenol	29-94 %R
Matrix Spike	1 per 20	2,4-Dinitrotoluene	24-96 %R			2,4-Dinitrotoluene	24-96 %R
Duplicate (Not	samples or less (if requested)	Pentachlorophenol	9-103 %R	Flag outliers		Pentachlorophenol	9-103 %R
Required)		Pyrene	26-127 %R			Pyrene	26-127 %R
		Phenol	0-42 %RPD			Phenol	0-42 %RPD



QAPP Worksheet #28c: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixAqueousAnalytical GroupDioxins/FuransAnalytical Method/SOP ReferenceEPA 1613B

QC Sample:	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Method Blank	1 per 20 samples	TCDD/F <0.5 pg/sample, PeCDD/F, HxCDD/F, HpCDD/F <1.0 pg/sample, OCDD/F <5 pg/sample unless sample concentrations > 10* blank levels (per SOP)	If samples non-detect or if lowest sample result is >10 times the blank-no action; otherwise redigest and reanalyze	Laboratory Analyst	No analyte > QL
Laboratory Duplicate	1 per 20 samples	± 20% mean for concentrations >10*QL	Investigate and correct; Flag outliers	Laboratory Analyst	± 20% of mean if sample concentration >10x DL ¹
Initial Precision and Recovery	Prior to sample analysis	Per laboratory SOP	Investigate and correct	Laboratory Analyst	Per method/laboratory SOP
Ongoing Precision and Recovery	1 per batch of 20 samples	Per laboratory SOP or 70-130%R	Identify source of problem, make other adjustments; redigest if needed and reanalyze	Laboratory Analyst	Individual laboratory established limits per SOP
Split Samples	1 per 20 samples	None	Data assessor to inform PM if MPC is exceeded; address in data quality assessment	CDM Smith ASC	≤ 40% RPD (for results ≥ 5QL)
Surrogates	1 per 20 samples	25-120%R-warning limit 17-130%R-control limit	Investigate and correct	Laboratory Analyst	25-120%R-warning limit 17-130%R-control limit
Temperature Blank	1 per cooler	0 to 6 degrees C	Note outlier in laboratory narrative. Inform CDM Smith of failure and need for additional coolant; check packing procedure	Laboratory Analyst	≤ 10 degrees C for data validation

The DLs referenced in laboratory SOP are equivalent to the QLs or sample reporting limits.



QAPP Worksheet #28e: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixAqueousAnalytical GroupTCL PesticidesAnalytical Method/SOP ReferenceSOM01.2

QC Sample:	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
Split Samples	1 per 20 samples	None		Notify PM and flag duplicate results	CDM Smith ASC and PM	≤50% RPD when both samples ≥ CRQL otherwise ABS ≤ 5xQL	
Temperature Blank	1 per cooler	0 to 6 degrees C		Laboratory - inform RSCC/CDM Smith and note in data narrative. CDM Smith - check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation	
Method Blank	1 per ≤20 samples or whenever samples extracted	No analyte > CRQL		Suspend analysis; re-extract and reanalyze blank and affected samples	DESA or CLP Laboratory GC/ECD Technician	Analyte ≤ CRQL	
	1 per ≤20 samples; if	gamma-BHC (Lindane)	56-123 %R	Flag outliers		gamma-BHC	56-123 %R
		Heptachlor	40-131 %R			Heptachlor	40-131 %R
		Aldrin	40-120 %R		DESA or CLP Laboratory GC/ECD Technician	Aldrin	40-120 %R
Matrix Spike	requested	Dieldrin	52-126 %R			Dieldrin	52-126 %R
		Endrin	56-121 %R	_		Endrin	56-121 %R
		4,4'-DDT	38-127 %R	_		4,4'-DDT	38-127 %R
		gamma-BHC	0-15 %RPD	Flag outliers		gamma-BHC	0-15 %RPD
		Heptachlor	0-20 %RPD			Helptachlor	0-20 %RPD
Matrix Spike	1 per ≤20 samples; if	Aldrin	0-22 %RPD	1		Aldrin	0-22 %RPD
•	requested	Dieldrin	0-18 %RPD	1		Dieldrin	0-18 %RPD
		Endrin	0-21 %RPD	1		Endrin	0-21 %RPD
		4,4'-DDT	0-27 %RPD			4,4'-DDT	0-27 %RPD

QAPP Worksheet #28e: Analytical Quality Control and Corrective Action (continued)

QC Sample:	Frequency/Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Perforn	nance Criteria
		Heptachlor epoxide	50-150 %R	Check calculations and instruments, reanalyze affected samples	DESA or CLP Laboratory GC/ECD Technician	Heptachlor epoxide	50-150 %R
		Dieldrin	30-130 %R			Dieldrin	30-130 %R
	1 per 20 samples	gamma-BHC	50-120 %R			gamma-BHC	50-120 %R
LCS [cont'd]		4,4'-DDE	50-150 %R			4,4'-DDE	50-150 %R
[cont u]		Endrin	50-120 %R			Endrin	50-120 %R
		Endosulfan sulfate	50-120 %R			Endosulfan sulfate	50-120 %R
		gamma-Chlordane	30-130 %R			gamma-Chlordane	30-130 %R
Surrogate	all samples	30-150 %R		Check calculations and instruments, reanalyze affected samples	DESA or CLP Laboratory GC/ECD Technician	30-150 %R	

QAPP Worksheet #28f: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixAqueousAnalytical GroupTCL PCBsAnalytical Method/SOP ReferenceSOM01.2

QC Sample:	Frequency/Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
Split Samples s	1 per 20 samples	None		Notify PM and flag duplicate results	CDM Smith ASC and PM	50% RPD when PCBs in both samples ≥ QL Otherwise ABS ≤ 5xQL	
Temperature Blank	1 per cooler	0 to 6 degrees C		Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation	
Method Blank	1 per ≤20 samples or whenever samples extracted	No analyte > CRQL		Suspend analysis – reextract and reanalyze affected samples	DESA or CLP Laboratory GC/ECD Technician	No analyte > CRQL No target analyte concentrations ≥ ½ CRQL	
Matrix Spike		Aroclor-1016	29-135 %R			Aroclor-1016	29-135 %R
	1 per ≤20 samples;	Aroclor-1260	29-135 %R	Flag outliers	DESA or CLP Laboratory	Aroclor-1260	29-135 %R
Matrix Spike	if requested	Aroclor-1016	0-15 %RPD	Tiug outliers	GC/ECD Technician	Aroclor-1016	0-15 %RPD
Duplicate		Aroclor-1260	0-20 %RPD			Aroclor-1260	0-20 %RPD
		Aroclor-1016	50-150 %R	Check calculations and	DESA or CLD Laboratory	Aroclor-1016	50-150 %R
LCS	1 per ≤20 samples Aroclor-126		50-150 %R	instruments, reanalyze affected samples	DESA or CLP Laboratory GC/ECD Technician	Aroclor-1260	50-150 %R
Surrogate	all samples	30-150 %R		Check calculations and instruments, reanalyze affected samples	DESA or CLP Laboratory GC/ECD Technician	30-150 %R	



QAPP Worksheet #28g: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixAqueousAnalytical GroupPCB CongenersAnalytical Method/SOP ReferenceEPA 1668A

QC Sample:	Frequency/ Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Method Blank	1 per 20 samples	Concentration < 2 pg, 10 pg or 50 pg/sample-See SOP. Sum of all congeners < 300 pg /sample unless sample concentrations > 10* blank levels	If samples non-detect or if lowest sample result is >10 times the blank-no action; otherwise redigest and reanalyze	Laboratory Analyst	No analyte > QL
Analysis (Laboratory) Duplicate	1 per 20 samples	± 20% mean for concentrations >10*QL	Flag outliers	Laboratory Analyst	RPD ≤ 40% for concentrations >10x DL¹; otherwise ABS <ql< td=""></ql<>
Quality Control Sample	Periodically at least quarterly	50-150%R;	Check standards; recalibrate if required	Laboratory Analyst	70-130%R;
Calibration Verification Sample	Beginning of each 12-hour shift	70-130%R;	Adjust and/or recalibrate	Laboratory Analyst	70-130%R
Initial Precision and Recovery	Prior to sample analysis	Per laboratory SOP	Investigate and correct	Laboratory Analyst	60-140%R ≤ 40% RSD
Ongoing Precision and Recovery	1 per batch of 20 samples	Per laboratory SOP	Identify source of problem, recalibrate if needed/ make other adjustments and reanalyze	Laboratory Analyst	Warning 70-130%R; Accept 50-150%R
Split Samples	1 per 20 samples	None	Data assessor to inform PM if MPC is exceeded; address in data quality assessment	CDM Smith ASC	RPD ≤ 40%; ABS <ql <5x<br="" for="" samples="">QL</ql>
Temperature Blank	1 per cooler	0 to 6 degrees C	Note outlier in laboratory narrative. Inform CDM Smith of failure and need for additional coolant; check packing procedure	Laboratory Analyst	≤ 10 degrees C for data validation

Notes: 1. The DLs referenced in the laboratory SOP are equivalent to the QLs or sample reporting limits.



QAPP Worksheet #28h: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Aqueous

Analytical Group TAL Metals ICP MS/AES

Analytical Method/SOP Reference ISM01.3 or current method

QC Sample:	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Split Samples	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤50%RPD, ABS ≤ 5xCRQL when any result ≤ CRQL
Temperature Blank	1 per cooler	0 to 6 °C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Preparation Blank	1 per ≤20 samples	No constituent > CRQL	Suspend analysis rectify source; redigest and reanalyze affected samples	DESA or CLP Laboratory ICP Technician	No constituent > CRQL
Spike	1 per ≤20 samples	75-125%R*	Flag outliers	DESA or CLP Laboratory ICP Technician	75-125%R*
Laboratory Duplicate	1 per ≤20 samples	± 20% RPD**	Flag outliers	DESA or CLP Laboratory ICP Technician	≤20% RPD**
Post-Digestion Spike	after any analyte (except Ag) fails spike %R	75-125%R	Flag outliers	DESA or CLP Laboratory ICP Technician	75-125%R
Interference Check Sample [ICP Analysis Only]	Beginning of each run	± CRQL + true value or ± 20% of true value, whichever is greater	Check calculations and instruments, reanalyze affected samples	DESA or CLP Laboratory ICP Technician	± 2 times CRQL of true value or ± 20% of true value, whichever is greater
LCS	1 per ≤20 samples	70-130%R	Suspend analysis until source rectified; redigest and reanalyze affected samples	DESA or CLP Laboratory ICP Technician	70-130%R

^{*}except when the sample concentration is greater than 4 times the spike concentration, then disregard the recoveries; no data validation action taken



^{**} except when the sample and/or duplicate concentration is less than 5 times the CRQL, then \pm CRQL.

QAPP Worksheet #28i: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Aqueous

Analytical Group TAL – Total Mercury

Analytical Method/SOP Reference ISM01.3Cold Vapor Atomic Absorption (CVAA)

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Split Samples	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤50% RPD, ABS ≤ 5xCRQL when either result ≤ CRQL
Temperature Blank	1 per cooler	0 to 6 degrees C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Preparation Blank (PB)	1 per ≤20 samples	No analyte > CRQL	Suspend analysis; redigest and reanalyze	DESA or CLP Laboratory Technician	No analyte > CRQL
Laboratory Duplicate	1 per ≤20 samples	<u>+</u> 20% RPD*	Flag outliers	DESA or CLP Laboratory Technician	±_20% RPD
Spike Sample	1 per ≤20 samples	75 – 125 %R	Flag outliers	DESA or CLP Laboratory Technician	75 – 125 %R

^{*}Reference EPA Region 2 SOP No. HW-2c, Revision 15 - (include ABS criteria)



QAPP Worksheet #28j: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixAqueousAnalytical GroupMercury

Analytical Method/SOP Reference EPA 1631 – Atomic fluorescence spectroscopy

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Split Samples	1 per 20 samples	20% RPD	Notify PM and address in data quality report	CDM Smith ASC and PM	≤ 40% RPD (for results ≥ 5QL) or ABS≤QL
Temperature Blank	1 per cooler	0 to 6 degrees C	Note in laboratory narrative. CDM Smith will use more coolant; check packing procedure	CDM Smith FOS	≤ 10 degrees C for data validation
Preparation Blank (PB)	1 per 20 samples	No analyte > QL (greater of 0.4 ng or <0.1xsample)	Suspend analysis; redigest and reanalyze if sample<10*blank result		No analyte > QL
Laboratory duplicate	1 per 20 samples	Per laboratory SOP	Investigate and correct; Flag outliers; Note in case narrative. Multiple failures require re-distillation and reanalysis.	Laboratory Analyst	≤ 35% RPD if result >5QL
Ongoing Precision and Recovery Samples	1 per 20 samples or 12-hour shift	Per laboratory SOP	Check calculations and instruments, reanalyze affected samples. Report in case narrative.		70-130%R for OPR <20 RSD for IPR 75-125%R for IPR
MS/MSD	1 per 20 samples or with each group of field samples	Per laboratory SOP	Investigate matrix effects and note in data narrative.		70-130%R RPD ≤35% (30 per method)



QAPP Worksheet #28k: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Aqueous

Analytical Group TAL - Total Cyanide

Analytical Method/SOP Reference ISM01.3 or current method – Colorimeter or Spectrophotometer

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Split Samples	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤50% RPD, ABS ≤ 5xCRQL when either result ≤ CRQL
Temperature Blank	1 per cooler	0 to degrees 6 C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Preparation Blank (PB)	1 per ≤ 20 samples	No analyte > CRQL	Suspend analysis; redistill and reanalyze	DESA or CLP Laboratory Technician	No analyte > CRQL
Laboratory Duplicate	1 per ≤ 20 samples	<u>+</u> 20% RPD*	Flag outliers	DESA or CLP Laboratory Technician	<20% RPD
Spike Sample	1 per ≤ 20 samples	75 – 125 %R	Flag outliers	DESA or CLP Laboratory Technician	75 – 125 %R

^{*}Reference EPA Region 2 SOP No. HW-2c, Revision 15 - (include ABS criteria)



QAPP Worksheet #28l: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Soil/Sediment

Analytical Group TCL VOCs
Analytical Method/SOP Reference SOM01.2

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Sample Splits	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤100% RPD ABS ≤ 5xQL when either result < 2*CRQL
Temperature Blank	1 per cooler	0 to 6 °C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Method Blank	1 every 12 hours	No analyte > CRQL*	Suspend analysis unit source recertified	DESA/ CLP Laboratory GC/MS Technician	No analyte > CRQL*

^{*}With the exception of methylene chloride, 2-butanone & acetone which can be up to 2 times the CRQL. (EPA CLP National Functional Guidelines, Final, July 2007)



QAPP Worksheet #28l: Analytical Quality Control and Corrective Action (TCL VOCs Soils continued)

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
		Vinyl chloride-d3	68-122 %R			Vinyl chloride-d3	68-122 %R
		Chloroethane-d5	61-130 %R			Chloroethane-d5	61-130 %R
		1,1-Dichloroethene-d2	45-132 %R]	DESA/ CLP	1,1-Dichloroethene-d2	45-132 %R
		2-Butanone-d5	20-182 %R]		2-Butanone-d5	20-182 %R
		Chloroform-d	72-123 %R	limits (Section 11.3.4, Page D45 of SOM01.2)		Chloroform-d	72-123 %R
		1,2-Dichloroethane-d4	79-122 %R			1,2-Dichloroethane-d4	79-122 %R
Davitanatad		Benzene-d6	80-121 %R			Benzene-d6	80-121 %R
Deuterated Monitoring	all samples	1,2-Dichloropropane-d6	74-124 %R		Laboratory	1,2-Dichloropropane-d6	74-124 %R
Compounds	·	Toluene-d8	78-121 %R		GC/MS Technician	Toluene-d8	78-121 %R
		trans-1,3-Dichloropropene-d4	72-130 %R			trans-1,3-Dichloropropen e-d4	72-130 %R
		2-Hexanone-d5	17-184 %R			2-Hexanone-d5	17-184 %R
		1,4-Dioxane-d8	50-150 %R]		1,4-Dioxane-d8	50-150 %R
		1,1,2,2-Tetrachloroethane-d2	56-161 %R	-		1,1,2,2-Tetrachloroethane -d2	56-161 %R
		1,2-Dichlorobenzene-d4	70-131 %R			1,2-Dichlorobenzene-d4	70-131 %R
Internal Standards	all samples	50-200% of area, <u>+</u> 30 second retention time		Check calculations/ instruments reanalyze affected samples	DESA or CLP Laboratory GC/MS Technician	50-100% of area, <u>+</u> 30 seco	nd retention



QAPP Worksheet #28m: Analytical Quality Control and Corrective Action (TCL VOCs Soils continued)

Matrix Soil/Sediment

Analytical Group TCL VOCs
Analytical Method/SOP Reference SOM01.2

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criter	
		1,1-Dichloroethene	59-172 %R	Flag outliers		1,1-Dichloroethene	59-172 %R
	1 per 20	Trichloroethene	62-137 %R			Trichloroethene	62-137 %R
Matrix Spike (Not samples or Required) (if request	samples or less	Benzene	66-142 %R			Benzene	66-142 %R
	(ii requested)	Toluene	59-139 %R			Toluene	59-139 %R
		Chlorobenzene	60-133 %R		DESA/ CLP Laboratory GC/MS Technician	Chlorobenzene	60-133 %R
		1,1-Dichloroethene	0-22 %RPD			1,1-Dichloroethene	0-22 %RPD
Matrix Spike	1 per 20	Trichloroethene	0-24 %RPD			Trichloroethene	0-24 %RPD
Duplicate (Not	samples or less (if requested)	Benzene	0-21 %RPD	Flag outliers		Benzene	0-21 %RPD
Required)		Toluene	0-21 %RPD			Toluene	0-21 %RPD
		Chlorobenzene	0-21 %RPD			Chlorobenzene	0-21 %RPD



QAPP Worksheet #28n: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixSoil/SedimentAnalytical GroupTCL SVOCsAnalytical Method/SOP ReferenceSOM01.2

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Sample Splits	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤100% RPD ABS ≤ 5xCRQL when either results≤ 2*CRQL
Temperature Blank	1 per cooler	0 to 6 degrees C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Method Blank	1 per 20 samples or less whenever samples are extracted	No analyte > CRQL*	Suspend analysis and reanalyze blank and affected sample	DESA or CLP Laboratory GC/MS Technician	No analyte > CRQL*

^{*}with the exception of bis (2-Ethylhexyl) phthalate which can be up to 5 times the CRQL. (EPA CLP National Functional Guidelines, Final, July 2007)



QAPP Worksheet #28n: Analytical Quality Control and Corrective Action (TCL SVOC Soils continued)

QC Sample	Frequency /Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Data Quality Indicator	Measurement Performance Criteria	
				TCL SVOCs – Soil	Continued			
Deuterated		Phenol-d5	17-103 %R	Check	DESA or CLP	Accuracy	Phenol-d5	17-103 %R
Monitoring Compounds	:	Bis(2-chloroethyl)ethe r-d8	12-98 %R	instruments,	Laboratory GC/MS Technician		Bis(2-chloroethyl)ether-d8	12-98 %R
		2-Chlorophenol-d4	13-101 %R	reanalyze			2-Chlorophenol-d4	13-101 %R
		4-Methylphenol-d8	8-100 %R	DMCs may fail to meet recovery			4-Methylphenol-d8	8-100 %R
		Nitrobenzene-d5	16-103 %R				Nitrobenzene-d5	16-103 %R
		2-Nitrophenol-d4	16-104 %R				2-Nitrophenol-d4	16-104 %R
		2,4-Dichlorophenol-d3	23-104 %R				2,4-Dichlorophenol-d3	23-104 %R
all samples	4-Chloroaniline-d4	1-145 %R	11.3.4, Page			4-Chloroaniline-d4	1-145 %R	
	Dimethylphthalate-d6	43-111 %R				Dimethylphthalate-d6	43-111 %R	
		Acenaphthylene-d8	20-97 %R	D48/SVOC of			Acenaphthylene-d8	20-97 %R
		4-Nitrophenol-d4	16-166 %R	SOM01.2)			4-Nitrophenol-d4	16-166 %R
		Fluorene-d10	40-108 %R				Fluorene-d10	40-108 %R
		4,6-Dinitro-2-methylp henol-d2	1-121 %R				4,6-Dinitro-2-methylphen ol-d2	1-121 %R
		Anthracene-d10	22-98 %R				Anthracene-d10	22-98 %R
		Pyrene-d10	51-120 %R				Pyrene-d10	51-120 %R
		Benzo(a)pyrene-d12	43-111 %R				Benzo(a)pyrene-d12	43-111 %R
Internal Standards	all samples	50-200% of area, <u>+</u> 30 second retention time shift		Check calculations and instruments, reanalyze affected samples	DESA or CLP Laboratory GC/MS Technician	Accuracy	50-200% of area, <u>+</u> 30 secon time shift	nd retention



QAPP Worksheet #28n: Analytical Quality Control and Corrective Action (TCL SVOCs Soils continued)

Matrix Soil/Sediment

Analytical Group TCL SVOCs
Analytical Method/SOP Reference SOM01.2

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria	
		Phenol	26-90 %R			Phenol	26-90 %R
	1 per 20	2-Chlorophenol	25-102 %R			2-Chlorophenol	25-102 %R
	(if requested)	N-Nitroso-di-n-propylamine	41-126 %R	Flag outliers		N-Nitroso-di-n-propylamine	41-126 %R
nequired)	(requestes,	4-Chloro-3-methylphenol	26-103 %R			4-Chloro-3-methylphenol	26-103 %R
		Acenaphthene	31-137 %R		Lahoratory	Acenaphthene	31-137 %R
		Phenol	0-35 %RPD			Phenol	0-35 %RPD
Matrix Spike	1 per 20	2-Chlorophenol	0-50 %RPD		Technician	2-Chlorophenol	0-50 %RPD
Duplicate (Not	samples if requested (if	N-Nitroso-di-n-propylamine	0-38 %RPD	Flag outliers		N-Nitroso-di-n-propylamine	0-38 %RPD
Required)	requested)	4-Chloro-3-methylphenol	0-33 %RPD			4-Chloro-3-methylphenol	0-33 %RPD
		Acenaphthene	0-19 %RPD			Acenaphthene	0-19 %RPD



QAPP Worksheet #280: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Soil/Sediment
Analytical Group PCDD/PCDF
Analytical Method/SOP
EPA 1613B

Reference

QC Sample:	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Method Blank	1 per 20 samples	Per laboratory SOP	If samples non-detect or if lowest sample result is >10 times the blank-no action; otherwise redigest and reanalyze	Laboratory Analyst	No analyte > QL
Laboratory Duplicate	1 per 20 samples	Per laboratory SOP	Investigate and correct; Flag outliers	Laboratory Analyst	± 20% of mean if sample concentration >10x DL
Initial Precision and Recovery	Prior to sample analysis	Per laboratory SOP	Investigate and correct	Laboratory Analyst	Per method/laboratory SOP
Ongoing Precision and Recovery	1 per batch of 20 samples	Per laboratory SOP	Identify source of problem, make other adjustments; redigest if needed and reanalyze	Laboratory Analyst	Individual laboratory established limits per SOP
Sample splits	1 per 20 samples	None	Data assessor to inform PM if MPC is exceeded; address in data quality assessment	CDM Smith ASC	≤ 40% RPD (for results ≥ 5QL)
Temperature Blank	1 per cooler	0 to 6 degrees C	Note outlier in laboratory narrative. Inform CDM Smith of failure and need for additional coolant; check packing procedure	Laboratory Analyst	≤ 10 degrees C for data validation

Laboratory and SOPs are TBD. The laboratory will be assigned per FASTAC policy.



QAPP Worksheet #28q: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixSoil/SedimentAnalytical GroupTCL PesticidesAnalytical Method/SOP ReferenceSOM01.2

QC Sample	Frequency/ Number	Method, Acceptan	•	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Perfor	rmance Criteria
Sample Splits	1 per 20 samples	None		Notify PM and flag duplicate results	CDM Smith ASC and PM	≤100% RPD ABS ≤ 5xCRQL when a results ≤ 2*CRQL	
Temperature Blank	1 per cooler	0 to 6 degrees C		Laboratory will inform and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation	
Method Blank	1 per 20 samples or whenever samples extracted	No analyte > CRQL		Suspend analysis unit source recertified	DESA or CLP Laboratory GC/ECD Technician	No analyte > CRQL	
		gamma-BHC (Lindane)	46-127 %R		DESA or CLP Laboratory	gamma-BHC (Lindane)	46-127 %R
	4 20	Heptachlor	35-130 %R			Heptachlor	35-130 %R
Matrix Spike	1 per 20	Aldrin	34-132 %R	Flag outliers		Aldrin	34-132 %R
	samples	Dieldrin	31-134 %R		GC/ECD Technician	Dieldrin	31-134 %R
		Endrin	42-139 %R			Endrin	42-139 %R
		4,4-DDT	23-134 %R			4,4-DDT	23-134 %R
		gamma-BHC	0-50 %RPD			gamma-BHC	0-50 %RPD
		Heptachlor	0-31 %RPD			Heptachlor	0-31 %RPD
Matrix Spike	1 per 20	Aldrin	0-43 %RPD	Flag outliers	DESA or CLP Laboratory	Aldrin	0-43 %RPD
Duplicate	samples	Dieldrin	0-38 %RPD	Tiag outliers	GC/ECD Technician	Dieldrin	0-38 %RPD
		Endrin	0-45 %RPD			Endrin	0-45 %RPD
		4,4-DDT	0-50 %RPD			4,4-DDT	0-50 %RPD



QAPP Worksheet #28q: Analytical Quality Control and Corrective Action (continued)

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteri	
		gamma-BHC	50-120 %R			gamma-BHC	50-120 %R
	Heptachlor epoxide	50-150 %R			Heptachlor epoxide	50-150 %R	
	-111	Dieldrin	30-130 %R	Check calculations and	DESA or CLP Laboratory GC/ECD Technician	Dieldrin	30-130 %R
LCS	all samples	4,4'-DDE	50-150 %R	instruments, reanalyze affected samples		4,4'-DDE	50-150 %R
		Endrin	50-120 %R			Endrin	50-120 %R
		Endosulfan sulfate	50-120 %R			Endosulfan sulfate	50-120 %R
		gamma-Chlordane	30-130 %R			gamma-Chlordane	30-130 %R
Surrogate	all samples	30-150 %R		Check calculations and instruments, reanalyze affected samples	DESA or CLP Laboratory GC/ECD Technician	30-150 %R	

QAPP Worksheet #28r: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixSoil/SedimentAnalytical GroupTCL PCBsAnalytical Method/SOP ReferenceSOM01.2

QC Sample	Frequency/ Number	Method/SOP QC Acceptance Limits		Corrective Action	Person(s) Responsible for Corrective Action	Measurement I Crite	
Sample Splits	1 per 20 samples	None		Notify PM and flag duplicate results	CDM Smith ASC and PM	≤100% RPD ABS ≤ 5xCRQL when either results≤ 2xCRQL	
Temperature Blank	1 per cooler	0 to 6 degrees	С	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation	
Method Blank	1 per 20 samples or whenever samples extracted	No analyte > CRQL		Suspend analysis unit source recertified	DESA or CLP Laboratory GC/ECD Technician	No analyte > CRQL	
Matrix Spike	1 per 20 samples	Aroclor-1016	29-135 %R	Flag outliers	DESA or CLP Laboratory	Aroclor-1016	29-135 %R
	1 per 20 samples	Aroclor-1260	29-135 %R	Tiag Outliers	GC/ECD Technician	Aroclor-1260	29-135 %R
Matrix Spike	1 per 20 samples	Aroclor-1016	0-15 %RPD	Flag outliers	DESA or CLP Laboratory	Aroclor-1016	0-15 %RPD
Duplicate	1 per 20 samples	Aroclor-1260	0-20 %RPD	Flag Oddlers	GC/ECD Technician	Aroclor-1260	0-20 %RPD
LCS	all camples	Aroclor-1016	50-150 %R	Check calculations and instruments, reanalyze	DESA or CLP Laboratory	Aroclor-1016	50-150 %R
LCS all samples	an samples	Aroclor-1260	50-150 %R	affected samples	GC/ECD Technician	Aroclor-1260	50-150 %R
Surrogate	all samples	30-150%R		Check calculations and instruments, reanalyze affected samples	DESA or CLP Laboratory GC/ECD Technician	30-150%R	



QAPP Worksheet #28s: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixSoil/SedimentAnalytical GroupPCB CongenersAnalytical Method/SOP ReferenceEPA 1668A

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Method Blank	1 per 20 samples	< QL	If samples non-detect or if lowest sample result is >10 times the blank-no action; otherwise redigest and reanalyze	Laboratory Analyst	No analyte > QL
Laboratory Duplicate	1 per 20 samples	≤ 20% RPD; ±QL for samples <10x QL	Flag outliers	Laboratory Analyst	RPD ≤ 20%
Quality Control Sample	Periodically at least quarterly	70-130%R;	Check standards; recalibrate if required	Laboratory Analyst	70-130%R;
Calibration Verification Sample	Beginning of each 12-hour shift	70-130%R;	Adjust and/or recalibrate	Laboratory Analyst	70-130%R;
Initial Precision and Recovery	Prior to sample analysis	Per laboratory SOP	Investigate and correct	Laboratory Analyst	60-140%R ≤ 40% RSD
Ongoing Precision and Recovery	1 per batch of 20 samples	Per laboratory SOP	Identify source of problem, recalibrate if needed/ make other adjustments and reanalyze	Laboratory Analyst	Warning 70-130%R; Accept 50-150%R
Sample splits	1 per 20 samples	None	Data assessor to inform PM if MPC is exceeded; address in data quality assessment	CDM Smith ASC	≤ 40% RPD (for results ≥ 5QL)
Temperature Blank	1 per cooler	0 to 6 degrees C	Note outlier in laboratory narrative. Inform CDM Smith of failure and need for additional coolant; check packing procedure	Laboratory Analyst	≤ 10 degrees C for data validation

Laboratory and SOPs are TBD. The laboratory will be assigned per FASTAC policy.



QAPP Worksheet #28t: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixSoil/SedimentAnalytical GroupTAL Metals

Analytical Method/SOP Reference ISM01.3 or current method

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Sample Splits	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤100% RPD ABS ≤ 5xCRQL when either result ≤ 2xCRQL
Temperature Blank	1 per cooler	0 to 6 degrees C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Preparation Blank	1 per 20 samples	No constituent > CRQL	Suspend analysis until source rectified; re-digest and reanalyze affected samples		No constituent > CRQL
Spike	1 per 20 samples	75-125%R*	Flag outliers		75-125%R*
Laboratory Duplicate	1 per 20 samples	≤35% RPD**	Flag outliers	DESA or CLP Laboratory	≤ 35% RPD**
Post-Digestion Spike	after any analyte (except Ag and Hg) fails spike %R	75-125%R	Flag outliers	ICP-AES/ICP-MS Technician	75-125%R
Interference Check Sample [ICP Analysis Only]	beginning, end and periodically during run (2 times every 8 hours)	Within ± 2 times CRQL of true value or ± 20% of true value, whichever is greater	Check calculations and instruments, reanalyze affected samples		Within ± 2 times CRQL of true value or ± 20% of true value, whichever is greater
LCS	1 per 20 samples	Control limits established by EPA*	Suspend analysis rectify source; re-digest and reanalyze affected samples		Control limits established by EPA*

^{*}except when the sample concentration is greater than 4 times the spike concentration, then disregard the recoveries; no data validation action taken.

^{***} If the EPA LCS is unavailable, other EPA QC samples or other certified materials may be used. In such cases, control limits for the LCS must be documented and provided.



^{**}Reference EPA Region 2 SOP No. HW-2, Revision 13/Evaluation of Metals Data for CLP - (include ABS criteria) except when the sample and/or duplicate concentration is less than 5 times the CRQL.

QAPP Worksheet #28u: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Soil/Sediment

Analytical Group TAL –Total Mercury

Analytical Method/SOP ISM01.3, or current method – Cold Vapor Atomic Absorption

Reference (CVAA)

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Sample Splits	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤100% RPD ABS ≤ 5xCRQL when either result ≤ 2xCRQL
Temperature Blank	1 per cooler	0 to 6 degrees C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Preparation Blank (PB)	1 per ≤20 samples	No analyte > CRQL	Suspend analysis; redigest and reanalyze	DESA or CLP Laboratory Technician	No analyte > CRQL
Laboratory Duplicate	1 per ≤20 samples	<u>≤</u> 20% RPD	Flag outliers	DESA or CLP Laboratory Technician	≤ 35% RPD
Spike Sample	1 per ≤20 samples	75 – 125 %R	Flag outliers	DESA or CLP Laboratory Technician	75 – 125 %R



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QAPP Worksheet #28v: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

MatrixSoil/SedimentAnalytical GroupTotal Mercury

Analytical Method/SOP
Reference EPA 1631

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Sample splits and field duplicates	1 per 20 samples	None	Data assessor to inform PM if MPC is exceeded; address in data quality assessment	CDM Smith ASC	≤ 40% RPD (for results ≥ 5QL)
Preparation Blank	3 per 20 samples or batch	Per laboratory SOP	Reanalyze. Suspend analysis until source rectified; re-distill and reanalyze affected samples if results are <10 times the blank	Laboratory Analyst	No result > 5MDL
Laboratory duplicate	1 per 20 samples	Per laboratory SOP	Investigate and correct; Flag outliers; Note in case narrative. Multiple failures require re-distillation and reanalysis.	Laboratory Analyst	≤ 35% RPD if result >5CRQL
Ongoing Precision and Recovery Samples	1 per 20 samples or with each group of field samples	Per laboratory SOP	Check calculations and instruments, reanalyze affected samples. Report in case narrative.	Laboratory Analyst	70-130%R for OPR <20 RSD for IPR 75-125%R for CRM/IPR
MS/MSD	1 per 20 samples or with each group of field samples	Per laboratory SOP	Investigate matrix effects and note in data narrative.	Laboratory Analyst Laboratory Analyst	70-130%R RPD ≤35% (30 per method)

Laboratory and SOPs are TBD. The laboratory will be assigned per FASTAC policy.



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QAPP Worksheet #28w: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

Matrix Soil/Sediment

Analytical Group TAL – Total Cyanide

Analytical Method/SOP Reference ISM01.3 or current method—Colorimeter or Spectrophotometer

QC Sample	Frequency/Number	Method/SOP QC Acceptance Limits	Corrective Action	Person(s) Responsible for Corrective Action	Measurement Performance Criteria
Sample Splits	1 per 20 samples	None	Notify PM and flag duplicate results	CDM Smith ASC and PM	≤100% RPD ABS ≤ 5xCRQL when either result ≤ 2*CRQL
Temperature Blank	1 per cooler	0 to 6 degrees C	Laboratory will inform RSCC/CDM Smith and note in data narrative. CDM Smith will check packing procedure and increase coolant	Laboratory Analyst and CDM Smith FOS	≤ 10 degrees C for data validation
Preparation Blank (PB)	1 per ≤20 samples	No analyte > CRQL	Suspend analysis; redigest and reanalyze	DESA or CLP Laboratory Technician	No analyte > CRQL
Laboratory Duplicate	1 per ≤20 samples	<u>±</u> 20% RPD	Flag outliers	DESA or CLP Laboratory Technician	< 35% RPD
Spike Sample	1 per ≤20 samples	75 – 125 %R	Flag outliers	DESA or CLP Laboratory Technician	75 – 125 %R



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QAPP Worksheet #28x: Analytical Quality Control and Corrective Action (UFP-QAPP Manual Section 3.4 and Tables 4, 5, and 6) (EPA 2106-G-05 Section 2.3.5)

PROCEDURE FOR FIELD BLANK COLLECTION/ FIELD RINSATE BLANK COLLECTION (Continued)

Cooler Temperature Indicators

One cooler temperature indicator or "temperature blank" will be placed in each cooler containing samples (solid and aqueous) being sent to the laboratory for analysis. The temperature blank will consist of a sample container filled with non-preserved water (potable or distilled). The container will be labeled "COOLER TEMPERATURE INDICATOR" and dated.

Matrix Spikes

Matrix spikes (MS) are laboratory QC samples drawn from excess volumes of existing samples to demonstrate the accuracy of laboratory analysis. In accordance with EPA Region 2, matrix spikes will be designated on environmental samples at a rate of one per sample delivery group (SDG). This designation will be noted on the sample container labels and the sample paperwork. An SDG is defined as one of the following:

- 1. All samples of an analytical case if the sample number is less than 20 (including environmental duplicates and QC blanks) and if sampling is completed within 7 calendar days.
- 2. Each group of 20 samples within an analytical case (including environmental duplicates, but excluding QC blanks) if the number is greater than 20.
- 3. Each 7-day calendar day period during which samples within an analytical case are received. This period begins with the receipt of the first sample in the SDG.



QAPP Worksheet #29: Project Documents and Records (UFP-QAPP Manual Section 3.5.1) (EPA 2106-G-05 Section 2.2.8)

	Sample Collection and Field Records						
Record	Generation (CDM Smith)	Verification (CDM Smith)	Storage location/archival				
Air Bills	FOS	FOS or Designee	Project File				
ANSETS	CDM Smith Project Manager	ASC	Project File				
Audit plans and reports	Auditor	QA Manager or Designee	Project File				
Corrective Action Reports	Project Manager	PM Designee	Project File				
Correspondence	Project Manager	Project Manager or Designee	Project File				
Daily QC Reports	FOS or Designee	PM Designee	Project File				
Daily Sign-In Sheet	FOS or Designee	PM	Project File				
Data usability assessment report	ASC or Designee	Chemist	Project File				
Data validation report	Data validator	Chemist	Project File				
Data verification checklists	FOS	ASC	Project File				
Deviations – Field Change Request	FOS	PM	Project File				
Field logbook or data collection sheets	FOS	FOS	Project File				

Note: field forms see Appendix D.



QAPP Worksheet #29: Project Documents and Records (UFP-QAPP Manual Section 3.5.1) (EPA 2106-G-05 Section 2.2.8)

Project Assessments					
Record	Generation	Verification	Storage location/archival		
Photographic log	FOS or Designee	Task Manager or Designee	Project File		
Sample Tracking Forms	Sample Manager or Designee	FOS or Designee	Project File		
Scribe Chain-of-Custody Forms	Sample Manager or Designee	FOS or Designee	Project File		
Self-Assessment Checklist	Site Manager or Designee	QA Specialist	Project File		
Subcontractor Laboratory Sample Tracking Log	Sample Manager or Designee	FOS or Designee	Project File		
	Laboratory	ecords			
Record	Generation	Verification	Storage location/archival		
Bid Sheets, scopes of work	PM or Designee	Technical Reviewer and Procurement Specialist	Procurement File		
Subcontract Laboratory certifications	Laboratory QA Officer	Chemist or QA Specialist	Procurement File		
Subcontract Laboratory QA Plans	Laboratory QA Officer	Chemist or QA Specialist	Procurement File		
SOPs	Laboratory QA Officer	Chemist or QA Specialist	Procurement File		

Note: field forms see Appendix D.



QAPP Worksheet #29: Project Documents and Records (UFP-QAPP Manual Section 3.5.1) (EPA 2106-G-05 Section 2.2.8)

	Laboratory Data Deliverables				
Record	SOM01.2- VOCS, SVOC, Pesticides, PCBs	ISM01.3 -TAL Metals	PCB Congeners and Dioxin/Furans	Trace Mercury	
Narrative	Х	Х	Х	Х	
COC	Х	Х	Х	Х	
Summary Results	Х	Х	Х	Х	
Analytical sample results	Х	Х	Х	Х	
QC Results	Х	Х	Х	Х	
Chromatograms	Х	NA	Х	NA	
Sample Preparation Log	Х	Х	Х	Х	
Sample Run Log	Х	Х	Х	Х	
Tentatively Identified Compounds (TICs)	NA	NA	NA	NA	
Raw Data	Х	Х	Х	Х	

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QAPP Worksheet #30: COMBINED WITH WORKSHEET #19



QAPP Worksheet #31, 32 & 33: Assessments and Corrective Action (UFP-QAPP Manual Sections 4.1.1 and 4.1.2) (EPA 2106-G-05 Section 2.4 and 2.5.5)

Assessment Type	Number/ Frequency	Organization	Responsible Party	Assessment Deliverable and Due Dates	Party to Identify and Implement Corrective Actions	Person(s) Responsible for Monitoring Effectiveness of Corrective Actions
					Title and Organ	nizational Affiliation
Project Readiness Review	Prior to field work	CDM Smith	FOS	Immediately; to within 24 hours of review	PM, CDM Smith	Paul Hagerman (PM), CDM Smith
Sample Collection and Documentation	Once	CDM Smith	FOS	Email within 24 hours	PM, CDM Smith	Jeniffer Oxford (QAS) or field auditor, CDM Smith
Health and Safety	Once if warranted	CDM Smith	FOS and PM, CDM Smith	Memorandum & Checklist (Notify by phone immediately. Report 1 week after audit)	PM, CDM Smith	Shawn Oliveira, H&S Manager or designee, SSHO, CDM Smith
QAPP	Annually	CDM Smith	Approved CDM Smith QA Staff or QA Coordinator	E-mail / FCR if required.	PM, CDM Smith	Paul Hagerman (PM), CDM Smith
Data Review	Once	CDM Smith	Vanessa Macwan (ASC) or designee,	Memorandum based on project requirements	Project Chemist, FOS, or PM depending on nature of issue	Paul Hagerman (PM), CDM Smith

Notes:

- 1. The CDM Smith QA Manager (QAM) will determine if an office audit is required. If CDM Smith PM requests self-assessments in lieu of the project audit, the QAM will review and approve or reject the self-assessments being considered.
- 2. Office audits are performed by trained and approved QA Staff members.
- 3. Findings and deviations from plans will require corrective actions which will be documented and discussed appropriately. The USACE PM and EPA RPM will be notified by CDM Smith PM.



QAPP Worksheet #34: Data Verification and Validation Inputs (UFP-QAPP Manual Section 5.2.1 and Table 9) (EPA 2106-G-05 Section 2.5.1)

Item	Input	Description	Verification (completeness)	Validation (conformance to specifications)
		Planning Documents/Records		
1	QAPP		Х	Х
2	Contractor Quality Control Plan (CQCP)	All planning documents will be available to reviewers to allow reconciliation with	Х	
3	Field TSOPs, The Group's QAPP and SAP	planned activities and objectives.	Х	Х
4	Laboratory SOPs		Х	Х
		Field Records		
5	Field logbooks	Field notes will be prepared daily by the Field Team and will be complete, appropriate to the project tasks, and legible. The FOS will review logbooks and	Х	Х
6	Equipment calibration records	records for accuracy and completeness. Upon completion of field work, logbooks and records will be placed in the project files. Field reports will be verified to ensure correct reporting of information. Review will be conducted prior to completion of each report.	Х	Х
7	сос	Sample manager, FOS or designee will review the COC forms against the samples packed in the each cooler prior to shipment. COCs will be sent with the samples to the laboratory and copies retained for the Trip Report and project files. The data validator will be review upon completion of analytical activities and verified against the laboratory report.	Х	Х
8	Sampling Trip Reports	FOS or designee; Laboratory coordinator will review these for each case of field sampling for which samples are sent to a CLP laboratory. Information will be reviewed against the COC forms, and potential discrepancies will be discussed with field personnel to verify locations, dates, etc.	Х	Х
9	Sampling Figures/ Diagrams/Surveys	Data user will review during evaluation and completion of data report.	Х	Х
12	Correspondence	Relevant correspondence will be used to reconcile field records and data.	Х	Х



QAPP Worksheet #34: Data Verification and Validation Inputs (UFP-QAPP Manual Section 5.2.1 and Table 9) (EPA 2106-G-05 Section 2.5.1)

Item	Input	Description	Verification (completeness)	Validation (conformance to specifications)
13	Field Change Requests	ASC and data evaluator will review during completion of each data usability assessment/measurement report.	Х	Х
		Analytical Data Package		
14	Laboratory analytical data packages	Laboratory analyst and QA officer will review/verify internally the completeness and technical accuracy of data prior to submittal. All laboratory data will be verified by the laboratory performing the analysis prior to submittal. EPA DV contractor-data validator or CDM Smith data validator will review data packages for content and sample information upon receipt. Data packages will be evaluated for completeness and compliance. Table 9 of the IDQTF UFP-QAPP shows items for compliance review.	х	Х
15	Communication Records	Relevant correspondence will be used to reconcile analytical data.	Х	Х
16	Electronic Data Deliverables (EDDs) fields	Data Manager will determine whether required EQuIS compatible EDD fields and format were provided.	х	х
17	Outputs of the EQuIS database	Project task leader and team will compile the project data results in a sample project report. Data tables, figures and reported entries will be reviewed/ verified against hardcopy information or EQuIS output.	Х	х
18	Data validation and audit reports, QAPP, and FCRs	Data assessor will prepare the project data quality and usability assessment report. The data will be evaluated against project DQOs and measurement performance criteria, such as completeness. Evaluate whether field sampling procedures were followed with respect to equipment and proper sampling support.	Х	Х

QAPP Worksheet #35: Data Verification Procedures (UFP-QAPP Manual Section 5.2.2) (EPA 2106-G-05 Section 2.5.1)

Requirement Documents	Records Reviewed	Process Description	Responsible Person /Organization
QAPP, TSOP 4-1	Field logbook	Verify that records are present and complete for each day of field activities. Verify that all planned samples including field QC samples were collected and that sample collection locations are documented. Verify that meteorological data were provided for each day of field activities. Verify that changes/exceptions are documented and were reported in accordance with requirements. Verify that any required field monitoring was performed and results are documented.	Daily - FOS At conclusion of field activities - Project QC staff
SOPs	Field logbook and FCRs	Ensure that the sampling methods/procedures outlined in QAPP were followed, and that any deviations were noted/approved. Determine potential impacts from noted/approved deviations, in regard to PQOs.	FOS
QAPP, TSOP 1-2	Chain-of-custody forms	Verify the completeness of chain-of-custody records. Examine entries for consistency with the field logbook. Check that appropriate methods and sample preservation have been recorded. Verify that the required volume of sample has been collected and that sufficient sample volume is available for QC samples (e.g., MS/MSD). Verify that all required signatures and dates are present. Check for transcription errors.	Daily - FOS At conclusion of field activities - Project Chemist or Data Assessor
QAPP, TSOP 1-2	сос	Examine traceability of data from sample collection to generation of project reported data. Provides sampling dates and time; verification of sample ID; and QC sample information.	At conclusion of field activities - Project QC staff (data coordinator, data validator)
QAPP	Laboratory data package	Examine packages against QAPP and laboratory contract requirements, and against COC forms (e.g., holding times, sample handling, analytical methods, sample identification, data qualifiers, QC samples, etc.). Determine potential impacts from noted/approved deviations, in regard to PQOs.	ESAT Data Validation Personnel, EPA Region 2 or CDM Smith Data validator



QAPP Worksheet #35: Data Verification Procedures (UFP-QAPP Manual Section 5.2.2) (EPA 2106-G-05 Section 2.5.1)

Requirement Documents	Records Reviewed	Process Description	Responsible Person /Organization
	Laboratory Deliverable	Verify that the laboratory deliverable contains all records specified in the subcontract SOW. Check sample receipt records to ensure sample condition upon receipt was noted, and any missing/broken sample containers were noted and reported according to plan. Compare the data package with the COCs to verify that results were provided for all collected samples. Review the narrative to ensure all QC exceptions are described. Check for evidence that any required notifications were provided to project personnel as specified in the QAPP. Verify that necessary signatures and dates are present.	Before release – Laboratory QAM Upon receipt - Project Chemist or Data Validator [ESAT or CDM Smith Data Validation Personnel or ASC]
QAPP	Audit Reports, Corrective Action Reports	Verify that all planned audits were conducted. Examine audit reports. For any deficiencies noted, verify that corrective action was implemented according to plan.	Contract QAS
	Field duplicates	Compare results of field duplicate (or replicate) analyses with RPD criteria.	
	Methods	Verify that records support implementation of the SOP - sampling and analysis.	
	Data Narrative	Determine deviations from methods and contract and the impact.	
	Audit Report	Confirm reports are used to validate compliance of field sampling, handling and analysis activities with the QAPP.	CDM Smith ASC, Data Validator or
	Project Quantitation Limit	Verify achievement of PQLG as established in the QAPP and that the laboratory successfully analyzed a standard at the QL.	Data Assessor
	Field and Laboratory data and QC report	A summary of all QC samples and results will be verified for measurement performance criteria, completeness, and 10 percent verified to field and laboratory data reports from vendors. A report on meeting the established criteria shall be prepared within 30 days of receipt.	



QAPP Worksheet #36: Data Validation Procedures (UFP-QAPP Manual Section 5.2.2) (EPA 2106-G-05 Section 2.5.1)

Validation Code and Label Identifier Table

Validation Code*	Validation Label	Description/Referen	ice
S2BVM	Stage 2b Validation Manual	Stage 2B Validation - Verification and validation based on completeness and compliance checks of sample receipt conditions and BOTH sample-related and instrument-related QC results.	EPA 540-R-08-005
S4VEM	Stage 4 Validation Electronic and Manual	Stage 4 Validation - Verification and validation based on completeness and compliance checks of sample receipt conditions, both sample-related and instrument-related QC results, AND recalculation checks.	
NV	Not Validated		
S3VEM	Stage 3 Validation Electronic and Manual	Stage 3 Validation - Verification and validation based on completeness and compliance checks of sample receipt conditions, both sample-related and instrument-related QC results, AND recalculation checks.	
S2bVEM	Stage 2b Validation Electronic and Manual	Stage 2B Validation - Verification and validation based on completeness and compliance checks of sample receipt conditions and BOTH sample-related and instrument-related QC results.	

Note:

The following data qualifiers will be applied during data validation by a third party. Potential impacts on project data quality objectives will be discussed in the data validation report.

- NM Measurement Performance Criteria contained in WS 12 were not met.
- J The result is an estimated value. The nature of the bias will be discussed in the data validation report.
- E Erroneous result (e.g., improper calculation, peak integration, etc.)
- R- rejected data



QAPP Worksheet #36: Data Validation Procedures (UFP-QAPP Manual Section 5.2.2) (EPA 2106-G-05 Section 2.5.1)

Analytical Group/Method	Data deliverable requirements	Analytical specifications	Measurement performance criteria	Percent of data packages to be validated ¹	Percent raw data review/% results to recalculate	Validation Procedure ³	Validation code	Electronic validation program/versi on	Data Validator
			F	ASTAC Tiers 1 and	d 2 (DESA or CLP)				
VOCs	SEDD Stage 3	SOM01.2				SOP HW-34, Rev 3, DESA Worksheet #35 or NFG	S3VEM	EXES	ESAT DV Staff, or DESA
SVOCs + SIM	SEDD Stage 3	SOM01.2				SOP HW-35, Rev 2, DESA Worksheet #35	S3VEM	EXES	ESAT DV Staff, or DESA
TAL Metals, Mercury (ICP-AES)	SEDD Stage 2B	ISM01.3	Appendix A	100%	100%/10%	SOP HW-2 a, Rev 15 or NFG	S2BVEM	EXES	
TAL Metals, Mercury (ICP-MS)	SEDD Stage 2B	ISM01.3				SOP HW-2 b, Rev 15 or NFG	S2BVEM	EXES	ESAT/DESA or CDM Smith DV Staff
Cyanide	SEDD Stage 2B	ISM01.3				SOP HW-2 c, Rev 15 or NFG	S2BVEM	EXES	2 1 3td11

			Tie	r 4 (CDM Smith Sub	contract Laboratory				
Trace mercury	EQuIS Region 2 compliant EDD	FPA 1631F	Appendix A	100%	100%/1 SDG	NFG modified by WS #12, 28, 15, 19 and 24	S2BVM	NA ⁴	CDM Smith ASC/ designee
Dioxin/Furans	SEDD Stage 3	WS 28, & EPA 1613B (Isotope dilution)	Appendix A	100%	100%/1 SDG	EPA SOP HW-25, Revision 3 or NFG	S3VEM	EXES	CDM Smith
PCB Congeners + homologs	SEDD Stage 3	EPA 1668				SOP HW-46 or NFG	S3VEM	EXES	ESAT DV Staff, or CDM Smith



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QAPP Worksheet #37: Data Usability Assessment (UFP-QAPP Manual Section 5.2.3 including Table 12) (EPA 2106-G-05 Section 2.5.2, 2.5.3, and 2.5.4)

The Data Comparability Report in lieu of data usability assessment will be prepared by CDM Smith. Paul Hagerman, CDM Smith Project Manager, will be responsible for its content and for assigning work to the CDM Smith personnel who will be supporting this assessment. The Data Comparability Report presents the overall comparison of the split sample data and the Groups parent sample data. Data comparison will be conducted on parameters that were analyzed and detected by both sample pairs. Data quality will be evaluated in the data validation reports.

Split samples for the selected parameters will be compared using the following criteria:

- Average ratio criteria: average ratio of the Groups s to CDM Smith split sample. Ration criteria of 30% will be used to evaluate the data pairs.
- Percent difference criteria: percent difference of the Respondents to CDM Smith split sample. Percent difference of 50% will be used to evaluate the data pairs.
- Statistical test criteria: Paired Prentice-Wilcoxon test will be employed at significance level (p-value) of 0.05. A p-value greater than or equal to 0.05 indicating that there is no significant statistical difference between the data pairs.

The sample parameter being evaluated is considered comparable if at least two of the three criteria are met.



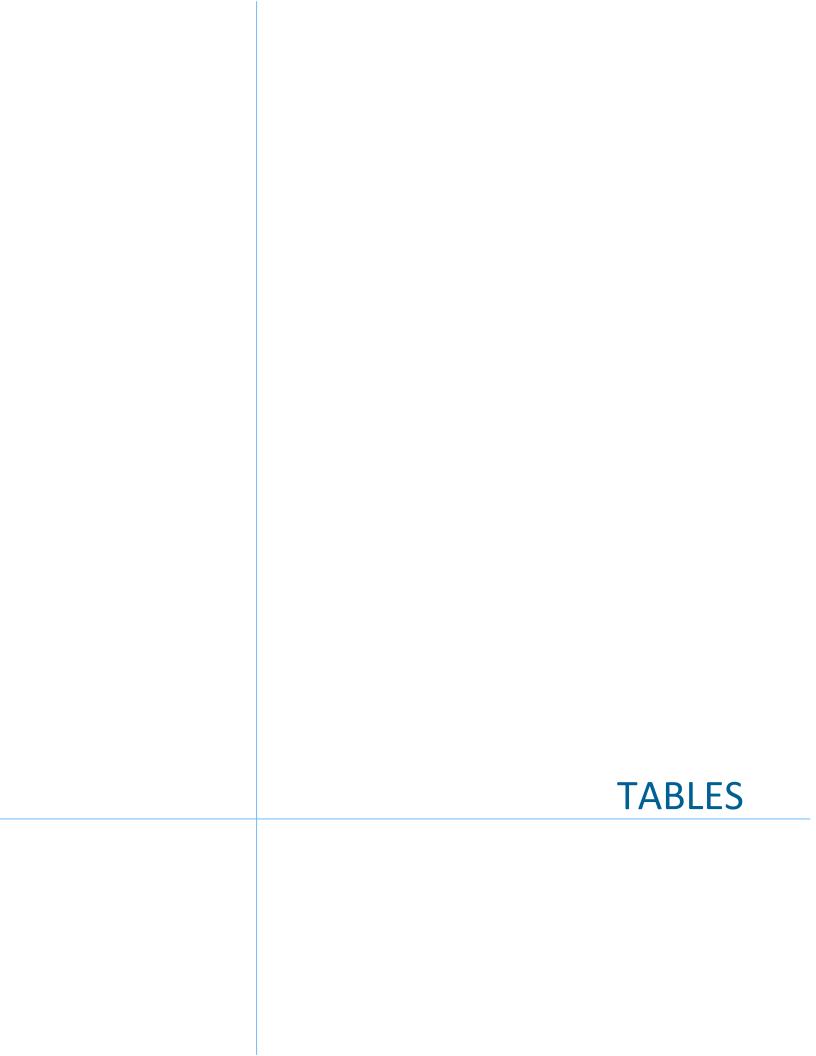


Table 1

Sample Locations, Depths, and Analyses
Data Gaps Sampling and Analysis Plan
Rolling Knolls Landfill Superfund Site
Chatham, New Jersey

								PF	RP La	bora	tor	v A	nalys	es				
Sample ID	Sample Media	Depth Interval (Feet)	Sample Collection Method	VOCs	SVOCs	SVOCs - SIM	PCBs (as Aroclors)	Pesticides	TAL Metals and Cvanide		Sul		TAL Metals and Cyanide (unfiltered)	TAL Metals and Cyanide (filtered)	Low-Level Mercury	Hardness	pH, TOC, Grain Size	Notes
Soil Samples	5																	
SS-125	Soil	0.0-1.0	Macrocore			X	X					X						
SS-126	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-125
SS-127	Soil	0.0-1.0	Macrocore			Χ	X			X		X						
SS-128	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-127
SS-129	Soil	0.0-1.0	Macrocore			Χ	Χ			X		X						
SS-130	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-129
SS-131	Soil	0.0-1.0	Macrocore			Χ	Χ					X						
SS-132	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-131
SS-133	Soil	0.0-1.0	Macrocore			Χ	X					X						
SS-134	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-133
SS-135	Soil	0.0-1.0	Macrocore			Χ	X					X						
SS-136	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-135
SS-137	Soil	0.0-1.0	Macrocore			Χ	Χ			X		Χ						
SS-138	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-137
SS-139	Soil	0.0-1.0	Macrocore			X	X			X		X						
SS-140	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-139
SS-141	Soil	0.0-1.0	Macrocore			X	X					X						
SS-142	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-141
SS-143	Soil	0.0-1.0	Macrocore			Χ	Χ					X						

Table 1 Sample Locations, Depths, and Analyses

Data Gaps Sampling and Analysis Plan Rolling Knolls Landfill Superfund Site Chatham, New Jersey

									Lab	orate	ory	Ana	alyses					
Sample ID	Sample Media	Depth Interval (Feet)	Sample Collection Method	VOCs	SVOCs	SVOCs - SIM	PCBs (as Aroclors)	Pesticides	TAL Metals and	PCB Congeners,	Dioxins, Furans	Full TCL/TAL	TAL Metals and Cyanide (unfiltered)	TAL Metals and Cyanide (filtered)	Low-Level Mercury	Hardness	pH, TOC, Grain Size	Notes
SS-144	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-143
SS-145	Soil	0.0-1.0	Macrocore			X	Χ					Χ						
SS-146	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-145
SS-147	Soil	0.0-1.0	Macrocore			X	Χ					Χ						
SS-148	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-147
SS-149	Soil	0.0-1.0	Macrocore			X	Χ					Χ						
SS-150	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-149
SS-151	Soil	0.0-1.0	Macrocore			X	Χ					Χ						
SS-152	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-151
SS-153	Soil	0.0-1.0	Macrocore			X	Χ					Χ						
SS-154	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-153
SS-155	Soil	0.0-1.0	Macrocore			X	Χ					Χ						
SS-156	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-155
SS-157	Soil	0.0-1.0	Macrocore			X	Χ					Χ						
SS-158	Soil	0.0-1.0	Macrocore															*Contingent on sample SS-157
SS-159	Soil	0.0-1.0	Macrocore			X	Χ			>	(Χ						
SS-160	Soil	0.0-1.0	Macrocore			X	Χ			>	(Χ						
SS-161	Soil	0.0-1.0	Macrocore			X	Χ			>	(Χ						Congeners, dioxins, and furans will be analyzed on up to 2 samples if PCBs are
SS-162	Soil	0.0-1.0	Macrocore			X	Χ			>	(Χ						detected in the TAL analysis.
SS-163	Soil	0.0-1.0	Macrocore			X	Χ			>	(Χ						·
SS-164	Soil	0.0-1.0	Macrocore			X	Χ			>	(Χ						

Table 1 Sample Locations, Depths, and Analyses

Data Gaps Sampling and Analysis Plan Rolling Knolls Landfill Superfund Site Chatham, New Jersey

									Labo	ratory	/ Ana	alyses					1
Sample ID	Sample Media Ionitoring Wells	Depth Interval (Feet)	Sample Collection Method	VOCs	SVOCs	SVOCs - SIM	PCBs (as Aroclors)	Pesticides	TAL Metals and Cyanide	PCB Congeners, Dioxins, Furans		TAL Metals and Cyanide (unfiltered)	TAL Metals and Cyanide (filtered)	Low-Level Mercury	Hardness	pH, TOC, Grain Size	Notes
TWP-1	Groundwater	TBD	Macrocore			Χ					Χ		Χ				
TWP-2	Groundwater	TBD	Macrocore			Х					X		X				
TWP-3	Groundwater	TBD	Macrocore			X					X		Х				
TWP-4	Groundwater	TBD	Macrocore			Χ					Χ		Χ				
TWP-5	Groundwater	TBD	Macrocore			X					Χ		Х				
TWP-6	Groundwater	TBD	Macrocore			Χ					Χ		Χ				
TWP-7	Groundwater	TBD	Macrocore			Χ					Χ		Χ				
TWP-8	Groundwater	TBD	Macrocore			X					Χ		Χ				
TWP-9	Groundwater	TBD	Macrocore			X					Χ		Χ				
Permanent N	Ionitoring Wells	(Existing)															
MW-1	Groundwater	14.5	Low flow			X					X		Χ				
MW-2	Groundwater	12.5	Low flow			X					X		Χ				
MW-3	Groundwater	12.5	Low flow			X					X		Χ				
MW-4	Groundwater	12.5	Low flow			X					X		Χ				
MW-5	Groundwater	12.5	Low flow			X					X		Χ				
MW-6	Groundwater	12.5	Low flow			X					X		Χ				
MW-7	Groundwater	12.5	Low flow			X					X		Χ				
MW-8	Groundwater	12.5	Low flow			X					X		Χ				
MW-9	Groundwater	12.5	Low flow			X					X		Χ				
MW-10	Groundwater	12.5	Low flow			X					X		Χ				
X-1	Groundwater	18	Low flow			X					X		Χ				

Table 1

Sample Locations, Depths, and Analyses
Data Gaps Sampling and Analysis Plan
Rolling Knolls Landfill Superfund Site
Chatham, New Jersey

				1					Laba		Λ						1
				ļ,					Labo	ratory	Ana	alyses					
Sample ID	Sample Media	Depth Interval (Feet)	Sample Collection Method	VOCs	SVOCs	SVOCs - SIM	PCBs (as Aroclors)	Pesticides	TAL Metals and Cyanide	PCB Congeners, Dioxins, Furans	Full TCL/TAL	TAL Metals and Cyanide (unfiltered)	TAL Metals and Cyanide (filtered)	Low-Level Mercury	Hardness	pH, TOC, Grain Size	Notes
X-2	Groundwater	20	Low flow			X					X		Χ				
X-3	Groundwater	23	Low flow			X					X		Χ				
X-4	Groundwater	15.5	Low flow			X					X		Χ				
X-5	Groundwater	13.2	Low flow			X					X		Х				This well was not sampled during previous sampling activities due to a
X-6	Groundwater	13	Low flow			X					X		Χ				
X-7	Groundwater	8.7	Low flow			X					X		Х				This well was not sampled during previous sampling activities due to a
Permanent M	Monitoring Wells	(Propose	d)														
MW-11	Groundwater	TBD	Low flow			X					X		Χ				
MW-12	Groundwater	TBD	Low flow			X					X		X				
MW-13	Groundwater	TBD	Low flow			X					X		X				
MW-14	Groundwater	TBD	Low flow			X					Χ		X				
MW-15	Groundwater	TBD	Low flow			X					Χ		X				
MW-16	Groundwater	TBD	Low flow			Χ					Χ		X				
MW-17	Groundwater	TBD	Low flow			X					X		X				
Pore Water S	Samples																
PW-1	Pore Water	0.0-0.5	PDB			Χ					X		X				
PW-2	Pore Water	0.0-0.5	PDB			Χ					Χ		Χ				
Surface Wat	er Samples																
SW-34	Surface Water	TBD	Teflon-lined bailer/direct dip			X					Х		Х	Х	Х		-

Table 1 Sample Locations, Depths, and Analyses

Data Gaps Sampling and Analysis Plan Rolling Knolls Landfill Superfund Site Chatham, New Jersey

									Labo	ratory	Ana	alyses					
Sample ID	Sample Media	Depth Interval (Feet)	Sample Collection Method	VOCs	SVOCs	SVOCs - SIM	PCBs (as Aroclors)	Pesticides	TAL Metals and Cyanide	PCB Congeners, Dioxins, Furans	Full TCL/TAL	TAL Metals and Cyanide (unfiltered)	TAL Metals and Cyanide (filtered)	Low-Level Mercury	Hardness	pH, TOC, Grain Size	Notes
SW-35	Surface Water	TBD	Teflon-lined bailer/direct dip			Х					Х		Χ	Χ	Х		
SW-36	Surface Water	TBD	Teflon-lined bailer/direct dip			Х					Х		Х	Χ	Χ		
SW-37	Surface Water	TBD	Teflon-lined bailer/direct dip			Х					Х		Х	Χ	Х		
SW-38	Surface Water	TBD	Teflon-lined bailer/direct dip			Х					Х		Х	Χ	Х		
SW-39	Surface Water	TBD	Teflon-lined bailer/direct dip			Х					Х		Х	Χ	Х		
SW-40	Surface Water	TBD	Teflon-lined bailer/direct dip			X					Х		Х	X	X		
SW-41	Surface Water	TBD	Teflon-lined bailer/direct dip			Х					Х		Х	Χ	Х		
SW-42	Surface Water	TBD	Teflon-lined bailer/direct dip			X					Х		Х	X	Χ		
SW-43	Surface Water	TBD	Teflon-lined bailer/direct dip			X					Х		Х	X	Χ		
SW-44	Surface Water	TBD	Teflon-lined bailer/direct dip			Х					Х		Х	X	X		

Table 1

Sample Locations, Depths, and Analyses
Data Gaps Sampling and Analysis Plan
Rolling Knolls Landfill Superfund Site
Chatham, New Jersey

								Labo	ratory	Ana	alyses					
Sample ID	Sample Media	Depth Interval (Feet)	Sample Collection Method	VOCs	SVOCs - SIM	PCBs (as Aroclors)	Pesticides	TAL Metals and Cyanide	PCB Congeners, Dioxins, Furans	Full TCL/TAL	TAL Metals and Cyanide (unfiltered)	TAL Metals and Cyanide (filtered)	Low-Level Mercury	Hardness	pH, TOC, Grain Size	Notes
Sediment Sa	ımples															
SD-34	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-35	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-36	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-37	Sediment	0.0 - 1.0	Grab sample – Encore sampler		X					Х					Х	
SD-38	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-39	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-40	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-41	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-42	Sediment	0.0 - 1.0	Grab sample – Encore sampler		X					Х					Х	
SD-43	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	
SD-44	Sediment	0.0 - 1.0	Grab sample – Encore sampler		Χ					Х					Х	

Abbreviations:

VOCs = volatile organic compounds SVOCs = semivolatile organic compound PCBs = polychlorinated biphenyls PDB = passive diffusion bag TCL = Target Compound List TOC = total organic carbon TAL= Target Analyte List

Sample analyses will be conducted using the following analytical methods:

Target Compound List organics (VOCs, SVOCs, PCBs and pesticides) via SOM01.2, Contract Laboratory Program (CLP Statement of Work for Organic Ar. Target Analyte List metals and cyanide via ISM01.3, CLP Statement of Work for Inorganic Analyses

PCB Congeners via USEPA Method 1668A, Chlorinated Biphenyl Congeners in Water, Soil, Sediment and Tissue by HRGC/HRMS Dioxins and furans via USEPA Method 1613, Dioxins and Furans in Water, Soil, Sediment and Tissue by HRGC/HRMS.

Low-level mercury via USEPA Method 1631, Revision E.

Hardness via SM 2340C.

pH via USEPA Method 9045D.

TOC via the Lloyd Kahn method.

Grain size via ASTM D-422.



Attachment 1 QAPP WORKSHEET # 15a - VOCs **Project Action Limits and Laboratory-Specific Detection/Quantitation Limits** (UFP-QAPP Manual Section 2.6.2.3 and Figure 15)

(EPA 2106-G-05 Section 2.2.6)

				So	il/Sediment				1	Water	
	CAS	Sediment	Soil PAL	DESA MDL (LOW)	DESA RL (LOW)	(MEDIUM)	DESA RL (MEDIUM)	Water PAL	Groundwater	DESA TRACE	DESA TRACE RL
Analyte (method)	Number	PAL (mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(μg/L)	PAL (μg/L)	MDL (μg/L)	(μg/L)
TCL - Volatiles (SOM01.2)											
1,1,1-Trichloroethane	71-55-6	0.213	0.005	0.70	5.00	13.600	250.00	76	30	0.30	0.5
1,1,2,2-Tetrachloroethane	79-34-5	0.850	0.127	0.50	5.00	8.800	250.00	4.7	1.0	0.20	0.5
1,1,2-Trichloro-1,2,2-	76-13-1	NA	NA	0.60	5.00	45.000	250.00	NA	5500	0.30	0.5
1,1,2-Trichloroethane	79-00-5	0.518	2.0	0.50	5.00	14.900	250.00	13	3.0	0.40	0.5
1,1-Dichloroethane	75-34-3	0.027	8.0	0.50	5.00	14.700	250.00	0.29	50	0.60	0.5
1,1-Dichloroethene	75-35-4	0.019	8.3	0.70	5.00	30.700	250.00	4.7	1.0	0.30	0.5
1,2,3-Trichlorobenzene	87-61-6	NA	20	0.80	5.00	24.700	250.00	NA	0.70	0.20	0.5
1,2,4-Trichlorobenzene	120-82-1	5.1	20	1.00	5.00	21.000	250.00	21	9.0	0.30	0.5
1,2-Dibromo-3-	96-12-8	NA	0.080	N/A	N/A	N/A	N/A	NA	0.02	N/A	N/A
1,2-Dibromoethane	106-93-4	NA	0.008	0.40	5.00	14.700	250.00	NA	0.03	0.30	0.5
1,2-Dichlorobenzene	95-50-1	0.294	3.0	0.80	5.00	13.000	250.00	14	600	0.20	0.5
1,2-Dichloroethane	107-06-2	0.260	0.90	0.70	5.00	12.800	250.00	0.29	2.0	0.30	0.5
1,2-Dichloropropane	78-87-5	0.333	2.0	0.40	5.00	6.800	250.00	0.50	1.0	0.20	0.5
1,3-Dichlorobenzene	541-73-1	1.3	38	0.70	5.00	11.000	250.00	38	600	0.20	0.5
1,4-Dichlorobenzene	106-46-7	0.318	20	0.60	5.00	12.500	250.00	9.4	75	0.20	0.5
1,4-Dioxane	123-91-1	NA	7.0	N/A	N/A	N/A	N/A	NA	0.78	N/A	N/A
2-Butanone	78-93-3	0.270	3100	1.10	10.00	28.500	500.00	14000	300	1.90	5.0
2-Hexanone	591-78-6	0.022	20	0.60	10.00	18.200	500.00	99	3.8	2.20	5.0
4-Methyl-2-pentanone	108-10-1	0.033	630	0.80	10.00	19.300	500.00	170	120	2.30	5.0
Acetone	67-64-1	0.009	70000	1.70	10.00	34.000	500.00	1500	6000	1.80	5.0
Benzene	71-43-2	0.142	0.26	0.60	5.00	8.700	250.00	0.15	1.0	0.30	0.5
Bromochloromethane	74-97-5	NA	15	0.80	5.00	23.400	250.00	NA	8.3	0.30	0.5
Bromodichloromethane	75-27-4	NA	0.54	0.50	5.00	16.500	250.00	0.55	1.0	0.20	0.5
Bromoform	75-25-2	0.492	16	0.80	5.00	13.000	250.00	4.3	4.0	0.30	0.5
Bromomethane	74-83-9	0.001	0.24	0.90	5.00	42.400	250.00	16	10	0.90	0.5
Carbon Disulfide	75-15-0	NA	7800	0.50	5.00	21.500	250.00	0.92	700	0.20	0.5
Carbon Tetrachloride	56-23-5	1.5	0.60	0.60	5.00	24.800	250.00	0.33	1.0	0.30	0.5
Chlorobenzene	108-90-7	0.291	13	0.70	5.00	9.600	250.00	47	50	0.30	0.5
Chloroethane	75-00-3	NA	220	0.90	5.00	35.600	250.00	NA	NA NA	0.30	0.5
Chloroform	67-66-3	0.121	0.60	0.60	5.00	19.900	250.00	68	70	0.30	0.5
Chloromethane	74-87-3	NA	4.0	0.80	5.00	13.700	250.00	NA	19	0.40	0.5
cis-1,2-Dichloroethene	156-59-2	NA NA	230	0.70	5.00	24.500	250.00	NA NA	70	0.30	0.5
cis-1,3-Dichloropropene	10061-01-	NA	2.0	0.40	5.00	15.900	250.00	0.34	1.0	0.20	0.5
Cyclohexane	110-82-7	NA	650	0.90	5.00	33.000	250.00	NA	1300	0.30	0.5
Dibromochloromethane	124-48-1	NA	2.1	0.30	5.00	17.800	250.00	0.40	1.0	0.30	0.5
Dichlorodifluoromethane	75-71-8	NA	490	N/A	N/A	N/A	N/A	NA	1000	N/A	N/A
Ethylbenzene	100-41-4	0.175	5.2	0.80	5.00	17.300	250.00	14	700	0.30	0.5
Isopropylbenzene	98-82-8	NA	NA	0.90	5.00	11.300	250.00	NA	NA	0.30	0.5
m,p-Xylene	108-38-3 /	NA NA	NA	N/A	N/A	N/A	N/A	NA NA	NA	N/A	N/A
Methyl Acetate	79-20-9	NA NA	78000	9.90	5.00	26.500	250.00	NA NA	7000	0.40	0.5
Methyl tert-Butyl Ether	7.5 20-3	NA NA	110	9.90 N/A	N/A	N/A	N/A	70	7000	N/A	N/A
Methylcyclohexane	108-87-2	NA NA	NA	0.70	5.00	38.000	250.00	NA	NA	0.30	0.5
Methylene Chloride	75-09-2	0.159	4.1	N/A	N/A	N/A	N/A	2.5	3.0	N/A	N/A
o-Xylene	95-47-6	0.133 NA	65	0.80	5.00	13.500	250.00	NA	19	0.30	0.5
Styrene	100-42-5	0.254	4.7	0.60	5.00	13.800	250.00	32	100	0.30	0.5
Tetrachloroethene	127-18-4	1.0	2.0	0.50	5.00	13.100	250.00	0.34	1.0	0.30	0.5
Toluene	108-88-3	1.0	200	0.50	5.00	7.400	250.00	253	600	0.30	0.5
trans-1,2-Dichloroethene	156-60-5	0.654	0.78	0.80	5.00	13.000	250.00	590	100	0.20	0.5
trans-1,2-Dichloroethene trans-1,3-Dichloropropene	10061-02-	0.654 NA	2.0	0.80	5.00	15.800	250.00	0.34	1.0	0.30	0.5
Trichloroethene		0.112	7.0		N/A						
	79-01-6			N/A		N/A	N/A	1.0	1.0	N/A	N/A
Trichlorofluoromethane	75-69-4	NA 0.202	23000	0.60	5.00	39.500	250.00	NA 0.00	2000	0.30	0.5
Vinyl Chloride	75-01-4	0.202	0.65	N/A	N/A	N/A	N/A	0.08	1.0	N/A	N/A
Xylenes (Total)	1330-20-7	0.433	10	N/A	N/A	N/A	N/A	27	1000	N/A	N/A

Minimum screening levels for the respective medium were derived from the following EBSLs and human health criteria, in the following order

Sediment: NJDEP Ecological Screening Criteria; ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Sediment- Associated Biota (Jones et al. 1997)

Soil: NJDEP Ecological Screening Criteria or NJ Soil Remediation Standards; USEPA (2014) Regional Screening Level (RSL) Residential Soil (Cancer Risk = 1x10-6; NonCancer Hazard = 0.1). Surface Water: NJ GWQC (Freshwater Chronic or Human Health Criteria); ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Aquatic Biota (Suter and Tsao 1996).

Groundwater: NJDEP Groundwater Qualtiy Standards N.J.A.C. 7:9C; USEPA (2014) Regional Screening Level (RSL) Tapwater (Cancer Risk = 1x10-6; NonCancer; Hazard = 0.1).

 $\label{lem:conditional} \mbox{Additional screening levels may be included based on site characterization information.}$

 $\mu g/L$ - microgram per liter EBSL - Ecologically-Based Screening Level MDL - Method Detection Limit mg/kg - milligram per kilogram

NA - Not Available NJ GWQC - New Jersey Groundwater Quality Criteria NJDEP - New Jersey Department of Environmental Protection

ORNL - Oak Ridge National Laboratory RL - Reporting Limit

TCL - Target Compound List

CDM Smith Rolling Knolls Landfill Superfund Site Data Gap Investigation Oversight

QAPP WORKSHEET # 15b - SVOCs Project Action Limits and Laboratory-Specific Detection/Quantitation Limits (UFP-QAPP Manual Section 2.6.2.3 and Figure 15) (EPA 2106-G-05 Section 2.2.6)

TCL - Semivolatiles (SOM01.2) 1,1"-Biphenyl 1,2,4,5-Tetrachlorobenzene 2,2,2"-oxybis(1-Chloropropane) 108-60-1 2,3,4,6-Tetrachlorophenol 2,4,5-Trichlorophenol 2,4,5-Trichlorophenol 2,4,5-Trichlorophenol 2,4,5-Trichlorophenol 2,4,5-Trichlorophenol 2,4,5-Trichlorophenol 2,4,5-Trichlorophenol 2,4-Dimethylphenol 105-67-9 2,4-Dinitrophenol 2,4-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 2,6-Dinitrotoluene 2,6-Dinitrotoluene 2,1-Binitrotoluene 2,6-Dinitrotoluene 2,6-Dinitrotoluene 2,6-Dinitrotoluene 2,6-Dinitrotoluene 2,6-Dinitrotoluene 3,5-7-8 2-Methylnaphthalene 31-58-7 2-Mitroaniline 38-74-4 2-Nitroaniline 38-74-4 2-Nitroaniline 38-74-4 2-Nitroaniline 3-Nitroaniline 3-Nitroaniline 3-Nitroaniline 4-Chloro-3-methylphenol 4-Chloro-3-methylphenol 4-Chloro-3-methylphenol 4-Chlorophenyl-phenylether 4-Chlorophenyl-phenylether 4-Chlorophenyl-phenylether 100-47-8 4-Chlorophenyl-phenylether 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthylene Acetophenone 38-32-9 Acenaphthylene Acetophenone 98-86-2 Anthracene 110-12-7 Atrazine 1912-24-9 Benza(a)pyrene 50-32-8 Benzo(b)fluoranthene 100-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 100-56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 100-60-7 Carbazole Carbazole Chrysene 218-01-9 Dibenzo(a,h)anthracene 56-55-3 Benzo(b)phthalate 117-81-7 Butylbenzylphthalate 218-01-9 Dibenzo(a,h)anthracene 56-65-7 Caprolactam 105-60-2 Carbazole Chrysene 218-01-9 Dibenzo(a,h)anthracene 519-1-4-4 Bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) methane 111-91-	Sediment r PAL (mg/kg) 1.1 1.3 NA NA NA 0.208 0.082 0.304 0.006 0.014 NA 0.417 0.032 NA 0.0127 NA NA NA NA NA 0.127 NA NA NA 0.127 NA	Soil PAL (mg/kg) 3100 2.0 NA 180 4.0 4.0 88 0.010 0.061 0.700 0.012 0.243 230 310 310 NA 0.646 NA 6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	DESA MDL (LOW) (mg/kg) 103.00 108.00 119.00 83.00 226.00 N/A 353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 86.00 N/A 106.00 361.00 76.00 78.00 115.00 115.00 N/A	DESA RL (LOW) (mg/kg) 120 120 120 120 120 120 120 120 120 12	Water PAL (µg/L) 14 0.97 NA NA 1800 0.58 11 100 19 0.11 NA 0.40 24 NA 13 NA	Groundwater PAL (µg/L) 400 0.24 NA 200 700 20 20 100 40 10 10 600 40 3.6 NA 19 NA 30 NA	DESA MDL (LOW) (μg/L) 1.00 0.80 1.23 0.00 0.76 0.55 0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90 0.58	(μg/L) 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
TCL - Semivolatiles (SOM01.2)	1.1 1.3 NA NA NA NA 0.208 0.082 0.304 0.006 0.014 NA 0.417 0.032 NA 0.127 NA NA NA NA 0.127 NA NA NA 0.127 NA NA 0.127 NA NA NA NA 0.127 NA NA NA 0.127 NA NA NA 0.127 NA NA NA NA NA NA NA NA NA 0.013 0.007 0	3100 2.0 NA 180 4.0 4.0 4.0 0.61 0.700 0.700 0.701 0.243 230 310 39 NA 0.646 NA 12.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	103.00 108.00 119.00 83.00 226.00 N/A 353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 86.00 N/A 106.00 361.00 76.00 78.00 115.00 115.00 N/A	120 120 120 120 120 120 120 120 N/A 120 120 120 120 120 120 120 120 120 120	14 0.97 NA NA 1800 0.58 11 100 19 0.11 NA 0.40 24 NA	400 0.24 NA	1.00 0.80 1.23 0.00 0.76 0.55 0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
1,2,4,5-Tetrachlorobenzene 95-94-3 2,2'-oxybis(1-Chloropropane) 108-60-1 2,3,4,6-Tetrachlorophenol 58-90-2 2,4,5-Trichlorophenol 95-95-4 2,4,6-Trichlorophenol 38-06-2 2,4-Dichlorophenol 105-67-9 2,4-Dimethylphenol 105-67-9 2,4-Dinitrophenol 120-83-2 2,4-Dinitrophenol 121-14-2 2,6-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2,C-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitrophenol 88-75-5 3,3-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 106-47-8 4-Chlorophenyl-phenylether 100-01-6 4-Nitrophenol 100-02-7 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzo(a)pyrene 89-80-2 Benzo(a)pyrene 98-86-2 Benzo(b)fluoranthene 205-99-2 Benzo(a)pyrene 191-24-2 Benzo(a)phthalate 107-81-81 Benzo(a)phthalate 107-81-81 Butylbenzylphthalate 117-81-7 Benzo(a),h)perylene 191-24-2 Benzo(b)fluoranthene 207-08-9 bis(2-Chloroethyr) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Benzo(a,h)anthracene 33-70-3 Dibenzofuran 132-64-9 Dienzo(a,h)anthracene 131-11-3 Di-n-butylphthalate 34-76-2 Dimethylphthalate 34-76-2 Dimethylphthalate 34-76-8 Hexachlorobutadiene 37-68-3 Hexachlorobutadiene 37-69-3 Sophothalene 39-95-3 Sophothalene 39-95-3	1.3 NA NA NA NA NA 0.208 0.082 0.304 0.006 0.014 NA 0.417 0.032 NA 0.012 NA NA 1.2 NA	2.0 NA 180 4.0 4.0 88 0.010 0.061 0.700 0.700 0.012 0.243 230 310 39 NA 0.646 NA 6.0 NA 180 180 180 180 180 180 180 18	108.00 119.00 83.00 226.00 N/A 353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 86.00 229.00 86.00 108.00 76.00 76.00 78.00 115.00 115.00 N/A	120 120 120 120 120 120 N/A 120 120 120 120 120 120 120 120 120 120	0.97 NA NA 1800 0.58 11 100 19 0.11 NA 0.40 24 NA 13 NA	0.24 NA 200 700 20 20 100 40 10 10 600 40 3.6 NA 19 NA 30 NA	0.80 1.23 0.00 0.76 0.55 0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,2'-oxybis(1-Chloropropane) 108-60-1 2,3,4,6-Tetrachlorophenol 58-90-2 2,4,5-Trichlorophenol 95-95-4 2,4,6-Trichlorophenol 120-83-2 2,4-Dirichlorophenol 120-83-2 2,4-Dimethylphenol 105-67-9 2,4-Dinitrophenol 121-14-2 2,4-Dinitrophenol 121-14-2 2,6-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Acetophenone 98-86-2 Acetophenone 98-86-2 Benzo(a)anthracene 120-12-7 Benzo(a)anthracene 100-52-7 Benzo(a)pyrene 191-24-2 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyy) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 34-66-2 Dimethylphthalate 34-66-2 Dimethylphthalate 34-74-2 Dien-octylphthalate 34-74-2 Dien-octylphthalate 34-74-2 Dien-octylphthalate 34-74-2 Dien-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 206-44-0 Fluoranthene 37-68-3 Hexachlorobutadiene 37-69-3 Sophothalene 39-95-3 Sophothalene 39-95-3 Sophothalene 39-95-3	NA NA NA NA 0.208 0.082 0.304 0.006 0.014 NA 0.417 0.032 NA 0.012 NA NA 1.2 NA	NA 180 4.0 4.0 4.0 0.061 0.700 0.700 0.700 0.702 0.243 230 310 39 NA 0.646 NA 6.0 NA SA	119.00 83.00 226.00 N/A 353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 111.00 N/A 235.00 68.00 229.00 86.00 108.00 378.00 110.00 108.00 110.00 108.00 110.00 108.00 110.00 108.00	120 120 120 120 N/A 120 120 N/A 120 120 120 120 120 120 120 120 120 120	NA NA 1800 0.58 11 100 19 0.11 NA 0.40 24 NA 13 NA	NA 200 700 20 20 100 40 10 10 10 600 3.6 NA 19 NA	1.23 0.00 0.76 0.55 0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,3,4,6-Tetrachlorophenol 58-90-2 2,4,5-Trichlorophenol 95-95-4 2,4,6-Trichlorophenol 120-83-2 2,4-Dinitrophenol 105-67-9 2,4-Dinitrophenol 51-28-5 2,4-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2-Chloroaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-75-5 2-Nitrophenol 38-74-4 2-Nitrophenol 38-75-5 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chloroaniline 106-47-8 4-Chloroaniline 106-47-8 4-Nitroaniline 100-02-7 4-Chloroaniline 100-02-7 4-Chloroaniline 100-02-7 4-Chloroaniline 100-02-7 4-Chloroaniline	NA NA O.208 O.304 O.006 O.014 NA O.417 O.032 NA O.012 NA NA O.127 NA NA NA O.127 NA NA O.137 O.006 NA O.013 O.007	180 4.0 4.0 88 0.010 0.061 0.700 0.700 0.012 230 310 39 NA 0.646 NA SA	83.00 226.00 N/A 353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 115.00 115.00 N/A	120 120 N/A 120 N/A 120 N/A 120 120 120 120 120 120 120 120 120 120	NA 1800 0.58 11 100 19 0.11 NA 0.40 24 NA 13 NA	200 700 20 20 100 40 10 10 10 600 3.6 NA 19 NA 30 NA	0.00 0.76 0.55 0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,4,5-Trichlorophenol 95-95-4 2,4,6-Trichlorophenol 120-83-2 2,4-Dinitrophenol 105-67-9 2,4-Dinitrophenol 51-28-5 2,4-Dinitrophenol 51-28-5 2,4-Dinitrotoluene 606-20-2 2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-78-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Mitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-44-5 4-Nitrophenol 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzo(a)pyrene 50	NA	4.0 4.0 4.0 88 0.010 0.061 0.700 0.700 0.012 0.243 230 310 39 NA 0.646 NA 6.0 NA SA	226.00 N/A 353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 115.00 115.00 N/A	120 N/A 120 120 120 120 120 120 120 120 120 120	1800 0.58 11 100 19 0.11 NA 0.40 24 NA 13 NA	700 20 20 100 40 10 10 10 600 40 3.6 NA 19 NA 30 NA	0.76 0.55 0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,4,6-Trichlorophenol 88-06-2 2,4-Dichlorophenol 120-83-2 2,4-Dinitrophenol 105-67-9 2,4-Dinitrophenol 51-28-5 2,4-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Mitroaniline 88-74-4 2-Nitroaniline 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4/ 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthylene 83-32-9 Acetophenone 98-86-2 Arthracene 120-12-7 Atrazine	0.208 0.082 0.304 0.006 0.014 NA 0.417 0.032 NA 0.012 NA NA 0.127 NA NA 1.2 NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	4.0 88 0.010 0.061 0.700 0.700 0.012 0.243 230 310 39 NA 0.646 NA 6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	N/A 353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 115.00 115.00 N/A	N/A 120 120 N/A 120 120 N/A 120 120 120 120 120 120 120 120 120 120	0.58 11 100 19 0.11 NA 0.40 24 NA 13 NA	20 20 100 40 10 10 600 40 3.6 NA 19 NA NA NA NA NA NA NA NA NA NA NA NA NA	0.55 0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,4-Dichlorophenol 120-83-2 2,4-Dimethylphenol 105-67-9 2,4-Dinitrophenol 51-28-5 2,4-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2-Chlorophenol 91-58-7 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-74-4 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloro-amethylphenol 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-47-8 4-Nitrophenol 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzo(a)anthracene 50	0.082 0.304 0.006 0.014 NA 0.417 0.032 NA 0.012 NA NA 0.127 NA NA 1.2 NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	88 0.010 0.061 0.700 0.700 0.012 0.243 230 310 39 NA 0.646 NA 5.0 NA 12.7 NA 21 25 NA 20 682 780 1480 210 6100 0.60	353.00 325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 N/A	120 120 N/A 120 120 120 120 120 120 120 120 120 120	11 100 19 0.11 NA 0.40 24 NA 13 NA NA NA NA NA NA 1.5 NA	20 100 40 10 600 40 3.6 NA 19 NA NA NA NA NA NA NA NA NA NA	0.94 1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 20 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,4-Dimethylphenol 105-67-9 2,4-Dinitrophenol 51-28-5 2,4-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 38-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-47-8 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthylene 83-32-9 Acenaphthylene 33-32-9 Acenaphthylene 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzol(a)anthracene 56-5	0.304 0.006 0.014 NA 0.417 0.032 NA 0.012 NA NA 0.127 NA NA 1.2 NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	0.010 0.061 0.700 0.700 0.700 0.700 0.243 230 310 39 NA 0.646 NA 6.0 NA 12.7 NA 2.7 NA 21 25 NA 21 20 682 780 1480 210 6100 0.60	325.00 N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 115.00 115.00 N/A	120 N/A 120 120 120 120 120 120 120 120 120 120	100 19 0.11 NA 0.40 24 NA 13 NA NA 0.02 NA NA NA NA 1.5 NA	100 40 10 10 600 40 3.6 NA 19 NA 30 NA	1.81 0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.34 0.35 0.72 0.77 0.90	5 20 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,4-Dinitrophenol 51-28-5 2,4-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitrophenol 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 / 4-Nitroaniline 100-44-5 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzo(a)aphthracene 56-55-3 Benzo(b)fluoranthene 205-99	0.006 0.014 NA 0.417 0.032 NA 0.012 NA NA 0.127 NA NA 1.2 NA 0.013 0.007 0.006 NA 0.220 NA NA NA NA NA 0.320 0.370 10 0.170	0.061 0.700 0.700 0.700 0.012 0.243 230 310 39 NA 0.646 NA 6.0 NA SA	N/A 92.00 68.00 95.00 401.00 108.00 377.00 93.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 115.00 N/A	N/A 120 120 120 120 120 120 120 120 120 120	19 0.11 NA 0.40 24 NA 13 NA NA 0.02 NA 15 NA NA 1.5 NA	40 10 10 600 40 3.6 NA 19 NA 30 NA NA NA NA NA NA NA NA NA NA NA NA NA	0.33 0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	20 5 5 5 5 5 5 5 5 5 5 5 5 5
2,4-Dinitrotoluene 121-14-2 2,6-Dinitrotoluene 606-20-2 2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 100-47-8 4-Nitroaniline 100-01-6 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzolaphtylene 100-52-7 Benzolaphtylene 50-32-8	0.014 NA 0.417 0.032 NA 0.012 NA NA 0.127 NA NA NA 1.2 NA NA NA NA NA NA NA NA 0.0320 NA NA 0.320 0.370 10 0.170	0.700 0.700 0.700 0.700 0.012 0.243 230 310 39 NA 0.646 NA 6.0 NA 2.7 NA 2.7 NA 21 25 NA 20 682 780 1480 210 6100 0.60	92.00 68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 120 120 120 120 120 120 120	0.11 NA 0.40 24 NA 13 NA NA 13 NA NA NA NA NA 0.02 NA	10 10 600 40 3.6 NA 19 NA 30 NA	0.48 0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2,6-Dinitrotoluene 606-20-2 2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 53-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloro-3-methylphenol 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-47-8 4-Nitroaniline 100-01-6 4-Nitroaniline 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benza(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(k)fluoranthene	NA 0.417 0.032 NA 0.012 NA NA 0.127 NA NA 1.2 NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	0.700 0.012 0.243 230 310 39 NA 0.646 NA 6.0 NA SA 2.7 NA 225 NA 20 682 780 1480 210 6100 0.60	68.00 95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 120 120 120 120 120 120 120	NA 0.40 24 NA 13 NA NA 0.02 NA	10 600 40 3.6 NA 19 NA 30 NA NA NA NA NA NA NA NA NA NA NA NA NA	0.79 0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2-Chloronaphthalene 91-58-7 2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 53-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloro-3-methylphenol 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-47-8 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 56-55-3 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(g,h)iperylene	0.417 0.032 NA 0.012 NA NA 0.127 NA NA 1.2 NA NA NA NA 0.013 0.007 0.006 NA NA 0.220 NA NA 0.320 0.370 10 0.170	0.012 0.243 230 310 319 NA 0.646 NA 6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	95.00 401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 120 120 120 120 120 120 120	0.40 24 NA 13 NA 0.02 NA NA 1.5 NA	600 40 3.6 NA 19 NA 30 NA NA NA NA NA NA NA NA NA NA NA	0.80 1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2-Chlorophenol 95-57-8 2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-47-8 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzaldehyde 100-52-7 Benzaldehyde 100-52-7 Benzolajpyrene 50-32-8 Benzolojhfluoranthene 205-99-2 Benzolojhfluoranthene 205-99-2	0.032 NA 0.012 NA NA NA 0.127 NA NA 1.2 NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA NA 0.320 0.370 10 0.170	0.243 230 310 39 NA 0.646 NA 6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	401.00 108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 N/A	120 120 120 120 120 120 120 120 N/A 400 120 120 120 120 120 120 120 120 120 1	24 NA 13 NA NA NA NA NA 1.5 NA	40 3.6 NA 19 NA 30 NA NA NA NA NA 30 NA NA NA NA NA NA NA	1.43 0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2-Methylnaphthalene 91-57-6 2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-47-8 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylp	NA 0.012 NA NA 0.127 NA NA NA 1.2 NA NA NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA NA 0.320 0.370 10 0.170	230 310 39 NA 0.646 NA 6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	108.00 377.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 120 120 120 N/A 400 120 120 120 120 120 120 120 120 120 1	NA 13 NA NA 0.02 NA NA 1.5 NA	3.6 NA 19 NA 30 NA NA NA NA NA NA NA NA NA N	0.88 0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2-Methylphenol 95-48-7 2-Nitroaniline 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 / 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzolalehyde 100-52-7 Benzolalehyde 56-55-3 Benzolalpyrene 50-32-8 Benzolalpyrene 50-32-8 Benzolalpyrene 50-32-8 Benzolalpyrene 191-24-2 Benzolalpyrene 191-24-2 Benzolalpyrene 191-24-2 Benzolalpyrene 191-24-2	0.012 NA NA 0.127 NA NA 1.2 NA NA NA NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	310 39 NA 0.646 NA 6.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 NA 10.0 10.	377.00 93.00 93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 120 120 N/A 400 120 120 120 120 120 120 120 120 120 1	13 NA NA 0.02 NA NA 1.5 NA NA NA NA NA NA	NA 19 NA 30 NA	0.99 0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 10 5 5 5 5 5 5 4 7 4 7 7 8
2-Nitrophenol 88-74-4 2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 / 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)apyrene 56-55-3 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis(2-Chloroethyl) ether 111-44-4 bis(2-Chloroethyl) ether 111-91-1 bis(2-Chloroethyl) ether 111-41-3	NA NA O.127 NA NA 1.2 NA NA NA NA NA NA NA NA O.013 O.007 O.006 NA O.220 NA NA O.320 O.370 10 O.170	39 NA 0.646 NA 6.0 NA NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	93.00 378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 N/A 400 120 120 120 120 120 120 120 120 120 1	NA NA 0.02 NA NA 1.5 NA	19 NA 30 NA NA NA NA NA NA NA A NA NA NA NA NA N	0.70 1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5
2-Nitrophenol 88-75-5 3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-44-5 4-Nitroaniline 100-01-6 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Actoraphthylene 208-96-8 Actophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzo(a)anthracene 56-53-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 15-60-2 Carbazole	NA 0.127 NA NA NA NA NA NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA NA 0.320 0.370 10 0.170	NA 0.646 NA 6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	378.00 111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 N/A 400 120 120 120 120 120 120 120 120 120 1	NA 0.02 NA NA 1.5 NA NA NA NA NA NA 4440 NA	NA 30 NA NA NA NA NA NA NA 30 NA	1.08 0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 5 5 5 5 5 5 4 7 4 7 4 7 5 5 5 5
3,3'-Dichlorobenzidine 91-94-1 3-Nitroaniline 99-09-2 4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 106-44-5 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 17-81-7 Butylbenzylphthalate 85-68-7 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzofuran	0.127 NA NA NA 1.2 NA NA NA NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	0.646 NA 6.0 NA NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	111.00 N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 N/A 400 120 120 120 120 120 120 120 120 120 1	0.02 NA NA 1.5 NA NA NA NA NA NA NA	30 NA NA NA NA 30 NA NA 3.8 NA 400 NA	0.40 0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 10 5 5 5 5 5 5 4n/A 5 10 5 5
3-Nitroaniline	NA NA 1.2 NA NA NA NA NA NA NA NA O.013 O.007 O.006 NA O.220 NA NA O.320 O.370 10 O.170	NA 6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	N/A 235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 N/A	N/A 400 120 120 120 120 120 120 120 120 120 1	NA N	NA N	0.76 0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 10 5 5 5 5 5 4N/A 5 10 5 5
4,6-Dinitro-2-methylphenol 534-52-1 4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chlorophenyl-phenylether 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzolalphyde 100-52-7 Benzolalpyrene 56-55-3 Benzo(a)anthracene 56-55-3 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 117-81-7 Carbazole 86-74-8	NA 1.2 NA NA NA NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	6.0 NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	235.00 68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	400 120 120 120 120 120 N/A 120 400 120 120 120	NA 1.5 NA	NA NA NA 30 NA NA 34 NA NA 3.8 NA 400 NA 700	0.85 0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	10 5 5 5 5 #N/A 5 10 5 5
4-Bromophenyl-phenylether 101-55-3 4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 / 106-44-5 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzoldehyde 100-52-7 Benzolaphtracene 56-55-3 Benzo(a)ptrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethy) ether 111-91-1 bis(2-Chloroethy) ether 111-44-4 bis(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 117-81-7 Butylbenzylphthalate 117-81-7 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzofuran <td>1.2 NA NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10</td> <td>NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60</td> <td>68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 N/A</td> <td>120 120 120 120 120 N/A 120 400 120 120 120</td> <td>1.5 NA NA NA NA NA NA AA AA AA AA AA AA AA</td> <td>NA NA 30 NA NA 3.8 NA 400 NA 700</td> <td>0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90</td> <td>5 5 5 5 #N/A 5 10 5 5</td>	1.2 NA NA NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10	NA NA 2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	68.00 229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 N/A	120 120 120 120 120 N/A 120 400 120 120 120	1.5 NA NA NA NA NA NA AA AA AA AA AA AA AA	NA NA 30 NA NA 3.8 NA 400 NA 700	0.58 0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 5 #N/A 5 10 5 5
4-Chloro-3-methylphenol 59-50-7 4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 / 106-44-5 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 205-99-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis(2-Chloroethyl) ether 111-44-4 bis(2-Chloroethyl) ether 111-44-4 bis(2-Chloroethyl) ether 111-44-4 bis(2-Chloroethyl) ether 111-91-1 bis(2-Chloroethyl) ether 111-49-1 bis(2-Chloroethyl) ether 111-91-1 bis(2-Chloroethyl) ether 111-41-4 <td>NA NA NA NA NA NA NA O.013 O.007 O.006 NA O.220 NA NA O.320 O.370 10 O.170</td> <td>NA 2.7 NA 31 25 NA 20 682 1480 210 6100 0.60</td> <td>229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A</td> <td>120 120 120 N/A 120 400 120 120 120 120</td> <td>NA NA NA NA NA NA 60 38 4840 NA</td> <td>NA 30 NA NA 3.8 NA 400 NA 700</td> <td>0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90</td> <td>5 5 5 #N/A 5 10 5 5</td>	NA NA NA NA NA NA NA O.013 O.007 O.006 NA O.220 NA NA O.320 O.370 10 O.170	NA 2.7 NA 31 25 NA 20 682 1480 210 6100 0.60	229.00 82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 N/A 120 400 120 120 120 120	NA NA NA NA NA NA 60 38 4840 NA	NA 30 NA NA 3.8 NA 400 NA 700	0.62 0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 5 #N/A 5 10 5 5
4-Chloroaniline 106-47-8 4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 / 106-44-5 4-Nitroaniline 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) methane 111-91-1 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 118-74-1 Hexachlorobenzene 193-95-3 Nitrobenzene 98-95-3 Nitrobenzene 98-95-3	NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	2.7 NA 31 25 NA 20 682 780 1480 210 6100 0.60	82.00 86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 120 N/A 120 400 120 120 120 120	NA NA NA NA 60 38 4840 NA	30 NA NA 3.8 NA 400 NA 700	0.42 0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 5 #N/A 5 10 5 5 5
4-Chlorophenyl-phenylether 7005-72-3 3&4-Methylphenol 108-39-4 / 106-44-5 4-Nitrophenol 100-01-6 4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 15-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Dibenzofuran 132-64-9 Diethylphthalate 117-84-0 Piuoranthene 206-44-0 Fluoranthene 206-44-0	NA NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	NA 31 25 NA 20 682 780 1480 210 6100 0.60	86.00 N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 N/A 120 400 120 120 120 120	NA NA NA 60 38 4840 NA	NA NA 3.8 NA 400 NA 700	0.57 #N/A 0.34 0.35 0.72 0.77 0.90	5 #N/A 5 10 5 5 5
3&4-Methylphenol 108-39-4 / 106-44-5	NA NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	31 25 NA 20 682 780 1480 210 6100 0.60	N/A 106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	N/A 120 400 120 120 120 120	NA NA 60 38 4840 NA	NA 3.8 NA 400 NA 700	#N/A 0.34 0.35 0.72 0.77 0.90	#N/A 5 10 5 5 5
106-44-5 106-04-5 106-04-5 106-04-5 106-04-5 100-01-6 100-02-7	NA 0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	25 NA 20 682 780 1480 210 6100 0.60	106.00 361.00 76.00 78.00 116.00 51.00 115.00 N/A	120 400 120 120 120 120	NA 60 38 4840 NA	3.8 NA 400 NA 700	0.34 0.35 0.72 0.77 0.90	5 10 5 5 5
4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(b,fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 15-68-7 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzola,)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 34-66-2 Dimethylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-0 Hexachlorobutadiene 47-68-3 Hexachlorobutadiene 77-47-4 <td>0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170</td> <td>NA 20 682 780 1480 210 6100 0.60</td> <td>361.00 76.00 78.00 116.00 51.00 115.00 N/A</td> <td>400 120 120 120 120</td> <td>60 38 4840 NA</td> <td>NA 400 NA 700</td> <td>0.35 0.72 0.77 0.90</td> <td>10 5 5 5</td>	0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	NA 20 682 780 1480 210 6100 0.60	361.00 76.00 78.00 116.00 51.00 115.00 N/A	400 120 120 120 120	60 38 4840 NA	NA 400 NA 700	0.35 0.72 0.77 0.90	10 5 5 5
4-Nitrophenol 100-02-7 Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 205-99-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 15-68-7 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzofuran 132-64-9 Diethylphthalate 53-70-3 Diethylphthalate 131-11-3 Di-n-octylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 206-44-0 Fluoranthene 46-63-3 Hexachlorobutadiene 47-47-4 <	0.013 0.007 0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	NA 20 682 780 1480 210 6100 0.60	361.00 76.00 78.00 116.00 51.00 115.00 N/A	400 120 120 120 120	60 38 4840 NA	NA 400 NA 700	0.35 0.72 0.77 0.90	10 5 5 5
Acenaphthene 83-32-9 Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(b)fluoranthene 205-99-2 Benzo(b)fluoranthene 205-99-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 131-11-3 Di-n-butylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 86-73-7 Hexachlorobutadiene 87-68-3 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroothane <td< td=""><td>0.007 0.006 NA 0.220 NA NA 0.320 0.370 10</td><td>20 682 780 1480 210 6100 0.60</td><td>76.00 78.00 116.00 51.00 115.00 N/A</td><td>120 120 120 120</td><td>38 4840 NA</td><td>400 NA 700</td><td>0.72 0.77 0.90</td><td>5 5 5</td></td<>	0.007 0.006 NA 0.220 NA NA 0.320 0.370 10	20 682 780 1480 210 6100 0.60	76.00 78.00 116.00 51.00 115.00 N/A	120 120 120 120	38 4840 NA	400 NA 700	0.72 0.77 0.90	5 5 5
Acenaphthylene 208-96-8 Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzolalphyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-74-2 Di-n-octylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluoranthene 117-84-0 Fluoranthene 86-73-7 Hexachlorobutadiene 87-68-3 Hexachlorobutadiene 87-68-3	0.006 NA 0.220 NA NA 0.320 0.370 10 0.170	682 780 1480 210 6100 0.60	78.00 116.00 51.00 115.00 N/A	120 120 120	4840 NA	NA 700	0.77 0.90	5 5
Acetophenone 98-86-2 Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 17-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 48-76-2 Dimethylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobutadiene 118-74-1 Hexachlorocyclopentadiene 77-47-4 Hexachlorochane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5	NA 0.220 NA NA 0.320 0.370 10 0.170	780 1480 210 6100 0.60	116.00 51.00 115.00 N/A	120 120	NA	700	0.90	5
Anthracene 120-12-7 Atrazine 1912-24-9 Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 167-37 Hexachlorobenzene 118-74-1 Hexachlorobenzene 193-95-3 Nitrobenzene 98-95-3 Nitrobenzene 98-95-3	0.220 NA NA 0.320 0.370 10 0.170	1480 210 6100 0.60	51.00 115.00 N/A	120				
Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chlorethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 17-81-7 Butylbenzylphthalate 85-68-7 Carpolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluorene 86-73-7 Hexachlorobutadiene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorochane 67-72-1 Insophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3 </td <td>NA 0.320 0.370 10 0.170</td> <td>6100 0.60</td> <td>N/A</td> <td>120</td> <td>0.04</td> <td></td> <td>. 0.30</td> <td>5</td>	NA 0.320 0.370 10 0.170	6100 0.60	N/A	120	0.04		. 0.30	5
Benzaldehyde 100-52-7 Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chlorethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 17-81-7 Butylbenzylphthalate 85-68-7 Carpolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluorene 86-73-7 Hexachlorobutadiene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorochane 67-72-1 Insophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3 </td <td>NA 0.320 0.370 10 0.170</td> <td>6100 0.60</td> <td>N/A</td> <td></td> <td>NA</td> <td>3.0</td> <td>1.50</td> <td>5</td>	NA 0.320 0.370 10 0.170	6100 0.60	N/A		NA	3.0	1.50	5
Benzo(a)anthracene 56-55-3 Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bisi(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 17-81-7 Butylbenzylphthalate 85-68-7 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorochane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 99-20-3 Nitrobenzene	0.370 10 0.170	_		N/A	NA	200	0.10	5
Benzo(a)pyrene 50-32-8 Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Carpolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocthane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.370 10 0.170	_	49.00	120	0.03	0.10	0.58	5
Benzo(b)fluoranthene 205-99-2 Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 131-11-3 Di-n-butylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocthane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.170	0.20	39.00	120	0.00	0.10	0.55	5
Benzo(g,h,i)perylene 191-24-2 Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3		0.60	30.00	120	0.00	0.20	0.41	5
Benzo(k)fluoranthene 207-08-9 bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethoxy) ther 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorotenane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Nitrobenzene 98-95-3		119	35.00	120	7.64	NA	0.35	5
bis(2-Chloroethoxy) methane 111-91-1 bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 131-11-3 Di-n-butylphthalate 131-11-3 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorochane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.240	6	31.00	120	0.38	0.50	0.60	5
bis(2-Chloroethyl) ether 111-44-4 bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorochane 67-72-1 Insophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	NA	23	124.00	120	NA	5.9	0.97	5
bis-(2-Ethylhexyl)phthalate 117-81-7 Butylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluoranthene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroothane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	3.5	0.40	130.00	120	0.03	7.0	1.38	5
Burylbenzylphthalate 85-68-7 Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Pluoranthene 206-44-0 Fluoranthene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.182	0.93	124.00	120	0.30	3.0	0.68	5
Caprolactam 105-60-2 Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-65-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 44-74-2 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	2.0	0.24	38.00	120	23	100	0.49	5
Carbazole 86-74-8 Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	NA	31000	231.00	120	NA	990	1.00	5
Chrysene 218-01-9 Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	NA	24	72.00	120	NA	NA	1.20	5
Dibenzo(a,h)anthracene 53-70-3 Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 117-84-0 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.340	4.7	52.00	120	3.8	5.0	0.53	5
Dibenzofuran 132-64-9 Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 84-74-2 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorochane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.060	0.20	N/A	N/A	0.004	0.30	0.42	5
Diethylphthalate 84-66-2 Dimethylphthalate 131-11-3 Di-n-butylphthalate 84-74-2 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.420	7.2	64.00	120	3.7	0.79	0.72	5
Dimethylphthalate 131-11-3 Di-n-butylphthalate 84-74-2 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.295	25	87.00	120	110	6000	0.39	5
Di-n-butylphthalate 84-74-2 Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	NA	NA NA	60.00	120	NA NA	NA	0.47	5
Di-n-octylphthalate 117-84-0 Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachlorocethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	1.1	0.150	510.00	120	9.7	700	0.48	5
Fluoranthene 206-44-0 Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	NA	2400	42.00	120	708	100	0.57	5
Fluorene 86-73-7 Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.750	122	30.00	120	1.9	300	0.51	5
Hexachlorobenzene 118-74-1 Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.190	122	86.00	120	19	300	0.61	5
Hexachlorobutadiene 87-68-3 Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.020	0.199	63.00	120	0.00028	0.02	0.49	5
Hexachlorocyclopentadiene 77-47-4 Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.020	0.199	126.00	120	0.00028	1.0	1.02	5
Hexachloroethane 67-72-1 Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.027	0.755	25.00	120	40	40	0.92	5
Indeno(1,2,3-cd)pyrene 193-39-5 Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.584	0.733	124.00	120	1.4	7.0	1.35	5
Isophorone 78-59-1 Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.200	0.600	29.00	120	0.04	0.20	0.50	5
Naphthalene 91-20-3 Nitrobenzene 98-95-3	0.432	139	83.00	120	35	40	0.76	5
Nitrobenzene 98-95-3	0.176	0.099	125.00	120	13	300	1.05	5
	0.145	1.3	115.00	120	17	6	1.13	5
N-Nitroso-di-n-propylamine 86-30-6	NA	0.200	99.00	120	0.01	10	0.61	5
N-Nitrosodiphenylamine 621-64-7	NA	0.545	104.00	120	3.30	10	0.99	5
Pentachlorophenol 87-86-5	23	0.119	324.00	400	0.27	0.30	0.91	10
Phenanthrene 85-01-8	0.560	45.7	N/A	N/A	3.6	NA	0.47	5
Phenol 108-95-2	0.049	30.0	374.00	120	180	2000	1.36	5
Pyrene 129-00-0	0.490	78.5	37.00	120	0.30	200	0.53	5
Semivolatiles-SIM (SOM01.2)			<u> </u>					
2-Methylnaphthalene 91-57-6	NA	230	0.0002	0.0033	NA	3.6	0.88	5
Acenaphthene 83-32-9	0.007	20	0.0002	0.0033	38	400	0.72	5
Acenaphthylene 208-96-8	0.006	682	0.0006	0.0033	4840	NA	0.77	5
Anthracene 120-12-7	0.220	1480	0.0001	0.0033	0.035	2000	0.58	5
Benzo(a)anthracene 56-55-3	0.320	0.600	0.0003	0.0033	0.025	0.10	0.58	5
Benzo(a)pyrene 50-32-8	0.370	0.200	0.0003	0.0033	0.0038	0.10	0.55	5
Benzo(b)fluoranthene 205-99-2	10	0.600	0.0002	0.0033	0.0038	0.20	0.41	5
Benzo(g,h,i)perylene 191-24-2	0.170	119.0	0.0002	0.0033	7.64	NA	0.35	5
Benzo(k)fluoranthene 207-08-9		6.0	0.0002	0.0033	0.38	0.50	0.60	5
Chrysene 218-01-9	0.240	4.7	0.0002	0.0033	3.8	5.0	0.53	5
Dibenzo(a,h)anthracene 53-70-3		0.200	0.0002	0.0033	0.0038	0.30	0.42	5
Fluoranthene 206-44-0	0.240	122	0.0003	0.0033	1.9	300	0.51	5
Fluorene 86-73-7	0.240 0.340	122	0.0002	0.0033	19	300	0.61	5
Indeno(1,2,3-cd)pyrene 193-39-5	0.240 0.340 0.060 0.750	0.600	0.0002	0.0033	0.038	0.20	0.50	5
Naphthalene 91-20-3	0.240 0.340 0.060 0.750 0.190		0.0002	3.0000	13	300	1.05	5
Pentachlorophenol 87-86-5	0.240 0.340 0.060 0.750 0.190 0.200	0 099	0.0019	0.0067	0.27	0.30	0.91	10
Phenanthrene 85-01-8	0.240 0.340 0.060 0.750 0.190 0.200 0.176	0.099	0.0019	0.0033	3.6	NA	0.47	5
Pyrene 129-00-0	0.240 0.340 0.060 0.750 0.190 0.200 0.176 23	0.119		0.0033	0.30	200	0.53	5
Notes:	0.240 0.340 0.060 0.750 0.190 0.200 0.176		0.0002		0.55	200	0.55	

Minimum screening levels for the respective medium were derived from the following EBSLs and human health criteria, in the following order:

Sediment: NJDEP Ecological Screening Criteria; ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Sediment- Associated Biota (Jones et al. 1997).

Soil: NJDEP Ecological Screening Criteria or NJ Soil Remediation Standards; USEPA (2014) Regional Screening Level (RSL) Residential Soil (Cancer Risk = 1x10-6; NonCancer Hazard = 0.1).

Surface Water: NJ GWQC (Freshwater Chronic or Human Health Criteria); ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Aquatic Biota (Suter and Tsao 1996).

Groundwater: NJDEP Groundwater Quality Standards N.J.A.C. 7:9C; USEPA (2014) Regional Screening Level (RSL) Tapwater (Cancer Risk = 1x10-6; NonCancer; Hazard = 0.1). Additional screening levels may be included based on site characterization information.

 $\mu g/L$ - microgram per liter EBSL - Ecologically-Based Screening Level MDL - Method Detection Limit

mg/kg - milligram per kilogram NA - Not Available

NJ GWQC - New Jersey Groundwater Quality Criteria

NJDEP - New Jersey Department of Environmental Protection ORNL - Oak Ridge National Laboratory

RL - Reporting Limit
TCL - Target Compound List



QAPP WORKSHEET # 15c - Metals Project Action Limits and Laboratory-Specific Detection/Quantitation Limits (UFP-QAPP Manual Section 2.6.2.3 and Figure 15) (EPA 2106-G-05 Section 2.2.6)

			S	oil/Sediment			Water					
Analyte (method)	CAS Number	Sediment PAL (mg/kg)	Soil PAL (mg/kg)	DESA MDL (LOW) (mg/kg)	DESA RL (LOW) (mg/kg)	Surface Water PAL (μg/L)	Groundwater PAL (μg/L)	DESA TRACE MDL	DESA TRACE RL (μg/L)			
TAL - Metals (ISM01.3)												
Aluminum	7429-90-5	58030	50	0.0	10	87	200	0.0	10			
Antimony	7440-36-0	12	5.0	0.2	2	5.6	6.0	0.0	2			
Arsenic	7440-38-2	6	9.9	0.4	1	0.017	3.0	0.1	1			
Barium	7440-39-3	NA	283	0.2	10	220	6000	0.0	1			
Beryllium	7440-41-7	NA	10	0.0	0	3.6	1.0	0.0	1			
Cadmium	7440-43-9	0.60	0.360	0.0	0	3.4	4.0	0.0	1			
Calcium	7440-70-2	NA	NA	12.6	50	NA	NA	0.0	-			
Chromium	7440-47-3	26	0.40	0.3	1	42	70	0.3	1			
Cobalt	7440-48-4	50	0.14	0.0	2	24	100	0.0	1			
Copper	7440-50-8	16	5.4	0.3	1	1300	1300	0.3	1			
Iron	7439-89-6	NA	5500	*	5	158	300	0.0	-			
Lead	7439-92-1	31	11	0.2	1	5.0	5.0	0.0	1			
Magnesium	7439-95-4	NA	NA	5.1	50	NA	NA	0.0	-			
Manganese	7439-96-5	630	220	0.3	1	120	50	0.1	1			
Mercury	7439-97-6	0.20	0.0005	0.0	0	0.05	2.0	0.0	-			
Nickel	7440-02-0	16	13.6	0.1	2	500	100	0.5	1			
Potassium	7440-09-7	NA	NA	12.4	50	NA	NA	0.0	-			
Selenium	7782-49-2	NA	0.210	0.2	2	5.0	40	0.2	5			
Silver	7440-22-4	0.50	2.0	0.1	1	0.12	40	0.0	1			
Sodium	7440-23-5	NA	NA	22.5	100	NA	50000	0.0	-			
Thallium	7440-28-0	NA	1.0	3.1	2	0.24	2.0	0.0	1			
Vanadium	7440-62-2	NA	2.0	0.4	2	12	10	0.1	1			
Zinc	7440-66-6	120	8.5	1.6	2	7400	2000	0.2	1			
Cyanide	57-12-5	0.0001	1.3	N/A	N/A	5.2	100	N/A	N/A			
Low Level Mercury by 1631												
Mercury	7439-97-6	0.20	0.0005	0.0043	N/A	0.05	2.0	TBD for Sub-Lab	TBD for Sub-Lab			

Notes:

Minimum screening levels for the respective medium were derived from the following EBSLs and human health criteria, in the following order

Sediment: NJDEP Ecological Screening Criteria; ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Sediment- Associated Biota (Jones et al. 1997

Soil: NJDEP Ecological Screening Criteria or NJ Soil Remediation Standards; USEPA (2014) Regional Screening Level (RSL) Residential Soil (Cancer Risk = 1x10-6; NonCancer Hazard = 0.1

Surface Water: NJ GWQC (Freshwater Chronic or Human Health Criteria); ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Aquatic Biota (Suter and Tsao 199 Groundwater: NJDEP Groundwater Quality Standards N.J.A.C. 7:9C; USEPA (2014) Regional Screening Level (RSL) Tapwater (Cancer Risk = 1x10-6; NonCancer; Hazard = 0.1)

Additional screening levels may be included based on site characterization information.

μg/L - microgram per liter

EBSL - Ecologically-Based Screening Level

MDL - Method Detection Limit mg/kg - milligram per kilogram

NA - Not Available

NJ GWQC - New Jersey Groundwater Quality Criteria

NJDEP - New Jersey Department of Environmental Protection

ORNL - Oak Ridge National Laboratory

RL - Reporting Limit TAL - Target Analyte List



QAPP WORKSHEET # 15d - PCBs

Project Action Limits and Laboratory-Specific Detection/Quantitation Limits (UFP-QAPP Manual Section 2.6.2.3 and Figure 15)

(EPA 2106-G-05 Section 2.2.6)

			S	oil/Sediment		Water						
						Surface						
		Sediment	Soil PAL	Method MDL	Method RL (LOW)	Water PAL	Groundwater	Method MDL	Method RL			
Analyte (method)	CAS Number	PAL (mg/kg)	(mg/kg)	(LOW) (mg/kg)	(mg/kg)	(μg/L)	PAL (μg/L)	(μg/L)	(μg/L)			
TCL - PCB/Pesticides (SOM	01.2)											
2,4'-DDD	53-19-0	NA	NA	NA	0.0033	NA	NA	N/A	N/A			
2,4'-DDE	3424-82-6	NA	NA	NA	0.0033	NA	NA	N/A	N/A			
2,4'-DDT	789-02-6	NA	NA	NA	0.0033	NA	NA	N/A	N/A			
4,4'-DDD	72-54-8	0.008	0.758	NA	0.0033	0.0003	0.10	0005	N/A			
4,4'-DDE	72-55-9	0.005	0.596	NA	0.0033	0.0002	0.10	0.003	N/A			
4,4'-DDT	50-29-3	0.008	0.004	NA	0.0033	0.0002	0.10	0.004	N/A			
Aldrin	309-00-2	0.002	0.003	NA	0.0017	0.0000	0.04	0.001	N/A			
alpha-BHC	319-84-6	0.006	0.099	NA	0.0017	0.0026	0.02	N/A	N/A			
alpha-Chlordane	5103-71-9	NA	NA	NA	0.0017	NA	NA	0.002	N/A			
beta-BHC	319-85-7	0.005	0.004	NA	0.0017	0.009	0.04	0.002	N/A			
delta-BHC	319-86-8	NA	NA	NA	0.0017	2.20	NA	0.002	N/A			
Dieldrin	60-57-1	0.002	0.002	NA	0.0033	0.00005	0.03	0.004	N/A			
Endosulfan I	959-8-8	0.006	470	NA	0.0017	0.05	40	N/A	N/A			
Endosulfan II	33213-65-9	0.006	470	NA	0.0033	0.05	40	N/A	N/A			
Endosulfan sulfate	1031-07-8	0.035	0.036	NA	0.0033	2.22	40	0.002	N/A			
Endrin	72-20-8	0.003	0.010	NA	0.0033	0.036	2.0	0.004	N/A			
Endrin aldehyde	7421-93-4	0.480	0.011	NA	0.0033	0.059	NA	0.006	N/A			
Endrin ketone	53494-70-5	NA	NA	NA	0.0033	NA	NA	0.004	N/A			
gamma-BHC (Lindane)	58-89-9	0.003	0.005	NA	0.0017	0.026	0.03	0.001	N/A			
gamma-Chlordane	5103-74-2	NA	NA	NA	0.0017	NA	NA	0.001	N/A			
Heptachlor	76-44-8	0.001	0.006	NA	0.0017	0.000079	0.05	0.001	N/A			
Heptachlor epoxide	1024-57-3	0.005	0.070	NA	0.0017	0.000039	0.20	0.005	N/A			
Methoxychlor	72-43-5	0.014	0.020	NA	0.0170	0.03	40	0.032	N/A			
Toxaphene	8001-35-2	0.000	0.119	NA	0.0330	0.0002	2.0	0.049	N/A			
Aroclor-1016	12674-11-2	0.007	0.400	NA	0.0330	NA	0.14	0	N/A			
Aroclor-1221	11104-28-2	0.120	0.150	NA	0.0330	0.28	0.0046	0	N/A			
Aroclor-1232	11141-16-5	0.600	0.150	NA	0.0330	0.58	0.0046	0	N/A			
Aroclor-1242	53469-21-9	0.170	0.240	NA	0.0330	0.053	0.039	0.02	N/A			
Aroclor-1248	12672-29-6	0.030	0.240	NA	0.0330	0.081	0.039	0	N/A			
Aroclor-1254	11097-69-1	0.060	0.110	NA	0.0330	0.033	0.039	0.014	N/A			
Aroclor-1260	11096-82-5	0.005	0.240	NA	0.0330	94	0.039	0	N/A			
Aroclor-1262	37324-23-5	NA	NA	NA	0.0330	NA	NA	0	N/A			
Aroclor-1268	11100-14-4	NA	NA	NA	0.030	NA	NA	0	N/A			

Notes:

Minimum screening levels for the respective medium were derived from the following EBSLs and human health criteria, in the following order:

Sediment: NJDEP Ecological Screening Criteria; ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Sediment- Associated Biota (Jones et al. 1997).

Soil: NJDEP Ecological Screening Criteria or NJ Soil Remediation Standards; USEPA (2014) Regional Screening Level (RSL) Residential Soil (Cancer Risk = 1x10-6; NonCancer Hazard = 0.1).

Surface Water: NJ GWQC (Freshwater Chronic or Human Health Criteria); ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Aquatic Biota (Suter and Tsao 199 Groundwater: NJDEP Groundwater Quality Standards N.J.A.C. 7:9C; USEPA (2014) Regional Screening Level (RSL) Tapwater (Cancer Risk = 1x10-6; NonCancer; Hazard = 0.1).

Additional screening levels may be included based on site characterization information.

 $\mu g/L$ - microgram per liter

EBSL - Ecologically-Based Screening Level

MDL - Method Detection Limit mg/kg - milligram per kilogram

NA - Not Available

NJ GWQC - New Jersey Groundwater Quality Criteria

NJDEP - New Jersey Department of Environmental Protection

ORNL - Oak Ridge National Laboratory

RL - Reporting Limit

TCL - Target Compound List



QAPP WORKSHEET # 15d - PCBs Congeners Project Action Limits and Laboratory-Specific Detection/Quantitation Limits (UFP-QAPP Manual Section 2.6.2.3 and Figure 15)

(EPA 2106-G-05 Section 2.2.6)

					Soil/Sediment	Water					
						Achievable	Achievable				
		Sediment				Subcontract	Subcontract	Surface			
		PAL	Soil PAL	Method MDL	Method RL	Laboratory MDL	Laboratory RL	Water PAL	Groundwater	Method MDL	Method RL
Analyte (method)	CAS Number	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	(μg/L)	PAL (μg/L)	(μg/L)	(μg/L)
PCBs (EPA 1668a)**											
2',3,4,4',5-PeCB (PCB-123)	65510-44-3	NA	0.120	See Method	See Method	TBD	TBD	NA	0.02	See Method	0.0005
2,3',4,4',5-PeCB (PCB-118)	31508-00-6	NA	0.120	See Method	See Method	TBD	TBD	NA	0.02	See Method	0.0005
2,3,4,4',5-PeCB (PCB-114)	74472-37-0	NA	0.120	See Method	See Method	TBD	TBD	NA	0.02	See Method	0.0005
2,3,3',4,4'-PeCB (PCB-105)	32598-14-4	NA	0.120	See Method	See Method	TBD	TBD	NA	0.02	See Method	0.0002
3,3',4,4',5-PeCB (PCB-126)	57465-28-8	NA	0.000037	See Method	See Method	TBD	TBD	NA	0.000014	See Method	0.0005
2,3',4,4',5,5'-HxCB (PCB-167)	52663-72-6	NA	0.120	See Method	See Method	TBD	TBD	NA	0.02	See Method	0.0005
2,3,3',4,4',5-HxCB (PCB-156)	38380-08-4	NA	0.120	See Method	See Method	TBD	TBD	NA	0.02	See Method	0.0005
2,3,3',4,4',5'-HxCB (PCB-157)	69782-90-7	NA	0.120	See Method	See Method	TBD	TBD	NA	0.02	See Method	0.0005
3,3',4,4',5,5'-HxCB (PCB-169)	32774-16-6	NA	0.00012	See Method	See Method	TBD	TBD	NA	0.00002	See Method	0.0005
2,3,3',4,4',5,5'-HpCB (PCB-189)	39635-31-9	NA	NA	See Method	See Method	TBD	TBD	NA	0.02	See Method	NA

Notes:

Minimum screening levels for the respective medium were derived from the following EBSLs and human health criteria, in the following order:

Sediment: NJDEP Ecological Screening Criteria; ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Sediment- Associated Biota (Jones et al. 1997).

Soil: NJDEP Ecological Screening Criteria or NJ Soil Remediation Standards; USEPA (2014) Regional Screening Level (RSL) Residential Soil (Cancer Risk = 1x10-6; NonCancer Hazard = 0.1).

Surface Water: NJ GWQC (Freshwater Chronic or Human Health Criteria); ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Aquatic Biota (Suter and Tsao 1996).

Groundwater: NJDEP Groundwater Qualtiy Standards N.J.A.C. 7:9C; USEPA (2014) Regional Screening Level (RSL) Tapwater (Cancer Risk = 1x10-6; NonCancer Hazard = 0.1).

Additional screening levels may be included based on site characterization information.

** Only congeners with a screening value available were presented; however, all 209 congeners will be analyzed and reported.

 $\mu g/L$ - microgram per liter

EBSL - Ecologically-Based Screening Level

MDL - Method Detection Limit

mg/kg - milligram per kilogram

NA - Not Available

NJ GWQC - New Jersey Groundwater Quality Criteria

NJDEP - New Jersey Department of Environmental Protection

ORNL - Oak Ridge National Laboratory

PCB - Polychlorinated Biphenyl

RL - Reporting Limit

TCL - Target Compound List



Rolling Knolls Landfill Superfund Site Data Gap Investigation Oversight

QAPP WORKSHEET # 15e - Dioxin/Furans Project Action Limits and Laboratory-Specific Detection/Quantitation Limits (UFP-QAPP Manual Section 2.6.2.3 and Figure 15)

(EPA 2106-G-05 Section 2.2.6)

				Soil/S	Water								
Analista (mashbad)	CAS Number	Sediment	Soil PAL	Method MDL	Method RL	Achievable Subcontract Laboratory	Achievable Subcontract Laboratory RL	DAL (ma/l)	DOLG (112-(1))	Method	Method	Laboratory	Subcontract Laboratory
Analyte (method)	CAS Number	PAL (mg/kg)	(mg/kg)	(mg/kg)	(mg/kg)	MDL (mg/kg)	(mg/kg)	PAL (μg/L)	PQLG (μg/L)	IVIDL (μg/L)	RL (μg/L)	MDL (μg/L) TBD	RL (μg/L)
Dioxins by 1613		NA	NA	NA	NA	TBD TBD	TBD TBD	NIA	NA	NA	NA	TBD	TBD TBD
Total TCDD Total TCDF		NA NA	NA NA	NA NA	NA NA	TBD	TBD	NA NA	NA NA	NA NA	NA NA	TBD	TBD
Total PeCDD		NA NA	NA NA	NA NA	NA	TBD	TBD	NA NA	NA NA	NA	NA	TBD	TBD
Total PeCDF		NA	NA	NA	NA	TBD	TBD	NA NA	NA	NA	NA	TBD	TBD
Total HxCDD		NA	NA	NA	NA	TBD	TBD	NA	NA	NA	NA	TBD	TBD
Total HxCDF		NA	NA	NA	NA	TBD	TBD	NA NA	NA	NA	NA	TBD	TBD
Total HpCDD		NA	NA	NA	NA	TBD	TBD	NA NA	NA	NA	NA	TBD	TBD
Total HpCDF		NA	NA	NA	NA	TBD	TBD	NA	NA 2 22221	NA	NA	TBD	TBD
2,3,7,8-TCDD	1746-01-6	0.0000012	0.0000002	NA	1	TBD	TBD	0.000000003	0.00001	NA	0.00001	TBD	TBD
1,2,3,7,8,9-HxCDD	19408-74-3	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
OCDD	3268-87-9	NA	NA	NA	10	TBD	TBD	NA	NA	NA	0.000001	TBD	TBD
1,2,3,4,6,7,8-HpCDD	35822-46-9	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
OCDF	39001-02-0	NA	NA	NA	10	TBD	TBD	NA	NA	NA	0.000001	TBD	TBD
1,2,3,4,7,8-HxCDD	39227-28-6	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
1,2,3,7,8-PeCDD	40321-76-4	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
2,3,7,8-TCDF	51207-31-9	NA	NA	NA	1	TBD	TBD	NA	NA	NA	0.00001	TBD	TBD
1,2,3,4,7,8,9-HpCDF	55673-89-7	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
2,3,4,7,8-PeCDF	57117-31-4	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
1,2,3,7,8-PeCDF	57117-41-6	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
1,2,3,6,7,8-HxCDF	57117-44-9	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
1,2,3,6,7,8-HxCDD	57653-85-7	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
2,3,4,6,7,8-HxCDF	60851-34-5	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
1,2,3,4,6,7,8-HpCDF	67562-39-4	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
1,2,3,4,7,8-HxCDF	70648-26-9	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD
1,2,3,7,8,9-HxCDF	72918-21-9	NA	NA	NA	5	TBD	TBD	NA	NA	NA	0.00005	TBD	TBD

Notes:

Minimum screening levels for the respective medium were derived from the following EBSLs and human health criteria, in the following order:

Sediment: NJDEP Ecological Screening Criteria; ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Sediment- Associated Biota (Jones et al. 1997).

Soil: NJDEP Ecological Screening Criteria or NJ Soil Remediation Standards; USEPA (2014) Regional Screening Level (RSL) Residential Soil (Cancer Risk = 1x10-6; NonCancer Hazard = 0.1).

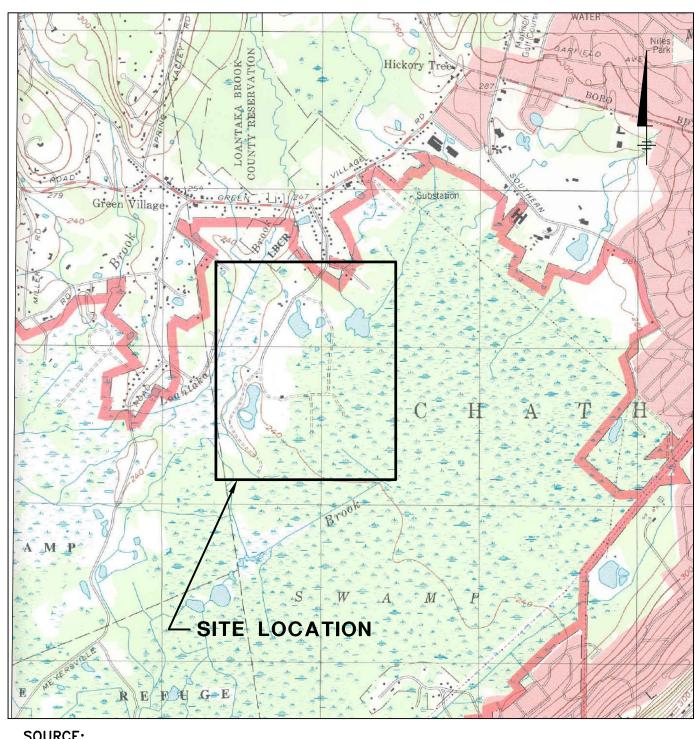
Surface Water: NJ GWQC (Freshwater Chronic or Human Health Criteria); ORNL Toxicological Benchmarks for Screening Potential Contaminants of Concern for Effects on Aquatic Biota (Suter and Tsao 1996).

Groundwater: NJDEP Groundwater Qualtiy Standards N.J.A.C. 7:9C; USEPA (2014) Regional Screening Level (RSL) Tapwater (Cancer Risk = 1x10-6; NonCancer; Hazard = 0.1).

Additional screening levels may be included based on site characterization information.

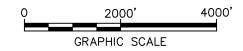


Rolling Knolls Landfill Superfund Site Data Gap Investigation Oversight



SOURCE:

CHATHAM QUADRANGLE, NJ 7.5 MINUTE SERIES CONTOUR INTERVAL 20 FEET





ROLLING KNOLLS LANDFILL SUPERFUND SITE CHATHAM, NEW JERSEY DATA GAPS SAMPLING AND ANALYSIS PLAN

SITE LOCATION



FIGURE

1

BY: MEYER, JULIE

PLOTTED: 9/16/2014 11:41 AM

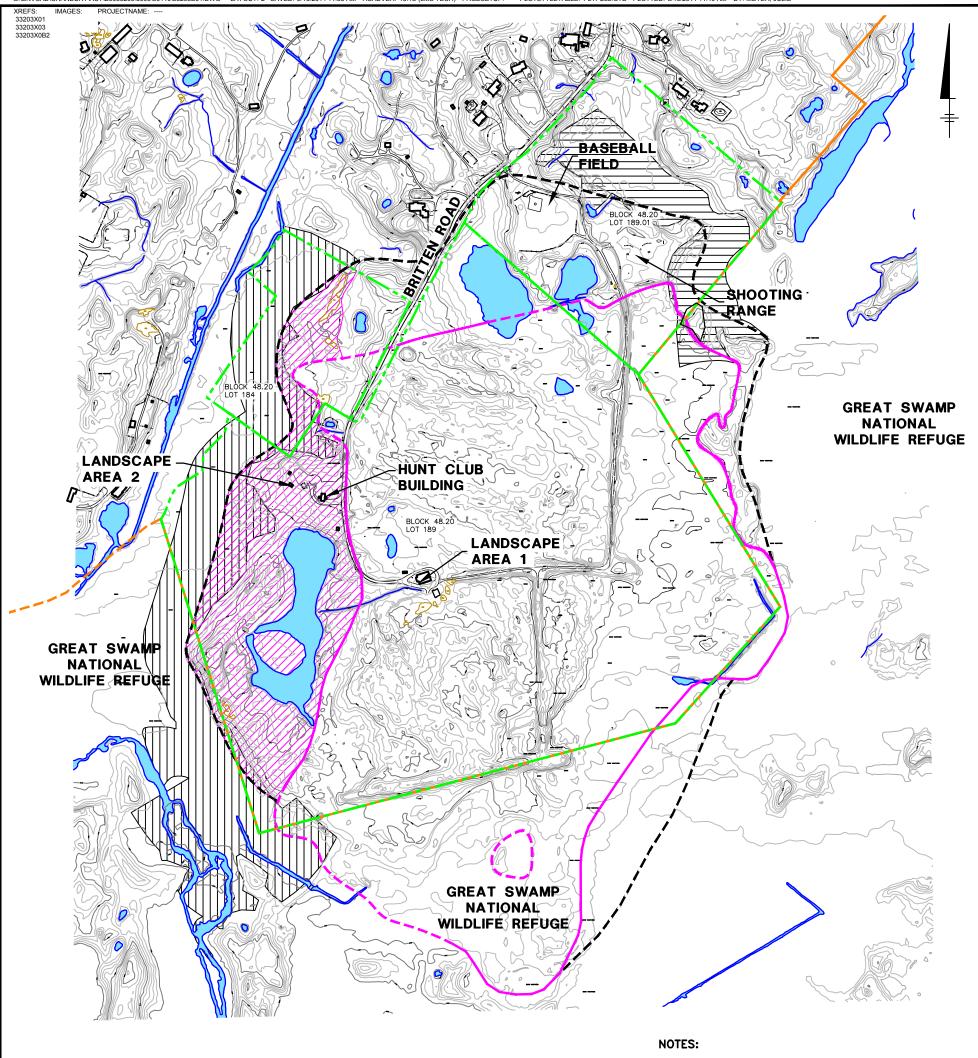
PLOTSTYLETABLE: PLTFULL.CTB

LYR: ON=";OFF=*REF* 'ER: 18.1S (LMS TECH)

PM: K.ROMAINE SAVED: 9/16/2014 1

PIC:

_: G M



LEGEND:

OPEN WATER

PRE-REMEDIAL INVESTIGATION PROJECTED EDGE OF LANDFILLED MATERIALS

EDGE OF LANDFILLED WASTES OBSERVED DURING TEST PIT ACTIVITIES (DASHED WHERE APPROXIMATE)

GREAT SWAMP NATIONAL WILDLIFE REFUGE PROPERTY BOUNDARY (DASHED WHERE APPROXIMATE)

TAX PARCELS

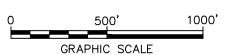
WASTE AND DEBRIS OBSERVED ON GROUND SURFACE BUT NOT OBSERVED OR ANTICIPATED TO BE BELOW GROUND SURFACE

POTENTIAL BOG TURTLE HABITAT AREA A (35.31 ACRES)

POTENTIAL BOG TURTLE HABITAT AREA B (10.89 ACRES)

SOURCES:

- 1. BASEMAP FROM JAMES M. STEWART INC., LAND SURVEYORS, PHILADELPHIA, PA., (ELECTRONIC FILE: 292406.DWG DATED: 6/30/06)
- 2. TAX PARCEL DATA FOR CHATHAM TOWNSHIP WAS PROVIDED BY CIVIL SOLUTIONS.



- THE PRE-REMEDIAL INVESTIGATION PROJECTED EDGE OF THE PRE-REMEDIAL INVESTIGATION PROJECTED EDGE OF LANDFILLED MATERIALS ON THIS FIGURE IS APPROXIMATE AS DRAWN AND IS BASED ON VISUAL OBSERVATIONS OF THE GROUND SURFACE MADE DURING SITE VISITS CONDUCTED JUNE 20, 2006 THROUGH JULY 14, 2006.
- THE EDGE OF LANDFILLED WASTES OBSERVED DURING TEST PIT ACTIVITIES IS DRAWN BASED ON OBSERVATIONS OF MATERIALS EXCAVATED DURING TEST PIT ACTIVITIES CONDUCTED FROM JULY 26, 2007 TO SEPTEMBER 6, 2007 AND MAPCH 26 2008 AND MARCH 26, 2008.
- THE PORTION OF THE GREAT SWAMP NATIONAL WILDLIFE THE PORTION OF THE GREAT SWAMP NATIONAL WILDLIFE REFUGE (GSNWR) PROPERTY BOUNDARY ON THIS FIGURE WITHIN CHATHAM TOWNSHIP, NJ WAS OBTAINED FROM CHATHAM TOWNSHIP TAX PARCEL DATA PROVIDED BY CIVIL SOLUTIONS. THE PORTION OF THE GSNWR PROPERTY BOUNDARY ON THIS FIGURE OUTSIDE OF CHATHAM TOWNSHIP IS APPROXIMATE AND WAS OBTAINED FROM THE UNITED STATES FISH AND WILDLIFE SERVICE (GEOGRAPHIC INFORMATION SYSTEMS AND SPATIAL DATA).
- BLOCK 48.20, LOTS 184 AND 189 ARE OWNED BY ROBERT J. MIELE AS TRUSTEE FOR THE TRUST CREATED BY THE LAST WILL AND TESTAMENT OF ANGELO J. MIELE. BLOCK 48.20, LOT 189.01 IS OWNED BY THE GREEN VILLAGE FIRE DEPARTMENT.

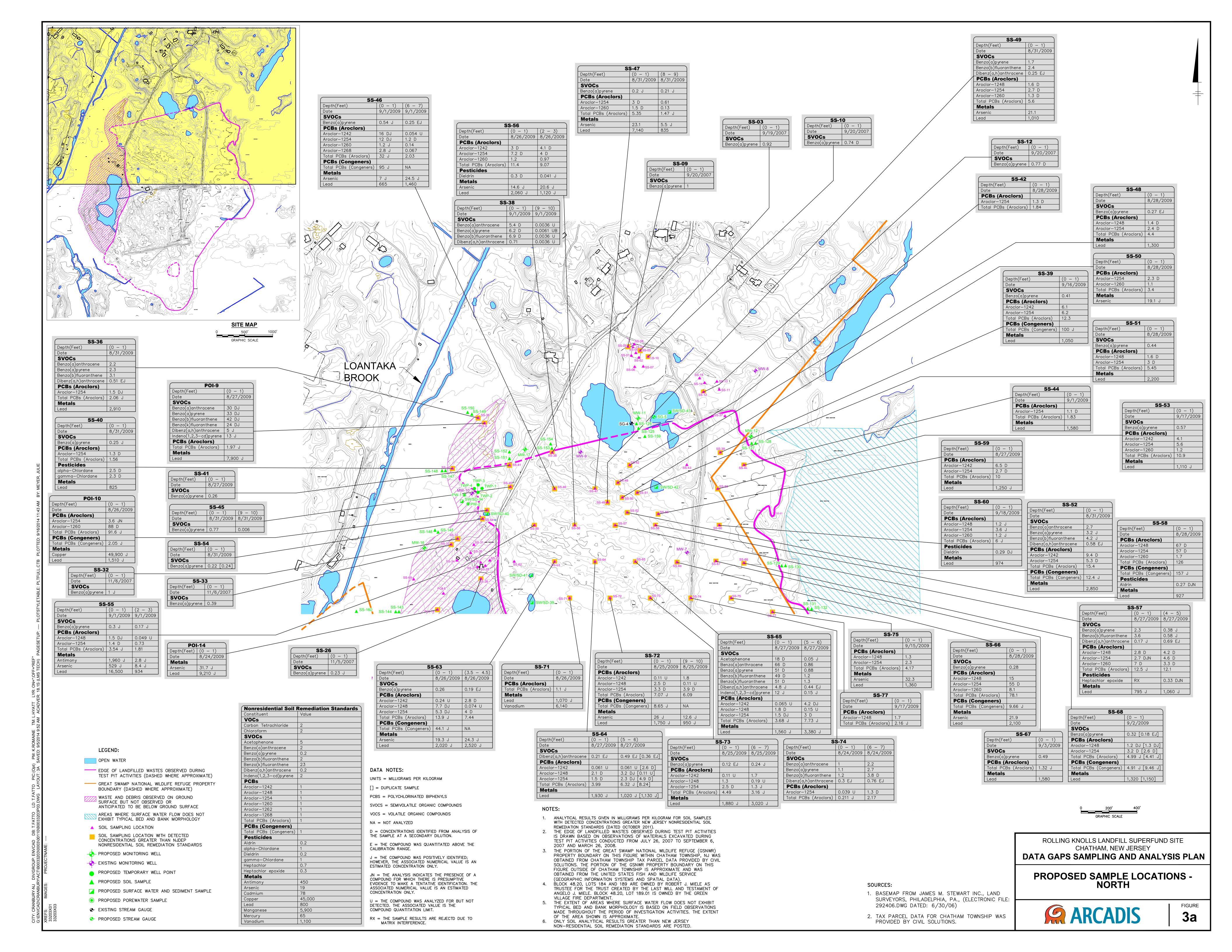
ROLLING KNOLLS LANDFILL SUPERFUND SITE CHATHAM, NEW JERSEY

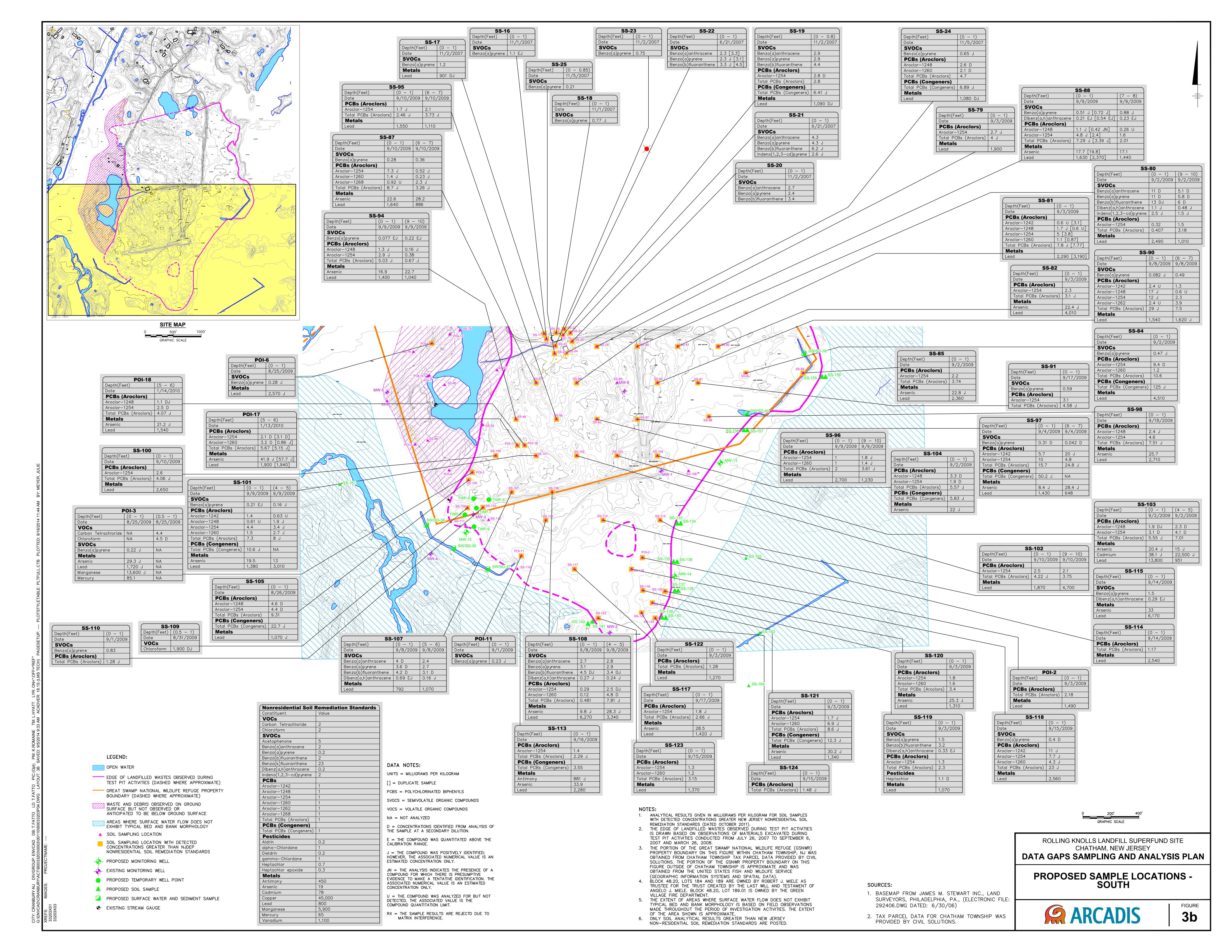
DATA GAPS SAMPLING AND ANALYSIS PLAN

SITE PLAN



FIGURE 2







ARCADIS U.S., Inc. 8 South River Road Cranbury New Jersey 08512 Tel 609 860 0590 Fax 609 860 0491

MEMO

Tanva Mitchell United States Environmental Protection Agency

Conies: John Persico

Suzy Walls

October 15, 2014

ARCADIS Project No.: B0033203.0004

Subject:

USEPA Comments dated October 9, 2014 on the Data Gaps Sampling and Analysis Plan and Quality Assurance Project Plan Rolling Knolls Landfill Superfund Site, Chatham, New Jersey

This memorandum is a summary of the conference call on October 15, 2014 to discuss USEPA's comments (dated October 9, 2014) on the Data Gaps Sampling and Analysis Plan (SAP) and Quality Assurance Project Plan (QAPP) for the Rolling Knolls Landfill Superfund Site in Chatham, New Jersey. The call was attended by:

- Tanya Mitchell, United States Environmental Protection Agency (USEPA);
- Michael Sivak, USEPA;
- Michael Clemetson, USEPA;
- Juan Fajardo, USEPA
- Paul Hagerman (CDM Smith);
- Joe Button (CDM Smith);
- Richard Ricci, Lowenstein Sandler LLP;
- Mickey Faigen, Issues, LLC;
- Andrew Gutherz, ARCADIS;
- John Persico, ARCADIS; and
- Suzy Walls, ARCADIS.

During the call, John Persico (ARCADIS) lead the discussion of general and specific comments from the USEPA's comment letter as discussed below. USEPA also reiterated the need to have USEPA counsel present if the Settling Parties' counsel would be present.

ARCADIS

General Comment 4: ARCADIS asked for clarification on which data USEPA would like to have in the Electronic Data Deliverable (EDD) format. USEPA clarified that beginning now, with the data gaps sampling, and moving forward, all data collected for the site must be submitted to the USEPA in the EDD format. Data previously collected for the Site Characterization Summary Report would not be submitted as part of this request. ARCADIS agreed to submit these deliverables moving forward.

Specific Comment 2: ARCADIS asked for clarification on the inclusion of the reference to New Jersey Department of Environmental Protection (NJDEP) Technical Requirements for Site Remediation, given that these have not been required by USEPA and have not been used during the site evaluations thus far. USEPA stated that this comment was only in regards to the current phase of sampling and would not open the door to applying these guidelines to previous phases of investigation at the site. ARCADIS asked if the comment could be reworded to include a specific request, such as needing vertical delineation in site boundary samples. USEPA stated that in general NJDEP regulations would need to be met, but that ARCADIS could submit a proposed response to the comment that USEPA would review and that they would also confer with NJDEP.

Specific Comment 5: ARCADIS requested that "approximately 200-acre" be removed from the revised site description due to the inconsistency in the paragraph when the landfill was later referred to as "approximately 170 acres." USEPA instead suggested that the paragraph be revised to remove the latter size reference of 170 acres. ARCADIS agreed to revise accordingly.

Specific Comment 9: ARCADIS reviewed their understanding of the permit equivalency process, which included revisiting areas of the site where work is currently proposed and filling out the proper permit equivalency forms. These forms were sent to the property owners for their review and signature and will be sent to NJDEP; however, NJDEP would not need to issue permits prior to the start of work. USEPA agreed that this process was also their understanding of the permit equivalency process but that this comment was sent from the NJDEP. USEPA agreed to contact NJDEP for clarification of this request and instructed ARCADIS to respond with the proposed approach stated above.

Specific Comment 10: ARCADIS asked for clarification on the request for vertical delineation of the proposed surface soil samples, given that the current samples were proposed for use in the risk assessments which only require surface evaluations for the identified receptors. Contrary to that objective, NJDEP guidance requires delineation to any depth necessary. USEPA requested a written explanation to the response for why vertical delineation was not necessary for this sampling.

ARCADIS also asked for clarification on the request for full Target Compound List (TCL)/Target Analyte List (TAL) parameters for all surface soil samples proposed. ARCADIS explained that the analyses chosen for each sample were based on historical sample results in each of those areas and that step-out samples would not require the full TCL/TAL list. USEPA acknowledged the reasoning but felt that full TCL/TAL parameters were required given the inconsistent occurrence of constituents in surface soil at the

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site. USEPA did not agree that reducing the list of analytes based on previous sample results was appropriate at this time.

Specific Comment 15: ARCADIS asked for clarification on the request for full TCL/TAL parameters for all temporary wells, given that these temporary wells were in very close proximity to one another and were being installed specifically to evaluate volatile organic compound (VOC) and metal concerns in MW-3 and MW-10. Further, ARCADIS stated that these wells were intended to be used for screening, were not meant for delineation and that permanent wells would be placed in these areas for future monitoring if USEPA felt TCL/TAL parameters were necessary in these areas. USEPA acknowledged the reasoning but felt that full TCL/TAL parameters were required given the extended time since the last round of groundwater data and the potential for constituents other than VOCs and metals to have migrated into groundwater. USEPA did not agree that targeting the list of analytes was appropriate at this time.

Specific Comment 16: ARCADIS asked for clarification on the request for full TCL/TAL parameters for the proposed porewater samples, given that the samples were only being collected to evaluate VOC and metal concerns in MW-3 and MW-10, and that these samples were not meant for delineation purposes. ARCADIS also raised the concern of finding suitable membranes for all of the TCL/TAL parameters, specifically polychlorinated biphenyls (PCBs), pesticides, and semi-volatile organic compounds (SVOCs), and for collecting enough volume to accommodate all of the analyses. USEPA did not agree that targeting the list of analytes was appropriate at this time; however, they acknowledged the concerns of finding a suitable membrane for the passive sampling and asked that ARCADIS look into the possible membranes. In the event an appropriate membrane could be found, USEPA requested collection of porewater in the following sequence: VOCs, PCBs, pesticides, SVOCs, and metals. If a suitable membrane cannot be identified, USEPA acknowledged that the analyses may be limited.

Specific Comment 20: ARCADIS asked for clarification on USEPA's proposed schedule change from 30 days between rounds of groundwater sampling to potentially 90 days or more to capture wet season and dry season conditions. While ARCADIS was not opposed to this change, we believe the extra time would not significantly change the outcome of the sampling but would add an additional delay in the schedule. USEPA did not believe that extending the time between sampling would drastically alter the overall schedule given that groundwater sampling was unlikely to delay biota sampling, which would also be occurring at the site.

ARCADIS also asked for clarification on the request for a second round of complete sampling from the existing monitoring wells on-site. USEPA explained that without sufficient groundwater data, monitored natural attenuation (MNA) could not be considered as a possible remedy during the feasibility study. ARCADIS asked if this second round of sampling would also require the full TCL/TAL parameters and USEPA confirmed that the full suite of constituents would be required. USEPA suggested that the Settling Parties review the requirements needed for each potential alternative that they would consider during the feasibility study. ARCADIS asked where those requirements were listed and USEPA said they could be

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found online, along with the New Jersey requirements for MNA remedies. ARCADIS stated that previous New Jersey projects required 8 rounds of groundwater monitoring prior to MNA remedies; however, USEPA said that in some cases fewer round of monitoring may be required if adequate trends or patterns could be demonstrated. USEPA stated that MNA closures were receiving higher scrutiny at USEPA at this time and reiterated that adequate rounds groundwater data would be needed to consider MNA as a potential remedy. ARCADIS asked if other data would also be required, including specific geochemistry data. USEPA stated that geochemistry data, along with a number of other types of data and trends, would be required.

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